Encyclopedia of Food Mycotoxins

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With 96 Figures and 9 Tables



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My Children Anna and Vincent

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Preface

The main emphasis of the present book is listing all foods which have been reported to be contaminated with mycotoxins (degree of contamination, concentration, country of origin/detection). To find out quickly whether a foodstuff is contaminated by a specific mycotoxin, the contaminated foods have been listed alphabetically under "Natural Occurrence" of a mycotoxin.

Products are listed by the country in which they were investigated for mycotoxin contamination. In some cases, the country of detection is **not** necessarily the country of origin, but information was lacking concerning the country of origin of such imports in the original literature. If only "imported" occurs after the country of investigation no more data were available in the original literature. Sometimes, e.g., in the case of nuts or spices, the original literature neither contained the producing country nor the addition "imported". In these cases also no indications were given in the Encyclopedia. However, in all cases where the origin of the investigated food was known, the name of the producing country was given.

The multiple listing of some countries in connection with mycotoxin contamination of food should not implicate a high rate of mycotoxin contamination of foods in these countries but primarily documents the efforts being made to detect toxic fungal metabolites in food.

The special data concerning the mycotoxin contamination of food, e.g. 6/12, means six positive (contaminated) products from a total of twelve. Means represent the mean of positively contaminated samples, except where indicated otherwise. An entry of < x generally refers to the limit of detection. Values above this level are included in calculating the mean of all positive samples.

The data concerning mycotoxin contamination of food listed in the Encyclopedia based on results predominantly published in recommended journals and scientific books in this field (mainly the literature given at the end of the book). In the case of commonly isolated mycotoxins, e.g. aflatoxins, trichothecenes, it was not possible to consider all the results published.

In the literature, sometimes contradictory information about the mycotoxin spectrum of mold species can be found. Therefore, only the "safe" and food relevant mycotoxins of a species and not all known toxic metabolites were listed. This information mainly based on Frisvad J (1988) Fungal species and their specific production of mycotoxins. In: Samson RA, Reenen-Hoekstra ES (Eds) Introduction of Food-borne Fungi, pp 239–249. Centralbureau voor Schimmelcultures, Baarn (*Aspergillus* and *Penicillium* species), Marasas WFO, Nelson PE, Tousson TA (1984) Toxigenic Fusarium Species, Identity and Mycotoxicology. The Pennsylvania State University Press, University Park, PA (*Fusarium* species), Samson RA, Hoekstra ES, Frisvad JC, Filtenborg O (1998) Introduction to Food-borne Fungi. Centraalbureau voor Schimmelcultures, Baarn (*Aspergillus* species and others). According to Ainsworth & Bisby's "Dictionary of the Fungi" all mycotoxigenic fungi listed in the Encyclopedia may be grouped easily to their corresponding family, order, phylum and kingdom.

The names used for all *Penicillium* species based on Pitt JI (1979) The Genus *Penicillium* and its Teleomorphic States *Eupenicillium* and *Talaromyces*, Academic Press, London.

Although in some cases more fungal species are known to produce a mycotoxin usually only the names of food relevant molds like *Aspergillus* spp., *Penicillium* spp. and/or *Fusarium* spp. are given.

Since in some cases various toxicological data of mycotoxins do exist for better comparison only the data of the per oral application in rats/mice (as far as possible) were chosen.

Gießen, Summer 2000

Martin Weidenbörner

Abbreviations

- BGY Bright greenish yellow (fluorescence)
- bm body mass
- bw body weight
- conc concentration
- d day(s)
- EC Esophageal cancer
- ELISA Enzyme linked immunosorbent assay
- EU European Union
- FAO Food and Agricultural Organization of the United Nations World Health Organization
- FDA United States Food and Drug Administration
- GC Gas chromatography
- GC-MS Gas chromatography-mass spectrometry h hour(s)
- h hour(s)
- HPLC High performance liquid chromatography
- HTST High temperature short time
- IARC International Agency for Research on Cancer
- ip intraperitoneal
- iv intravenous
- JECFA Joint Expert Committee on Food Additives
- kGy kilo Gray
- LD₅₀ Lethal dosis of e.g. aflatoxin that will cause acute toxicity in 50 % of the target population
- mc moisture content
- min minutes
- mp melting point
- mw molecular weight
- nc no comment (not stated, unclear)
- ND Not detected
- NOAEL No observed adverse effect level
- NMR Nuclear magnetic resonance
- po per os
- PTWI Provisional tolerable weekly intake
- sa sample(s)
- sc subcutaneous
- sqd semi-quantitative determination

TLC	Thin-layer chromatography
tr	traces
UAE	United Arabic Emirates
WHO	World Health Organization of the United Nations
kg	kilogram
mg	milligram = 10^{-3} g;
μg	1 mg/kg = 1:10 ⁶ = ppm = parts per million microgram = 10 ⁻⁶ g; 1 μ g/kg = 1:10 ⁹ = ppb = parts per billion
l ml µl	litre millilitre = 10^{-3} l; 1 ml/l = 1:10 ³ microlitre = 10^{-3} ml; 1 µl/l = 1:10 ⁶ = ppm = parts per million

XII

A

AAL-toxins is the abbreviation for *Alternaria alternata* f. sp. *lycopersici* toxins which possess a "sphingosine-like" structure (see Figure AAL-toxins). AAL-toxins include the two fractions T_A and T_B . T_A $(C_{13}H_{53}NO_{15}, MW = 679)$ consists of two esters (C_{13} or C_{14}) of 1,2,3-propane-tricarboxyclic acid and 1-amino-11,15dimethylheptadeca-2,4,5,13,14-pentol. T_B $(C_{13}H_{53}NO_{13}, MW = 647)$ consists of two esters (C_{13} or C_{14}) of 1,2,3-propane-tricarboxyclic acid and 1-amino-11,15dimethylheptadeca-2,4,13,14-tetrol. These

fractions contain four closely related compounds T_A -1, T_A -2, T_B -1 and T_B -2. Recently they were renamed alperisins A1, A2, B1, and B2. The alperisins are remarkably similar to the \rightarrow fumonisins.

CHEMICAL DATA

Empirical formula: $C_{13}H_{53}NO_{15}$, molecular weight: 679 (T_A) Empirical formula: $C_{13}H_{53}NO_{14}$, molecu-

lar weight: 663 (T_B) FUNGAL SOURCES

Alternaria alternata f. sp. lycopersici

NATURAL OCCURRENCE

There are no reports on the natural occurrence of these toxins in plant products, probably because *A. alternata* f. sp. *lycopersici* is a rarely occurring pathotype of *A. alternata*. However, AAL-toxins and fumonisins (FB₁, FB₂, FB₃) occur together in spores and mycelia of *A. alternata*.

Тохісіту

Like fumonisin B_1 the AAL-toxins caused stem cancer disease in "Earlypark-7" and other susceptible tomato cultivars. In addition, AAL-toxins and the fumonisins inhibited ceramide synthase in animal cells, cell prolifeartion in rat liver and dog kidney cells.

Acacia concinna (medicinal seeds) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: nc/nc, conc. range: 80-1130 μ g/kg, country: India \rightarrow citrinin incidence: nc/nc, conc. range: 10-760 μ g/kg, country: India

Acetoxyscirpenediol 4- or \rightarrow 15-acetylscirpentriol

3-Acetyldeoxynivalenol (Syn.: deoxynivalenol monoacetate) is a 3α -acetoxy- 7α ,15-trihydroxy-12,13-epoxytrichothec-9-en-8-one and belongs to the \rightarrow trichothecenes (\rightarrow mycotoxins) (see Figure 3-Acetyldeoxynivalenol).

	H3	CH_3 R_3 CH_3 R_2	R ₁ NH ₂ OH OH
AAL-TOXIN	R ₁	R ₂	R ₃
T _A -1	ОН	ОН	–O ₂ C–CH ₂ –CH(CO ₂ H)–CH ₂ –CO ₂ H
T _A -2	ОН	$-O_2C-CH_2-CH(CO_2H)-CH_2-CO_2H$	ОН
Т _в —1	н	ОН	–O ₂ C–CH ₂ –CH(CO ₂ H)–CH ₂ –CO ₂ H
Т _в –2	Н	$-O_2C-CH_2-CH(CO_2H)-CH_2-CO_2H$	ОН

AAL-toxins. Alternaria alternata f. sp. lycopersici (AAL) toxins

M. Weidenbörner, *Encyclopedia of Food Mycotoxins* © Springer-Verlag Berlin Heidelberg 2001

3-Acetyldeoxynivalenol

CHEMICAL DATA Empirical formula: $C_{17}H_{22}O_7$, molecular weight: 338

FUNGAL SOURCES \rightarrow Fusarium culmorum (W.G. Smith) Sacc., \rightarrow Fusarium graminearum Schwabe

NATURAL OCCURRENCE \rightarrow barley, \rightarrow maize, \rightarrow oats, \rightarrow rye, \rightarrow triticale, \rightarrow wheat

TOXICITY feed refusal (rats) LD₅₀ (ip): 49.4-49.9 mg/kg bw mice (ddS strain)

DETECTION ELISA, TLC, GC-MS, MS

Further Comments

Most Japanese strains of *F. graminearum* produced 3-acetyldeoxynivalenol. The same is true for Chinese strains although the 15-acetatedeoxynivalenol could be isolated from Chinese grain. \rightarrow deoxynivalenol

15-Acetyldeoxynivalenol belongs to the \rightarrow trichothecenes (\rightarrow mycotoxins) (see Figure 15-Acetyldeoxynivalenol).

CHEMICAL DATA Empirical formula: $C_{17}H_{22}O_7$, molecular weight: 338

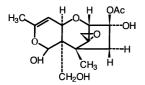
Fungal Sources

 \rightarrow Fusarium graminearum Schwabe

NATURAL OCCURRENCE \rightarrow maize, \rightarrow wheat

TOXICITY

In combination with \rightarrow deoxynivalenol and \rightarrow zearalenone the aforementioned



3-Acetyldeoxynivalenol

contaminated samples caused feed refusal in swine.

Detection GC-MS

Further Comments

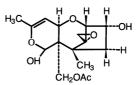
Nearly all strains of *F. graminearum* isolated in North America are able to produce this mycotoxin. This trichothecene mycotoxin occurred in naturally infected field maize samples (ca. 16,300 and 1510 μ g/kg) used for feed. 15-acetyldeoxinvalenol co-occurs with \rightarrow deoxynivalenol and \rightarrow zearalenone.

4-Acetylnivalenol → fusarenon X

4-Acetylscirpentriol (Syn.: 15-acetylscirpentriol)

Acute aflatoxicosis → aflatoxicosis

Acute cardiac beriberi (Syn.: Shoshinkakke) A probable \rightarrow mycotoxicosis which belongs to the complex of "yellow rice diseases" (\rightarrow yellow rice disease). It was first described in Japan at the end of the last century. The disease has mainly been reported from Asian countries where \rightarrow rice is a staple food and has been recognized for the past three centuries. The mold damaged rice is mainly contaminated with \rightarrow Penicillium citreonigrum Dierckx (synonyms P. citreoviride, P. *toxicarium*). \rightarrow Citreoviridin the most important mycotoxin (neurotoxin) of this mold which causes a very rapid \rightarrow paralysis of the respiratory muscles. In combination with \rightarrow convulsion, vomition, ascending \rightarrow paralysis, and lowering



15-Acetyldeoxynivalenol

of the body temperature, the patient usually dies within a short period of 1-3 days, once the disease started. There is no method available of saving the patient from acute cardiac beriberi. Because moldy "yellow rice" was thought to be responsible for this disease the sale of this rice was prohibited in Japan in 1910. Subsequently no more cases of acute cardiac beriberi have been reported. The disease is now of only historical interest in Japan. However, in other parts of Asia P. citreonigrum and its mycotoxin citreoviridin which is also produced by P. ochrosalmoneum may still contribute acute cardiac beriberi.

It is under discussion whether there are several types of beriberi (e.g. atropic and wet beriberi) having the same etiological origin. The difference in symptoms compared to acute cardiac beriberi may be due to dose and duration of intake of the mycotoxin. In these cases severe \rightarrow paretic signs were not observed. In contrast to acute cardiac beriberi, the cause of beriberi is a nutritional disease, an avitaminosis (vitamin B). This is proved by the following facts: the slower course of the disease, no dilation of the right ventricle, and no hypertrophy of adrenal medulla. In addition, administration of liver removed from typical shoshin-kakke patients led to the recovery of vitamin B1-deficient animals. This indicates that adequate amounts of vitamin B_1 were present in the liver of these patients at the time of death. However, to prove beyond doubt that citreoviridin is the cause of acute cardiac beriberi, the etiology of the chemical pathway of this neurotoxin has to be clarified.

Aflatoxicol (Abbr.: AFL, AFR₀) AFL was first reported in microorganisms (\rightarrow mycotoxins) and is the cyclopentanol derivative (2,3,6a,9a-tetrahydro-1-

hydroxy-4-methoxy-cyclopenta[c]furo[3',2':4,5]furo[2,3-h][1]-benzopyran-11(1H)-one) of \rightarrow aflatoxin B₁ (see Figure Aflatoxicol).

CHEMICAL DATA

Empirical formula: $C_{17}H_{13}O_6$, molecular weight: 313

NATURAL OCCURRENCE \rightarrow human breast milk, \rightarrow pistachio nuts

Тохісіту

AFL resulted from the in vitro and in vivo metabolism of AFB₁ by soluble NADPH-dependent reductases of submitochondrial liver fractions from humans and several animal species (e.g. poultry, rabbits, trouts). A microsomal AFL-dehydrogenase catalyzes the enzymatically reversible reaction. AFL therefore may represent a storage reservoir of AFB₁ that enhances the toxicity of AFB₁. Mice or rats which are relatively resistant to AFB₁ produce only very little AFL. Therefore, the minor rate of transformation might be a determinant in the susceptibility of animals to the acute toxic action of AFB₁. AFL is reported to be 18 times less toxic than AFB₁ in the duckling biliary \rightarrow hyperplasia assay. In Fischer rats AFL shows nearly one half the hepatocarcinogenic potency of AFB1. Carcinogenicity and mutagenicity (\rightarrow mutagenic) were almost the same as for AFB₁ in rainbow trout and in Salmonella typhimurium, respectively. Biological activity of aflatoxicol B is unknown.

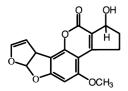
DETECTION see \rightarrow aflatoxins

Further Comments

Two stereoisomers of AFL are known, the "A" isomer, also referred to as aflatoxin R_0 , and the "B" isomer. The latter is only formed by microorganisms whereas AFR_0 also resulted from animal metabolism.

Aflatoxicol H_1 (Abbr.: AFLH₁) is the hydroxylated oxidative metabolite of

Aflatoxicol H₁





 \rightarrow aflatoxicol. It resulted from the metabolism of \rightarrow aflatoxin B₁ by microsomal and soluble enzymes of primate and human liver and from \rightarrow aflatoxin Q₁ incubated with cytosol enzymes.

TOXICITY

No toxicity has been reported in chick embryos and bacteria but it was \rightarrow mutagenic (2% that of AFB₁) in the case of *Salmonella typhimurium*.

Aflatoxicosis is caused by \rightarrow Aspergillus flavus Link and \rightarrow Aspergillus parasiticus Speare due to the formation of \rightarrow aflatoxins. Although these molds are of ubiquitous distribution, A. parasiticus predominates in tropical and subtropical countries. These \rightarrow storage fungi invade seeds and \rightarrow grains, particularly \rightarrow peanuts, \rightarrow maize (before harvest), and edible \rightarrow nuts. Saprophytic growth on a wide range of foodstuffs is possible. Certain climatic conditions favour preharvest invasion and aflatoxin contamination of maize and peanuts. Countries with colder climates do not support aflatoxin production. Here, aflatoxicosis may be imported by contaminated feeds and foods. Species which are mainly affected by aflatoxins are humans, \rightarrow cattle, dogs, \rightarrow poultry, pigs, and trout.

The aflatoxicosis can be divided into two forms: primary aflatoxicosis with the acute and chronic forms, and secondary aflatoxicosis.

Acute aflatoxicosis results from high and moderate aflatoxin concentrations which cause the death of the animal. The main symptoms are: fatty, pale, and decolorized livers; interference of normal blood clotting mechanisms with subsequent hemorrhages (\rightarrow haemorrhage); decrease in total serum proteins and increase in certain serum enzymes of the liver; accumulation of blood in the gastrointestinal canal. In addition, lesions of the kidney (glomerular \rightarrow nephritis) and congestions (\rightarrow congestion) in the lungs are possible.

The most severe case of acute aflatoxicosis has been observed in north-west India (1974). Ca. 25% of the exposed population (397 affected, 106 died) died after eating molded \rightarrow maize with aflatoxin levels ranging from 6250 to 15,600 μ g / kg. In contrast to females males were affected twice as often. Patients suffered from \rightarrow icterus, in general vomiting and \rightarrow anorexia preceded. \rightarrow Ascites and \rightarrow edema of the lower extremities subsequently occurred. In another case of acute aflatoxicosis (Kenya) patients showed similar clinical signs. Pathological changes in the liver were characteristic of toxic \rightarrow hepatitis. In addition, three children in the Province of Taiwan, China and one child in Uganda died from acute liver necrosis. Their death was associated with the ingestion of \rightarrow rice (200 µg aflatoxins/kg) and \rightarrow cassava (1700 µg aflatoxins/kg), respectively, which most probably caused the disease. The reported outbreaks are only seen as the tip of the iceberg of worldwide occurring aflatoxicosis.

Chronic aflatoxicosis is caused by long term consumption of moderate to low aflatoxin concentrations. Much more serious veterinary problem may arise compared to acute aflatoxicosis. Symptoms are: liver congestions with hemorrhagic and necrotic regions; proliferation of the hepatic parenchyma and epithelial cells of the \rightarrow bile duct; kidney congestion accompanied by occasional hemorrhagic \rightarrow enteritis. Reduced feed efficiency and retarded growth rate are common, the reproducive efficiency is decreased. Development of liver cancer (e.g. hatchery-reared trout) may result from longterm consumption of low levels of \rightarrow aflatoxins as extremely potent hepatocarcinogenes.

Secondary aflatoxicosis (low aflatoxin concentrations) impairs the native resistance by reduction of phagocytic effectiveness of macrophages and nonspecific humoral substances (complements). The immunosuppressive effects of aflatoxins predispose animals to secondary infections by bacteria, fungi and viruses. Epidemiological studies in different parts of Africa and Asia show that aflatoxins may cause liver cancer in humans, albeit in combination with the hepatitis B virus. People e.g. living in Kenya, Mozambique, Swaziland and Thailand showed a high incidence of hepatic carcinomas. In these countries \rightarrow foods and feeds are often contaminated with aflatoxins. In the Philippines AFM₁ has been detected in the 24 h urine samples of people who ingested \rightarrow peanut butter containing aflatoxin. A level as high as 10-15 $\mu g \rightarrow a flatoxin B_1$ in the diet seems to be sufficient for detection of \rightarrow aflatoxin M₁ in urine.

Aflatoxin B_1 (Abbr.: AFB₁) is a 2,3,6a,9atetrahydro-4-methoxy-cyclopenta[c]furo[3',2':4,5]furo[2,3-h][1]-benzopyran-1,11-dione (\rightarrow mycotoxins) generally produced in the largest amount both in nature and in culture (see Figure Aflatoxin B₁).

CHEMICAL DATA

Empirical formula: $C_{17}H_{12}O_6$, molecular weight: 312

FUNGAL SOURCES

→ Aspergillus flavus Link, → Aspergillus nomius Kurtzman et al. → Aspergillus parasiticus Speare.

NATURAL OCCURRENCE

 \rightarrow Acacia concinna, almonds, \rightarrow ammi,

 \rightarrow apples, \rightarrow baby food, \rightarrow bacon,

 \rightarrow barley, \rightarrow bean jam, \rightarrow beans, \rightarrow beefburger, \rightarrow beer, burukutu, \rightarrow beer, pito, \rightarrow beer, sorghum, \rightarrow Blepharis edulis, \rightarrow bondakaledkai, \rightarrow Brazil nuts, \rightarrow bread, \rightarrow buckwheat, \rightarrow buckwheat flour, \rightarrow cabbage, \rightarrow Caesalpinea digyna, \rightarrow Cassia fistula, \rightarrow cardamom, \rightarrow cardamom, greater, \rightarrow cashew nuts, \rightarrow cayenne pepper, \rightarrow cereals, \rightarrow cheese, \rightarrow cheese, blue, \rightarrow cheese, pepper, \rightarrow cheese, Tilsit, \rightarrow cheese rind, \rightarrow cheese trimmings, \rightarrow cherries, \rightarrow chicken liver, \rightarrow cocoa beans, \rightarrow congressbele, \rightarrow copra, \rightarrow coriander, \rightarrow corn flakes, \rightarrow cumin, \rightarrow curcuma, \rightarrow dairy products, \rightarrow duck, \rightarrow emu aran, \rightarrow equsi meal, \rightarrow fennel, \rightarrow fenugreek, figs, \rightarrow galgant, \rightarrow garlic, \rightarrow garlic / onions, \rightarrow ginger, \rightarrow groundnut toffee, \rightarrow ham, \rightarrow hare, \rightarrow hazelnuts, \rightarrow hot dog, \rightarrow human breast milk, \rightarrow Hydnocarpus laurifolia, \rightarrow Indian cassia, \rightarrow ingwer, \rightarrow job's-tears, \rightarrow kubeba, \rightarrow lemmons, \rightarrow lentils, \rightarrow libritos, \rightarrow lineseed oil, \rightarrow lineseeds, \rightarrow mackarel, \rightarrow maize flour, \rightarrow maize grits, \rightarrow mango, \rightarrow meat, luncheon, \rightarrow milk, \rightarrow milk powder, \rightarrow miso, \rightarrow muesli, \rightarrow nutmeg, \rightarrow nuts (mixed), \rightarrow oats, \rightarrow oat flakes, \rightarrow ogbono, \rightarrow ogiliugba, \rightarrow ogoro, \rightarrow oil seeds, \rightarrow oil seed rape, \rightarrow olive oil, \rightarrow olives, \rightarrow oranges, \rightarrow pastries, \rightarrow peaches, \rightarrow peanut brittle, \rightarrow peanut butter, \rightarrow peanut oil, \rightarrow peanut products, \rightarrow peas, \rightarrow pecans, \rightarrow persipan, \rightarrow pheasants, \rightarrow pig liver, \rightarrow pine nuts, \rightarrow Piper betle, \rightarrow pipian paste, \rightarrow pop corn, \rightarrow rice, \rightarrow rice cake, \rightarrow roe deer, \rightarrow rye, \rightarrow sago, \rightarrow salami, \rightarrow sausages, \rightarrow shrimp, \rightarrow sorghum, \rightarrow soybean, \rightarrow spices, \rightarrow sunflower seeds, \rightarrow sunflower seed oil, \rightarrow taro, \rightarrow tomatoes, \rightarrow tomato ketchup, \rightarrow tumeric, \rightarrow vegetables, walnuts, \rightarrow wheat For further information see \rightarrow aflatoxins and \rightarrow aflatoxin G₂. Plant commodities which may be highly contaminated with \rightarrow aflatoxins are \rightarrow nuts such as \rightarrow peanuts, Brazil nuts, \rightarrow pistachio nuts as well as copra,

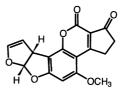
→ maize, and cottonseeds. Agricultural products with a slightly lower potential of aflatoxin contamination are → almonds, → figs, pecans, spices, and → walnuts. Animal products are less likely substrates, e.g. → milk, animal tissue.

TOXICITY

It is the strongest natural carcinogen and the main hepatocarcinogen in animals, although effects vary with species, age, sex, and general nutrition. For example trout, duckling, and pig, are highly susceptible, whereas e.g. sheep and \rightarrow cattle, are more resisant. The liver is the primary organ affected (induction of liver lesions, liver carcinoma, bile duct proliferation). In Fischer rats and rainbow trout AFB₁ is the most potent hepatocarcinogen. Changes in other organs (e.g. kidneys, lung) have been observed. From primate data the doses of AFB1 required to cause acute \rightarrow aflatoxicosis in humans were extrapolated. It was estimated that the intake of \rightarrow food contaminated with 1700 μ g/kg bw for a short time could be sufficient for severe liver damage while a single dose of 75,000 µg/kg bw could result in death. Apparent acute aflatoxicosis would not occur if 340 μ g AFB₁ / kg bw is consumed per day. In the USA the ingestion of AFB₁ with maize and peanut products contributes to a greater risk of hepatic cancer in adults than AFM₁ in milk and \rightarrow dairy products. In comparison to these agricultural products the human intake of aflatoxins by meat and meat products is negligible. The IARC (1993) evaluated AFB_1 as a Class 1 human carcinogen.

 LD_{50} (po): 5.5-7.2 mg/kg bw male rats (weight: 100 g), 17.9 mg/kg bw female rats (weight: 150 g)

DETECTION see \rightarrow aflatoxins



Aflatoxin B₁

Further Comments

Spiking commercially manufactured cigarettes with AFB_1 (100-300 µg/kg) did not result in any contamination of the gas phase or the ashes.

Aflatoxin B₂ (Abbr.: AFB₂) is the dihydro derivative of \rightarrow aflatoxin B₁ (2,3,6a,8,9ahexahydro-4-methoxy-cyclopenta[c]furo[3',2':4,5]furo[2,3-h][1]-benzopyran-1,11-dione) and synthesized by the reduction of the single double bond in the terminal dihydrofuran ring (\rightarrow mycotoxins) (see Figure Aflatoxin B₂).

CHEMICAL DATA Empirical formula: $C_{17}H_{14}O_6$, molecular weight: 314

FUNGAL SOURCES

→ Aspergillus flavus Link, → Aspergillus nomius Kurtzman et al., → Aspergillus parasiticus Speare

NATURAL OCCURRENCE

 AFB_2 occurs in the same commodities as AFB_1 but AFB_2 is found in smaller amounts. Via milk it is secreted as \rightarrow aflatoxin M₂.

TOXICITY

This carcinogenic (?) and \rightarrow genotoxic substance shows toxic properties similar to AFB₁ but has markedly reduced toxic potency in comparison to AFB₁. Instead of 3.9 µg AFB₁ 50 µg AFB₂ are necessary to produce similar bile duct proliferation in ducklings. Estimated lethal dose for human beings 1-10 mg/kg.

 LD_{50} (po): 84.8 µg / 50 g bw one-day old ducklings

DETECTION see \rightarrow aflatoxins

Aflatoxin B_{2a} (Abbr.: AFB_{2a}) (Syn.: AFB_1 hemiacetyl, aflatoxin W, hydroxydihydroaflatoxin B_1) represents the corresponding "water adduct" (2-hydroxy derivative) of \rightarrow aflatoxin B₁ (\rightarrow mycotoxins) which resulted from the hydration of the 2,3-vinyl ether bond of this aflatoxin (2,3,6a,8,9,9a-hexahydro-8-hydroxy-4methoxy-cyclopenta[c]furo[3',2':4,5]furo[2,3-h][1]-benzopyran-1,11-dione). Conversion occurs rapidly under mildly acidic conditions. Although this blue fluorescing compound is 60-100 (200) times less toxic to ducklings it may be dehydrated to the highly toxic AFB₁. Furthermore, AFB_{2a} is a biotransformation / detoxification product of AFB₁ produced by hepatic microsomes in vitro of some animals (e.g. mouse, guinea-pig, avian). It is under discussion whether AFB_{2a} reacts readily with free amino groups of functional proteins (see Figure Aflatoxin B22).

CHEMICAL DATA Empirical formula: $C_{17}H_{14}O_7$, molecular weight: 330

Fungal Sources

 \rightarrow Aspergillus flavus Link, \rightarrow Aspergillus parasiticus Speare

Тохісіту

In the standard duckling assay (initiation of \rightarrow bile duct proliferation) both AFB_{2a} and AFG_{2a} are very much less toxic than AFB₁ (60-100 times) after oral application. In Khaki Campbell ducklings (day-



old) no acute toxicity was noted at levels up to 1200 μ g / duckling.

Aflatoxin B₃ (Abbr.: AFB₃) (Syn.: parasiticol) Older cultures of \rightarrow Aspergillus flavus Link and \rightarrow Aspergillus parasiticus Speare may contain high amounts of this 6-methoxy-7-(2'hydroxyethyl) difurocoumarin (7a,10a-dihydro-4-(2-hydroxyethyl)-5-methoxy-2H-furo[3',2':4,5]-furo[2,3-h]-1-benzopyran-2-one) as a possible precursor of \rightarrow aflatoxins. On the other hand it seems to be the first step in the biological degradation of \rightarrow aflatoxin G₁ by e.g. *Rhizopus* spp. (see Figure Aflatoxin B₃).

CHEMICAL DATA

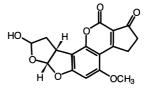
Empirical formula: $C_{16}H_{14}O_6$, molecular weight: 302

TOXICITY

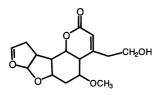
Parasiticol has the same acute toxicity to ducklings as \rightarrow aflatoxin B₁. However, the tendency to cause biliary \rightarrow hyperplasia is low. In chick embryo studies toxicity was only 1/100 than that of AFB₁.

Aflatoxin D₁ is a major product (10-30%) - besides the 206-molecular weight compound (3-10%) - from the reaction of aflatoxin B₁ with heated ammonium hydroxide. aflatoxins

Aflatoxin G_1 is a mycotoxin (\rightarrow mycotoxins) that has a structure very similar to that of \rightarrow aflatoxin B_1 (3,4,7a,10a-tetrahydro-5-methoxy-1H,12H-furo[3',2':4,5]furo[2,3-h]pyrano[3,4-c][1]-benzopyran-1,12-dione) but there are two lactone functions rather than one and the two



Aflatoxin B_{2a}



Aflatoxin B₃

dihydrofuran rings are fused in a *cis* configuration (see Figure Aflatoxin G_1).

CHEMICAL DATA

Empirical formula: $C_{17}H_{12}O_7$, molecular weight: 328

FUNGAL SOURCES

→ Aspergillus flavus Link, → Aspergillus nomius Kurtzman et al., → Aspergillus parasiticus Speare

NATURAL OCCURRENCE

Same commodities as AFB_1 , in addition, \rightarrow celery seeds.

Тохісіту

This carcinogenic (liver- and kidney carcinoma) and \rightarrow genotoxic mycotoxin possesses a similar toxicity to that of AFB₁, although acute toxicity was less than AFB₁ but greater than AFB₂. It is a slightly less potent liver carcinogen but a slightly more potent kidney carcinogen, with a comparable carcinogenic potency to aflatoxin B_1 i.e. within a factor of 10. Ducklings treated with AFG₁ showed the same lesions as AFB₁-treated animals. The zone in affected rat liver lobule was the same as in B_1 . However, a consistent pattern as seen with AFB₁ was absent. The LD_{50} in the rat was twice that of AFB₁.

DETECTION see \rightarrow aflatoxins

tion is 30 °C.

FURTHER COMMENTS Optimum temperature for AFG₁ produc-

Aflatoxin G₂ is the dihydro derivative of \rightarrow aflatoxin G₁ (3,4,7a,9,10,10a-hexahy-dro-5-methoxy-1H,12H-furo[3',2':4,5]-

furo[2,3-h]pyrano[3,4-c][1]-benzopyran-1,12-dione) and synthesized by the reduction of the single double bond in the terminal dihydrofuran ring (see Figure Aflatoxin G_2).

CHEMICAL DATA Empirical formula: $C_{17}H_{14}O_7$, molecular weight: 330

FUNGAL SOURCES \rightarrow Aspergillus flavus Link, \rightarrow Aspergillus nomius Kurtzman et al., \rightarrow Aspergillus parasiticus Speare

NATURAL OCCURRENCE

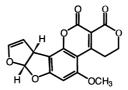
→ beans, → brazil nuts, → cocoa beans, → cumin, → ginger, → Indian cassia, → lemons, → maize, → mango, → olive oil, → oranges, → peanuts, → peanut brittle, → pepper, → pop corn, → rice, → sausages, → sesame seeds, → shoyu, → sunflower seeds, → tumeric, → walnuts For further information see → aflatoxins and → aflatoxin B₁. ToxICITY This carcinogenic (?) and → genotoxic

mycotoxins possesses the least acute toxicity of the 4 major naturally occurring \rightarrow aflatoxins.

 LD_{50} (po): 172.5 µg/50 g bw one day old ducklings.

DETECTION see \rightarrow aflatoxins

Aflatoxin G_{2a} (Abbr.: AFG_{2a}) Aflatoxin G₁ is converted by strong acids to the corresponding "water adduct" (2-hydroxy derivative = AFG_{2a}) which retains its



Aflatoxin G₁

toxicity (3,4,7a,9,10,10a-hexahydro-9hydroxy-5-methoxy-1H,12H-furo[3',2':4,5]furo[2,3-h]pyrano[3,4-c][1]benzopyran-1,12-dione). Livers of certain animals ingesting \rightarrow aflatoxin G₁ produce AFG_{2a} which might be a detoxification mechanism (see Figure Aflatoxin G_{2a}).

CHEMICAL DATA

Empirical formula: $C_{17}H_{14}O_7$, molecular weight: 330

FUNGAL SOURCES

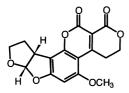
 \rightarrow Aspergillus flavus Link, \rightarrow Aspergillus parasiticus Speare

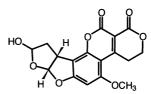
TOXICITY

No significant differences in growth and characteristic liver lesions occurred in day-old Khaki Cambell ducklings (1600 μ g/duckling). LD₅₀ of AFB₁ in the same assay was 18.2 μ g/duckling.

Aflatoxin GM₁ is a 4-hydroxylated derivative of \rightarrow aflatoxin G₁ but only minor quantities have been detected in \rightarrow Aspergillus flavus Link cultures.

Aflatoxin M₁ (Abbr.: AFM₁) is the 4hydroxylated derivative of \rightarrow aflatoxin B₁ (2,3,6a,9a-tetrahydro-1,9a-dihydroxy-4methoxy-cyclopenta[c]furo[3´,2´:4,5]furo[2,3-h][1]-benzopyran-11(1H)-one). It is found in liver, kidneys, blood, bile, feces, urine, and \rightarrow milk of mammals (\rightarrow mycotoxins). Hydroxylation mainly occurs in the liver in the benzylic position at the junction of the two furan rings. It was the first \rightarrow aflatoxin B₁ metabolite identified which was originally (early 1960s) found in cow's milk. Struc-





Aflatoxin G_{2a}

tural elucidation was first achieved in 1966. Subsequently isolation of AFM_1 has also been reported from other kinds of milk as well as \rightarrow dairy products (see Figure Aflatoxin M₁).

Chemical Data

Empirical formula: $C_{17}H_{12}O_7$, molecular weight: 328

FUNGAL SOURCES \rightarrow Aspergillus flavus Link, \rightarrow Aspergillus parasiticus Speare

NATURAL OCCURRENCE \rightarrow cheese, \rightarrow cheese, blue, \rightarrow cheese, Blue Haverti, \rightarrow cheese, Brie, \rightarrow cheese, butter, \rightarrow cheese, Camembert, \rightarrow cheese, Camembert & Brie, \rightarrow cheese, Cheddar, \rightarrow cheese, Cheshire, \rightarrow cheese, Chester, \rightarrow cheese, Cottage, \rightarrow cheese, Comte, \rightarrow cheese, Cream, \rightarrow cheese, Double Gloucester, \rightarrow cheese, Edam, \rightarrow cheese, Emmental, \rightarrow cheese, Fresh, \rightarrow cheese, Gouda, \rightarrow cheese, Grana Padano, \rightarrow cheese, Lancashire, \rightarrow cheese, Leicester, \rightarrow cheese, Maribo, \rightarrow cheese, Mozarella, \rightarrow cheese, Parmesan, \rightarrow cheese, Romadur, \rightarrow cheese, Samsoe, \rightarrow cheese, Stilton, \rightarrow cheese, Wensleydale, \rightarrow cheese, Wine, \rightarrow cream, full, \rightarrow human breast milk, \rightarrow milk, \rightarrow milk powder, \rightarrow milk, pasteurized, \rightarrow milk, sterilized, \rightarrow milk, UHT, \rightarrow milk, camel, \rightarrow pistachio nuts, \rightarrow soybean milk powder, \rightarrow whey powder, → yogurt Besides milk and dairy products this mycotoxin (\rightarrow mycotoxins) is also a contaminant of stored white and yellow \rightarrow maize, freshly harvested yellow maize,

and acid treated stored yellow maize (1-

Aflatoxin G₂

Aflatoxin M₁

35 μ g/kg) as well as moldy \rightarrow peanuts. AFM₁ has also been found in \rightarrow human breast milk samples as a hydroxylated derivative of AFB₁ due to the activity of cytochrome P4501A2.

The ingestion of AFB₁-contaminated feed by mammals leads to the excretion of AFM₁ in milk (\rightarrow carry over ca. 0.3-3%, in dairy cows in early lactation up to 6%) and urine. 85% of dosed AFB₁ is secreted as AFM₁ via milk and urine within 48 hours. First dectection of AFM₁ within 12 hours. A milk sample taken after 96 hours was free of aflatoxin. Milk and dairy products are most probably the only toxic hazard from animal products. Concentration of AFM in body tissues is usually low with the majority of reports indicating undetectable levels in meat, blood, fat etc.

TOXICITY

 LD_{50} : 16.6 µg AFM₂ / day old duckling; 12 µg AFB₁ / day old duckling (simultaneous application)

A slightly less capacity in inducing \rightarrow hepatic carcinoma (trout, rats) has been observed compared to AFB₁. There was inadequate evidence of the human carcinogenicity of AFM₁ (IARC 1993). AFM₁ induced hepatocarcinoma in trout and occasionally subcutaneous \rightarrow sarcoma after injection.

DETECTION

see \rightarrow aflatoxins

FURTHER COMMENTS

In some countries the contamination of milk with AFM_1 may follow a seasonal trend. During summer months lower contamination levels are detected because less supplementary mixed feeds are added to the diets of dairy cattle. AFM_1 is associated with the protein fraction of the milk. This fact is responsible for the contamination of cheeses (3-5 fold enrichment).

In contrast to AFB₁, AFB₂, AFG₁, and AFG₂ (\rightarrow aflatoxins) AFM₁ also occurs in the absence of other aflatoxins. Human exposure is primarily due to milk and milk products from animals that ingested AFB₁-contaminated feed. AFM₁ may cause problems especially in infants with a high milk consumption because of relatively low body weight, high cell activity, and partially developed immune system.

Stability: AFM₁ is stable in raw milk. Processing of contaminated milk will not result in aflatoxin-free dairy products. No reduction was established after pasteurization or processing into cheese, \rightarrow yogurt, and \rightarrow cream (20-40% fat). However, other reports proved a 63% reduction after pasteurization, 80% after sterilization and 85% after dry milk processing. Depending upon the time a 100% degradation of AFM₁ was achieved by UV irradation.

Aflatoxin M₂ (Abbr.: AFM₂) is the 4hydroxylated derivative of \rightarrow aflatoxin B₂ (2,3,6a,8,9,9a-hexahydro-9a-hydroxy-4methoxy-cyclopenta[c]furo[3´,2´:4,5]furo[2,3-h][1]-benzopyran-1,11-dione) and found in liver, kidneys, urine, and \rightarrow milk of mammals (see Figure Aflatoxin M₂).

CHEMICAL DATA Empirical formula: $C_{17}H_{14}O_7$, molecular weight: 330

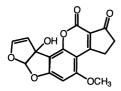
FUNGAL SOURCES \rightarrow Aspergillus flavus Link, \rightarrow Aspergillus parasiticus Speare

NATURAL OCCURRENCE \rightarrow human breast milk

TOXICITY

Compared to $AFM_1 AFM_2$ is considerably less toxic. This may be due to the lack of the double bond terminating the difuran ring system which is common in AFB_1 , AFG_1 , and AFM_1 .

Aflatoxins



Aflatoxin M1

 LD_{50} : 62 µg AFM₂/day old duckling; 12 µg AFB₁/day old duckling (simultaneous application).

DETECTION see \rightarrow aflatoxins

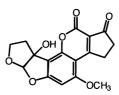
FURTHER COMMENTS

Compared to \rightarrow aflatoxin M₁ AFM₂ has a lower $R_{\rm f}$ with a violet fluorescence.

Aflatoxin M_4 (Abbr.: AFM₄) In 1986 the metabolite AFM₄ was isolated and identified in cow \rightarrow milk. Contamination of commercial milk samples with this aflatoxin has been reported in France and Italy. The particular nutritional condition of the cow may be decisive for the mammary excretion of AFM₄. The name, AFM₄, derived from the fact that the hydroxyl group was located at carbon 4 of the cyclopentenone ring of AFM₁ (2,3,6a,9a-tetrahydro-2-hydroxy-4-methoxy-cyclopenta[c]furo[3',2':4,5]furo[2,3h][1]-benzopyran-1,11-dione). \rightarrow aflatoxins

Aflatoxin P₁ (Abbr.: AFP₁) represents the principal urinary metabolite in rhesus monkeys after intraperitoneal injection of \rightarrow aflatoxin B₁. It showes considerably less toxicity than AFB₁. In mice and humans hepatic microsomes are also responsible for the metabolization of AFB₁ to AFP₁.

Aflatoxin Q₁ (Abbr.: AFQ₁) is the 3hydroxy metabolite of \rightarrow aflatoxin B₁. The major metabolic product of the metabolism in monkey, rat, and human liver



Aflatoxin M₂

microsomes preparations was approximately 18 times less toxic than AFB₁. No \rightarrow mutagenic activity was detected.

Aflatoxin $R_0 \rightarrow$ aflatoxicol

Aflatoxin W (Syn.: \rightarrow aflatoxin B_{2a})

Aflatoxins Aflatoxins as causing agents of the \rightarrow turkey "X" disease were responsible for the death of more than 100,000 \rightarrow turkey poults, aged three to six weeks, in south east England in 1960. A shipment of peanut meal ("Rosetti" meal) imported from Brazil as a by-product from the extraction of \rightarrow peanut oil, was contaminated by \rightarrow Aspergillus flavus Link (but actually \rightarrow Aspergillus parasiticus Speare) and contained four distinct, fluorescent highly toxic substances: \rightarrow aflatoxin B₁, \rightarrow aflatoxin B₂, \rightarrow aflatoxin G_1 , and \rightarrow aflatoxin G_2 (Aspergillus flavus toxin A-fla-toxin). Later it could be shown that \rightarrow cyclopiazonic acid was also involved in turkey "X" disease. Besides \rightarrow poultry which showed hemorrhages $(\rightarrow \text{hemorrhage})$ and liver necrosis frequently accompanied by lesions of the kidney e.g. ducklings, pigs, and \rightarrow cattle were also affected. Toxicity of the aflatoxins comprises hepatocarcinogenicity, reduced T-cell function, diminished antibody response, and suppressed phagocyte activity. For further information see each single aflatoxin.

Aflatoxins are polycyclic, unsaturated highly substituted coumarins and one of the most important \rightarrow mycotoxins.

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Aflatoxins

The proposed natural synthesis of aflatoxin B₁ is as follows: acetate, norslorinic acid, averantin, averufanin, averufin versiconal hemicacetal acetate, versicolorin A, sterigmatocystin, O-methylsterigmatocystin, AFB₁. The first substance in the pathway to contain the essential C_2 - C_3 double bond is versicolorin A. Aflatoxins possess a fused dihydrofuran configuration that is peculiar to a limited number of compounds of natural origin. The coumarin nucleus is fused to a reactive bifuran system on one side and either a pentanone (B-aflatoxins) or a six-membered lactone (G-aflatoxins) on the other. The aflatoxin molecule probably has two reactive (toxic/carcinogenic) sites, viz. the unsatured terminal site in the bihydrofuran moiety and the lactone ring of the coumarin part. Although approximately 20 aflatoxins have been identified only four of them, aflatoxins B₁, B₂, G₁ and G₂, occur naturally. The letters B₁, B₂, G₁, and G₂, are due to their intensive blue (B-aflatoxins) and green (G-aflatoxins) fluorescence in UV light. The subscripts indicate the relative chromatographic mobility. Two other familar aflatoxins, \rightarrow aflatoxin M₁ and \rightarrow aflatoxin M₂, are usually "metabolites" (mammal transformation products) of AFB₁ and AFB₂. They are labeled so because of their presence in "milk" (milk toxin) previously exposed to AFB₁ and AFB₂. However, isolation of the 4-hydroxylated aflatoxins has also been reported from peanuts and \rightarrow maize. The M toxins fluoresce blue to violet when exposed to long-wave UV light, but separate at a lower $R_{\rm f}$ value on TLC plates than AFB and AFG toxins. Besides the AFM-toxins further aflatoxins derived from AFB₁, AFB₂, AFG₁ and AFG₂ as metabolic products of microbial or animal systems (e.g. \rightarrow aflatoxin P₁, \rightarrow aflatoxin Q₁ and \rightarrow aflatoxicol) or produced spontaneously in response to the chemical environment (e.g. B_{2a} , G_{2a} , and D_1).

In spite of the worldwide distribution of A. flavus (A. parasiticus predominates in warmer climates, \rightarrow Aspergillus nomius Kurtzman et al. is a sporadic contaminant of \rightarrow food) different factors favor aflatoxin contamination of \rightarrow foods and feeds in distinct areas of the world. The \rightarrow a_w of the commodity and the surrounding relative humidity as well as temperature are most decisive in storage and in the field. Preharvest invasion with A. flavus and subsequent aflatoxin contamination occurs in the case of peanuts and maize. Factors that promote invasion and contamination are drought stress in plants, drought-enhanced insect damage, wet weather conditions in combination with high temperatures during harvest, and use of susceptible genotypes.

CHEMICAL DATA

 $\begin{array}{l} \mbox{see:} \rightarrow \mbox{aflatoxin } B_1, \rightarrow \mbox{aflatoxin } B_2, \\ \rightarrow \mbox{aflatoxin } B_{2a}, \rightarrow \mbox{aflatoxin } G_1, \rightarrow \mbox{aflat} \\ \mbox{toxin } G_2, \rightarrow \mbox{aflatoxin } G_{2a}, \rightarrow \mbox{aflatoxin } B_3, \\ \rightarrow \mbox{aflatoxin } M_1, \rightarrow \mbox{aflatoxin } M_2 \end{array}$

FUNGAL SOURCES

Only 3 species, *A. flavus* Link, *A. parasiticus* and *A. nomius*, are definite producers of aflatoxins. Approximately 50% of all *A. flavus* strains synthesize aflatoxin. A higher percentage is found in warmer climates than in cooler regions, e.g. Ex-CSSR only 6 of 694 strains were aflatoxin-positive. Aflatoxins are found in the mycelium of *A. flavus* Link, in the conidia (84 mg/kg AFB₁, 566 mg/kg AFG₁) and sclerotia (135 mg/kg AFB₁, 968 mg/kg AFG₁) (see Figure Aflatoxins).

The domesticated forms of A. flavus and A. parasiticus (\rightarrow Aspergillus oryzae (Ahlburg) Cohn, A. sojae) have completely lost their ability to produce aflatoxins and the corresponding precursors.

NATURAL OCCURRENCE

- \rightarrow bakery products, \rightarrow barley grits,
- \rightarrow beer, \rightarrow breakfast cereals, \rightarrow cassava,
- \rightarrow chilli, \rightarrow chilli pickles, \rightarrow chilli powder,

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 \rightarrow chocolate, \rightarrow cocoa nibs, \rightarrow cocoa presscake, \rightarrow coconut, \rightarrow coconut ice, \rightarrow coconut oil, \rightarrow coffee beans, \rightarrow cow peas, \rightarrow curry, \rightarrow curry paste, \rightarrow egg products, \rightarrow fig paste, \rightarrow fish, \rightarrow foods, \rightarrow garlic pickle, \rightarrow garlic powder, \rightarrow grains, \rightarrow maize bran, \rightarrow maize malt, \rightarrow maize meal, \rightarrow maize products, \rightarrow maize starch, \rightarrow maize, boiled, \rightarrow maize, canned, \rightarrow maize, dried, \rightarrow maize, shelled, \rightarrow manioc, \rightarrow marzipan, \rightarrow meat, \rightarrow melon balls snacks, \rightarrow melon seeds, \rightarrow millet, \rightarrow muesli ingredients, \rightarrow noodles, \rightarrow nuts, oil, \rightarrow peach kernels, \rightarrow peanut candy, \rightarrow peanut mix, \rightarrow peanut paste, \rightarrow peanut sauce, \rightarrow pigeon pea, \rightarrow pistachio candy, \rightarrow poppadoms, \rightarrow pumpkin seeds, \rightarrow small grains, \rightarrow soybean flour, \rightarrow tandoori, \rightarrow tubers, \rightarrow wine For further information see aflatoxin B_1 and aflatoxin G₂. Foods which show a predisposition for

aflatoxin contamination include maize and \rightarrow maize products, peanuts and \rightarrow peanut products, \rightarrow pecans, \rightarrow almonds, \rightarrow hazelnuts, \rightarrow Brazil nuts,

→ aimonds, → mazemuts, → brazimuts, → pistachio nuts, and → walnuts. Small food → grains, e.g. → soybeans, → barley, → rye, → rice, and → oats, are not a major source of aflatoxin exposure if stored under suitable conditions. Other kinds of foodstuff which were found positive for aflatoxin contamination are e.g. cassava, → peas, → cowpeas, millet, → sorghum, sesame, sweetpotatoes, → spaghetti.

Of the aflatoxins present in food AFB₁, AFG₁, and AFM₁ are of primary importance and, together with aflatoxicol, represent possible health concerns. Although AFB₁, AFB₂ and AFG₁ are common in the same food sample AFB₁ predominates (60-80% of the total aflatoxin content). Generally, AFB₂, AFG₁, and AFG₂ do not occur in the absence of AFB₁. In most cases AFG₁ is found in higher concentrations than AFB₂ and AFG₂. Maximum concentrations have been recorded from the following seeds: cotton: > 5 g aflatoxin / kg, peanuts: 1 g aflatoxin / kg, maize: 0.4 g aflatoxin / kg, pistachio nuts: 1.4 g AFB_1 / kg. Unprocessed foods of plant origin seem to be the most important potential sources of aflatoxins in the diet. In contrast, animal products are less likely substrates for aflatoxin contamination. Sugar, conventional jellies, sauerkraut, raisins and potatoes are generally free of aflatoxins.

The contamination of agricultural products with aflatoxins is not only a problem in less developed countries (tropics) but also in (warm) regions with a high developed agricultural standard (southern and sometimes mid-western USA). Plant stress, insufficient drying after harvest and storage at relatively high temperatures are the main reasons for contamination. Maize and peanuts are particularly susceptible. If these crops belong to the staple foods a higher exposure level to aflatoxins may be the consequence.

Τοχιζιτγ

AFB₁, AFM₁ and aflatoxicol belong to the group of \rightarrow genotoxic carcinogens with AFB₁ being the most potent. Aflatoxins with the index 1 are the most toxic ones. For this type of carcinogen, there is no threshold dose below which no tumor formation would occur. Only a zero level of exposure will result in no risk. Even very low concentrations, e.g. 1 ng aflatoxin / kg bw / day or less still contribute to the risk of liver cancer.

Besides their carcinogenic effect aflatoxins are \rightarrow mutagenic, \rightarrow teratogenic, and hepatogenic. In low levels they are responsible for weight gain losses, loss of reproducive capacity, and impairment of the immune systems (e.g. poultry, pigs, cattle). Conversion of AFB₁ and AFG₁ by hydroxylation to B_{2a} and G_{2a}, respectively, greatly reduces oral toxicity. The bio-

Aflatoxins

chemical effects are inhibition of adenosine triphosphatase (energy metabolism), reduction of hepatic glycogen levels (carbohydrate, lipid metabolism), binding with DNA and RNA (nucleic acid, protein metabolism).

The NOAEL for AFB₁ was estimated as 0.75 μ g/kg body weight per day, using Fisher exact (statistical) test. Similarly, for aflatoxicol and AFM₁, the respective NOAELs were 1.25 and < 2.5 μ g/kg bw per day.

Nutritional status of individuals exposed to aflatoxins seems to be very important for human health because malnutrition coexists in many parts of the world with aflatoxins and high incidences of liver disease, including liver cancer. Furthermore, susceptibility of monkeys to aflatoxins was significantly increased by reduced protein intake. The estimated LD_{50} for humans is about 1-10 mg AFB₁ / kg.

The carcinogenicity of aflatoxins is enhanced by e.g. gossypol, 3-methylcoumarin, cycloproprenoid fatty acids, malvalic acid and sterculic acid but possibly also by deoxynivalenol and nivalenol (synergistic effect).

DETECTION

ELISA, HPLC, IACA, RIA, TLC

The aflatoxin contamination (and other mycotoxins) of seeds is characterized by a negative binomial distribution function. Because very few seeds contain any significant level of aflatoxin but the majoritiy are non-contaminated, a representative sample from the lot must be taken.

Possible Mycotoxicosis

 \rightarrow Aflatoxicosis (acute), \rightarrow Indian childhood cirrhosis, \rightarrow Kwashiorkor, \rightarrow primary hepatocellular carcinoma (PHC), \rightarrow Reye's syndrom

FURTHER COMMENTS

Production: The highest amounts of aflatoxins are synthesized in the log phase (intense sporulation), while aflatoxin production starts at the same time as the formation of conidia. Usually after six days aflatoxin production decreases. Under a given set of conditions only two or three aflatoxins are produced. Although growth in culture yields higher AFG₁ concentration than AFB₁ in the case of natural contamination AFB₁ is frequently found in the highest concentration (e.g. "Rosetti meal" as causual agent of the "turkey X disease" contained 10,000 µg AFB₁/kg but negligible levels of G₁).

The limiting a_w for aflatoxin production (*A. flavus*) is between 0.83 and 0.87, which is close to the minimum for growth. Synthesis increased at a_w 0.95-0.99 with optimum temperatures ranging from 25 °C to 30 °C.

Low temperatures (8-10 °C) induce production of approximately equal amounts of aflatoxins B and G. However, total production is lowered and more time required. Aflatoxin B production is stimulated by higher temperatures relative to aflatoxin G.

Optimal AFB₁ production occurred between 24-28 °C whereas 30 °C is optimal for AFG₁ formation. Only a few strains are able to synthesize aflatoxins at 7.5 °C. Fluctuating temperatures (mean 25 °C, upper limit 40-50 °C) are less favorable for aflatoxin production than a constant temperature (25 °C). Fluctuations down to 10 °C did not cause any significant effect.

 CO_2 - > 10% or O_2 -concentration < 20% or > 90% suppresses toxin production. The addition of cadmium, iron and molybdenium increases aflatoxin production, zinc is a prerequisite. For any given strain of fungus, the substrate influences the amount of aflatoxin produced. Aflatoxin synthesis in the conidia of *A*. *flavus* is stimulated by irradation \leq 3 kGy. A dose of 2.5 kGy enhanced synthesis of AFB₁ and AFG₁ 50 times while 1 kGy was sufficient to induce aflatoxin formation of non-producing A. flavus strains.

Stability: Aflatoxins are extremely heat stable compounds in the dry state up to the melting point. Moisture lowers heat stability but in general these mycotoxins are also stable during thermal processing of most food products. At elevated temperatures a partial destruction could be observed during autoclaving or roasting of nuts (40 to 60%). Cooking processes (e.g. dry heating, boiling) of cereal products, extrusion of maize meal dough (150 °C), or fermentation of dough during breadmaking cause variable losses of aflatoxins. As little as 14-26% of AFB1 contamination of wheat was found to survive flour \rightarrow milling and bread baking. However, baking temperatures are usually not sufficient to cause significant losses of aflatoxin in bread.

Decomposition occurs after exposure to sunlight, ultraviolet ligth and ionizing radiation.

Destruction or removal of aflatoxins from \rightarrow food oils is achieved by alkaline treatments and refining, respectively. Aflatoxin concentration decreased in raw peanut butter and meat with increasing storage time but other studies do not report significant changes in aflatoxin levels of stored peanut meal and peanut butter. An essential stability (after one week) of AFB₁ and AFG₁ in Swiss cheese, bologna and cooked cornmeal was observed.

Cleaning and milling do not cause a general reduction of aflatoxin levels in cereal grains. A redistribution of the mycotoxins in the different fractions is most likely. E.g. dry milling of \rightarrow maize usually leads to increased AFB₁ levels in the germ, hull, and degermer fines fractions. However, the ultimate distribution depends on the original amount present in the seed. Although milling of \rightarrow rice and parboiled rice caused a significant decrease in AFB₁ and AFG₁ levels, afla-

toxin concentrations in the \rightarrow bran and polished fractions increased substantially. Increasing AFB₁ concentrations have also been found in the \rightarrow flour of durum wheat from the top grade to the second. The bran contained the highest amounts. Mashing and brewing caused a partial loss of AFB₁ while distillation destroyed total aflatoxins in excess of 90%. Fermentation of AFB₁ contaminated maize under conditions used in the spirits industry led to aflatoxin-free distilled ethyl alcohol. In completely processed \rightarrow beer only 18-27% of the original AFB₁ concentration was detected. Wort boiling and final fermentation steps mainly contribute to aflatoxin losses.

Detoxification: Detoxifiaction processes include degradation, destruction, or inactivation.

Physical methods: Heat - roasting temperatures (> 250 °C) are necessary for effective aflatoxin degradation; increasing the moisture content of the substrate will enhance degradation; irradiation - effective dose levels (X-rays, electron irradiation) cause destruction of the contaminated commodity; adsorption - \rightarrow bentonite adsorbed aflatoxins from \rightarrow milk and fluid products, hydrated sodium calcium aluminosilicate is suitable for the adsorption of AFB₁ from aqueous solutions. Similar effects have been reported for clays, charcoal, asbestos, aluminas, silicas, xeolites and aluminosilicates; solvent extraction - 90% aqueous acetone, 95% ethanol, hexane-ethanol, hexane-methanol, and 80% isopropyl alcohol have been used effectively.

Chemical methods:

Ammonia causes lactone ring opening of AFB₁, ultimate splitting off of the cyclopentenone part by NH₃. Several breakdown products of AFB₁ have been identified, e.g. \rightarrow aflatoxin D₁ and the 206 molecular weight compound. Both substances showed a 450-fold decrease in mutagenicity (Ames test) compared to

AFB₁. The treatment prevents both acute and chronic aflatoxicosis in animals and is generally believed to be the most effective decontamination method. Ammonia treatment is used on commercial scale for the decontamination of feedstuff including corn and peanut and cottonseed meals in France, Senegal, USA (Arizona, California, Georgia, Alabama). Acids effectively convert AFB₁ and AFG₁ to their corresponding hemiacetal forms \rightarrow aflatoxin B_{2a} und \rightarrow aflatoxin $G_{2a},$ but they have no effect on AFB₂ or AFG₂. Oxidising agents, ozone - destruction of AFB₁ and AFG₁ but not AFB₂; hydrogen peroxide - destruction of aflatoxins in peanuts; in combination with riboflavin destruction of AFM₁ in milk; bisulfite reaction with AFB₁ and AFG₁; vitamin C treatment.

Biotransformation: Microorganisms such as bacteria, actinomycetes, yeasts, molds, and algae cause degradation of aflatoxins. The most effective one, \rightarrow Flavobacterium aurantiacum, removes AFB₁ (and AFM₁) from milk, maize, \rightarrow maize oil, peanuts, \rightarrow peanut butter, and soybeans while AFG₁ and AFM₁ are also metabolized. Other microorganisms convert or transform AFB₁ to aflatoxicol which is a very slow (3 to 4 d) and incomplete process (60% of AFB_1 is converted to aflatoxicol). However, except for ammonification (see above) the remaining methods are only of limited realistic commercial benefit. Control: Control of aflatoxin contamination extends from growth of the crops in the field, through the storage of harvested crops, to the proper storage of prepared foods in the home.

Prevention of aflatoxin contamination of agricultural products, especially high-risk crops such as maize and peanuts, starts in the field. Growth of *A. flavus* and *A. parasiticus* is impaired / inhibited by breeding (using) resistant varieties, good agronomic practices from planting to cultivation and harvesting. Prevention of insect and mechanical damage as well as chemical plant protection favor the optimal development of the plants. During storage low moisture content and temperature, adequate aeration, and pest control (insects, mites) inhibit aflatoxin (mycotoxin) accumulation in the harvested crops. Especially in stored maize, hot and humid storage conditions contribute to elevated aflatoxin concentrations. In the home proper storage of prepared foods for prolonged periods at low humidity and temperature prohibits aflatoxin contamination. At least in the EU there are uncertainties about the dietary aflatoxin intake since detailed information concerning the ingestion of typically aflatoxin-containing foods like peanuts, pistachio and Brazil nuts, figs etc. are difficult to obtain.

Agranulocytosis Absence of granules in cells in cytoplasm. \rightarrow Alimentary toxic aleukia

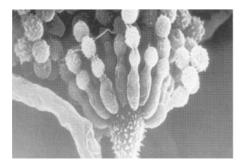
Akakabi byo disease (Syn.: \rightarrow red mold toxicosis, scabby grain intoxication)

Aleukia Absence of leukocytes (\rightarrow leukocytosis) from blood

Alimentary hemorrhagic aleukia \rightarrow Alimentary toxic aleukia

Alimentary mycotoxicosis → Alimentary toxic aleukia

Alimentary toxic aleukia (Abbr.: ATA) (Syn.: septic angina, alimentary panhematopathy, alimentary toxicosis, alitoxicosis, alimentary \rightarrow agranulocytosis, endemic panmyelotoxicosis, hemorrhagic (\rightarrow hemorrhage) syndrome) This \rightarrow mycotoxicosis was first described in 1913 with first indications in 1891. ATA occurred sporadically during the first three decades of the century (e.g. 1924,



Aflatoxins. Small vesicle with phialides and conidia of *Aspergillus flavus* Link

1934) in various parts of Russia, but from 1932 on it appeared in endemic form. It claimed many victims (mortality 2-80%) especially during World War II. People of the Siberian USSR and the Orenburg district were mainly affected. During spring 1944 the morbidity in this district exceeded 10% and a high mortality was observed in 9 of 50 counties. Until the postwar years (1947) the disease caused the death of hundreds of thousands of people.

Mild winters with heavy snow in combination with frequent alternate freezing and thawing in the spring favored fungal growth in grains, especially proso \rightarrow millet and \rightarrow wheat (most likely to be toxic) as well as \rightarrow barley, \rightarrow rye, \rightarrow oats, and \rightarrow buckwheat. Over 3500 fungal isolates were collected from more than 1000 samples of overwintered grains. These cultures belong to more than 40 genera with 200 species. 61 isolates were \rightarrow Fusarium poae (Peck) Wollenw. and 57 were \rightarrow Fusarium sporotrichioides Sherb. These molds produced high amounts of \rightarrow trichothecenes (e.g. \rightarrow T-2 toxin, \rightarrow diacetoxyscirpenol, \rightarrow HT-2 toxin, \rightarrow nivalenol etc.). Optimal toxin production of both Fusarium species occurred at 6-12 °C. Alteration of freezing and thawing temperatures caused maximal toxicity in culture.

Large numbers of people had to consume these overwintered, moldy and mycotoxin-contaminated grains (\rightarrow mycotoxins) because the shortage of manpower due to the war made harvesting impossible at the proper time. 2 to 3 weeks after ingestion of the toxic grain - at least 2 kg - the disease usually developed. A large number of victims died within 6-8 weeks after consuming at least 6 kg. However, breast-fed babies less than one year old did not show any symptoms. It seems that the toxic principle was not secreted into \rightarrow human breast milk. Ingestion of the \rightarrow grains resulted in the following clinical symptoms: First stage: burning sensation caused by inflammation of the mouth and fore-gut, emesis, diarrhoea, abdominal pain, no increase of body temperature, decrease of leukocyte counts ($\leq 2000 \text{ cells / mm}^3$); duration: 3 to 9 days. Second stage: disorder of bone marrow functions, pronounced and progressive \rightarrow aleukia; duration 3-4 weeks. Sudden onset of the third stage: petechial hemorraghes on head (face), trunk, and limbs, necrotic changes in the mouth, throat, and esophagus, bacterial infections (septic \rightarrow angina) occur, enlargement of the lymphatic glands, parenchymateous \rightarrow hepatitis resulting in \rightarrow jaundice (sometimes), further decrease of leukocyte counts ($\leq 100 \text{ cells}/\text{mm}^3$), significant decrease of erythrocyte and thromobocyte counts. Constriction of the glottis (strangulation) due to edemateous swelling caused the death of one-third of the vicitims. Among

death of one-third of the vicitims. Among survivors, intensity of toxicoses was decisive for the rate of recovery. In about 4 weeks the necrotic and hemorrhagic symptoms disappeared. However, two months or more were necessary for full recovery of the bone marrow function. Prophylaxis includes blood transfusion and administration of nucleic acid. Calcium preparations, vitamin C and K, and sulphonamide further contribute to avoid the development of severe symptoms. Based on the closely related if not identical syndromes it was concluded that \rightarrow moldy corn toxicosis and ATA have the same origin, viz. T-2 toxin and diacetoxyscirpenol, primarily produced by \rightarrow Fusarium sporotrichioides Sherb.

Alkaloids → ergot alkaloids

Almond paste → marzipan

Almonds (no specification) Contamination of maturing almonds with molds and \rightarrow mycotoxins may result from kernel damage due to the navel orange worm near the time of hull split. At this time the invading molds, frequently \rightarrow Aspergillus flavus Link, found moisture levels usually high enough to support growth and aflatoxin formation (\rightarrow aflatoxins).

The average probability of aflatoxin contamination in California almonds is one kernel in 26,500 unsorted in-shell nuts from the field.

Almonds may contain the following mycotoxins:

 \rightarrow aflatoxin B₁

incidence: 1/110*, conc.: 93 µg/kg, country: Finland, *imported incidence: 1/6*, conc.: 67 µg/kg, country:

Finland, *imported, bitter almonds incidence: 1/184*, conc.: $\leq 1 \ \mu g / kg$, country: Finland, *imported, sliced and

crushed

incidence: 7/198, conc. range: $< 5 \mu g/kg$ (6 samples), 12 $\mu g/kg$ (1 sa), country: Germany

incidence: 19*/23, conc. range: 39-4000 μg/kg, country: Germany, *moldy incidence: 87/261*, conc. range: < 5

 μ g/kg (44 samples), 11-189 μ g/kg (43 sa*), Ø conc.: 33 μ g/kg, country: Germany, *ground

incidence: 77/360*, Ø conc.: 28 μg/kg, country: Germany, *ground incidence: $2/4^*$, conc. range: $< 5 \mu g / kg$ (1 sample), 200 μ g/kg (1 sa), country: Germany, *sliced incidence: 43/907*, Ø conc.: 23 μg/kg, country: Germany, *sliced incidence: 15/19*, conc. range: 0.5-5 μ g/kg (14 samples), 6 μ g/kg (1 sa), country: UK, *ground \rightarrow aflatoxin B₂ incidence: 1/110*, conc.: 14 µg/kg, country: Finland, *imported incidence: $1/6^*$, conc.: 5 µg/kg, country: Finland, *imported, bitter almonds \rightarrow aflatoxins (no specification) incidence: 2*/78, conc. range: 5- > 25 μg/kg, country: Canada incidence: 47/327, conc. range: ?, country: Germany incidence: 77/360*, Ø conc.: 36 μg/kg, country: Germany, *ground incidence: 43/907*, Ø conc.: 35 μg/kg, country: Germany, *sliced incidence: 2/7*, conc. range: nc, country: UK, *ground incidence: 15/19*, conc. range: 0.5-5 μ g / kg (13 samples), 6-10 μ g / kg (2 sa) (with a maximum of 10 μ g/kg), country: UK, *shelled, ground incidence: 1*/4, conc.: 0.8 µg/kg, country: UK, *aflatoxin (AFB₁, AFB₂, AFG₁, AFG_2) incidence: 1/2*, conc.: nc, country: UK, *sugared incidence: 1/9*, conc. range: nc, country: UK, *unblanched incidence: 36/557, conc. range: 90 µg/kg, \emptyset conc.: 27 µg/kg, country: USA incidence: 28/345, conc. range: 2-94 μ g/kg, Ø conc.: 20 μ g/kg, country: USA incidence: 1/5, conc.: 10 μ g/kg, country: USA \rightarrow ochratoxin A incidence: 1/12, conc. range: 0.2-0.49 μg/kg, country: Germany \rightarrow nuts

Alperisins (Syn.: \rightarrow AAL-toxins) Altenuene (Abbr.: ALT) is a dibenzo- α pyrone derivative (2,3,4,4a-tetrahydro-2,3,7-trihydroxy-9-methoxy-4a-methyl-6H-dibenzo[b,d]pyran-6-one) produced by \rightarrow Alternaria spp. (\rightarrow mycotoxins) (see Figure Altenuene).

CHEMICAL DATA Empirical formula: $C_{12}H_{14}O_6$, molecular weight: 292

FUNGAL SOURCES

 \rightarrow Alternaria alternata (Fr.) Keissler, Alternaria citrii

NATURAL OCCURRENCE

 \rightarrow apples, \rightarrow olives, \rightarrow ragi, \rightarrow sorghum, \rightarrow tomatoes

Тохісіту

cytotoxic

LD₅₀ (ip): (50) 75-100 mg/kg bw mice. In chicks and rats no toxic effects occurred after feeding \rightarrow alternariol methyl ether, \rightarrow alternariol and ALT for 21 days at concentrations up to 24, 39, and 10 µg / g, respectively.

DETECTION see \rightarrow Alternaria mycotoxins

Alternaria (Syn.: Macrosporium) anamorphic \rightarrow Pleosporaceae, teleomorph Lewia (formerly Dematiaceae) Alternaria spp. are very common (airborne) fungi. Temperatures in the 18-22 °C range contribute to their growth. In addition, for substantial growth moisture contents of 28-34% in the substrate, i.e. water activities of $\geq \rightarrow a_w$ 0.84, are required.

This genus may be the principal fungus in \rightarrow wheat, \rightarrow barley, and \rightarrow sorghum, where in some years a nearly 100% infection has been recorded. Although *Alternaria* spp. cause rather limited damage (e.g. discoloration, black point of kernels) to cereal \rightarrow grains mycotoxin contamination may result from infection (\rightarrow mycotoxins). Seed moisture contents of $\approx 22\%$ due to heavy rainfall and high relative humidity at the time of harvest favor invasion.

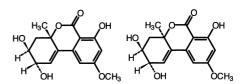
Besides grains, \rightarrow fruits and \rightarrow vegetables are also subject to *Alternaria* spp. infection both pre- and postharvest. Factors that promote invasion of these fungi are: surface physical damage, fruit imperfection, overripening, cold stress. Especially commodities held in cold storage may show significant spoilage because *Alternaria* spp. grow well at low temperatures.

About 70 secondary metabolites belonging to several chemical classes, e.g. anthraquinones, cyclic peptides, dibenzopyrones, lactones, perylenequinones, tetramic acids, are produced by this genus. From feeding studies it was estimated that 68% of the *Alternaria* strains are toxic.

Important Alternaria toxins which contaminate food are: alternariols (\rightarrow alternariol, \rightarrow alternariol methyl ether), and altenuens (\rightarrow altenuene, isoaltenuene), altertoxins (\rightarrow altertoxin I-III), \rightarrow tenuazonic acid. The mycotoxin stemphyltoxin III and Alternaria alternata f. sp. lycopersici toxins (\rightarrow AAL-toxins) are known from fungal cultures and infected plant material.

Alternaria alternata f. sp. lycopersici toxins \rightarrow AAL-toxins

Alternaria alternata (Fr.) Keissler (Syn.: A. longipes, A. tenuis) A. alternata is an extremely common saprophyte which contaminates various plants and foods.



Altenuene. Altenuene and isoaltenuene

Developing \rightarrow grains are infected by airborne spores of this fungus after anthesis when grain (\rightarrow wheat) moisture is as high as 70%. At high relative humidities this "black mold" causes a darkish discoloration ("black point") starting at the end of the grains (see Figure Alternaria alternata (Fr.) Keissler).

The relative production of alternariols and altertoxins is influenced by the water activity. Toxin production is best at water activities above $\rightarrow a_w 0.98$. Because formation of \rightarrow tenuazonic acid seems to be strain dependent a correlation between the relative toxicity of *Alternaria* infected grains and the degree of fungal invasion is doubtful.

Tobacco leaves are commonly invaded by A. alternata (A. longipes). Therefore, it has been suggested, that pulmonary exposures to \rightarrow Alternaria mycotoxins might be involved in lung diseases. However, no Alternaria mycotoxins have been found in infected tobacco leaves.

A. alternata may produce the following \rightarrow mycotoxins:

→ alternariol, → alternariol methyl ester, → altenuene, → altertoxins I-III, tenuazonic acid. In addition, *A. alternata* f. sp. *lycopersici* produces → fumonisins (FB₁, FB₂, FB₃).

Alternaria mycotoxins are produced by many *Alternaria* strains in relatively large amounts usually at the pre-harvest stage of the crop. \rightarrow Tenuazonic acid, \rightarrow alternariol methyl ether and \rightarrow alternariol occur most frequently while the occurrence of \rightarrow altenuene, isoaltenuene and \rightarrow altertoxin I-III has been reported occasionally.

Alternaria toxins are divided into 3 main structural classes: dibenzo- α -pyrones (i.e. alternariol, altenuene, altenuisol, altenusin, and dehydroaltenusin), tetramic acids (i.e. tenuazonic acid), and the altertoxins (i.e. altertoxin I-III).

NATURAL OCCURRENCE

- \rightarrow apples, \rightarrow barley, \rightarrow mandarin fruits,
- \rightarrow melon, \rightarrow oats, \rightarrow olives, \rightarrow pecans,
- \rightarrow pepper, \rightarrow ragi, \rightarrow rye, \rightarrow sorghum,

 \rightarrow sunflower seeds, \rightarrow tomatoes, \rightarrow triticale, \rightarrow wheat

Quite high levels of alternariol, alternariol methyl ether and tenuazonic acid occurred in \rightarrow apples and \rightarrow tomatoes as well as \rightarrow tomato paste. In \rightarrow wheat flour alternariol, alternariol methyl ether, altertoxin I, and tenuazonic acid could be detected after storage at 20 °C for 28 days.

Тохісіту

In comparison to the altertoxins, alternariol methyl ether is only slightly \rightarrow mutagenic but the possibility of synergistic toxic effects is high. Among the *Alternaria* toxins tenuazonic acid is probably the most acutely toxic. The involvement of *Alternaria* toxins in

The involvement of Alternaria toxins in human and animal health disorders is under discussion. It was suggested that A. alternata (formerly A. longipes) as a common pathogen in tobacco might be involved in lung disease of man due to the exposure to its mycotoxins. However, up to now no Alternaria toxins have been detected in this crop. Cereal samples (\rightarrow cereals) from farms with suspected mycotoxicosis showed a higher frequency of Alternaria mycotoxin contamination than brands from farms with healthy ani-



Alternaria alternata (Fr.) Keissler

mals. In addition, the contamination of fodder and feed with toxic *Alternaria* spp. could be responsible for the death of rabbits and poultry.

Although contamination of \rightarrow food and feed supplies with *Alternaria* mycotoxins in the developed countries seems to be low, these levels may cause chronic rather than acute disease. Synergistic effects of cooccurring mycotoxins should be taken into account.

Detection

HPLC & MS, LC

The detection and analysis of Alternaria mycotoxins often interferes with other commonly occurring \rightarrow mycotoxins (e.g. alternariol methyl ether $/ \rightarrow$ zearalenone, alternariol methyl ether and alternariol / \rightarrow aflatoxins). Although both Alternaria mycotoxins exhibit sky-blue fluorescence, it is brighter under short-wave than longwave ultraviolet light.

Possible Mycotoxicosis

Alternaria mycotoxins, especially tenuazonic acid, may responsible for the mycotoxicoses \rightarrow Onyalai.

FURTHER COMMENTS

25 °C and a_w 0.98 were the optimum conditions for the production of the three *Alternaria* mycotoxins alternariol, alternariol methyl ether, and alternaric acid.

Alternaria tenuissima (Kunze ex Pers.)

Wilts may produce the following

- \rightarrow mycotoxins:
- \rightarrow alternariol, \rightarrow alternariol methyl ester,
- → tenuazonic acid.

Alternariol (Abbr.: AOH) is a dibenzo- α -pyrone derivative (3,7,9-trihydroxy-1methyl-6H-dibenzo[b,d]pyran-6-one) produced by \rightarrow Alternaria spp. (\rightarrow mycotoxins) (see Figure Alternariol).

CHEMICAL DATA Empirical formula: $C_{14}H_{10}O_5$, molecular weight: 258 FUNGAL SOURCES

→ Alternaria alternata (Fr.) Keissler, A. cucumerina, A. dauci, A. kikuchiana, A. solani

NATURAL OCCURRENCE

→ apples, → barley, → mandarin fruits, → oats, → pecans, → pepper, → rye, → sorghum, → sunflower seeds, → tomatoes, → triticale, → wheat

TOXICITY

cytotoxic, fetotoxic, \rightarrow teratogenic Dosage (ip): 200 mg/kg bw mice (3 of 10 mice died)

AOH possesses a very weak acute toxicity. A synergistic effect between AOH and \rightarrow alternariol methyl ether could be shown.

DETECTION GC, HPLC, TLC

Possible Mycotoxicosis AOH may be involved in the "Fescue Foot Syndrome" (cattle).

Alternariol methyl ether (Abbr.: AME) is a dibenzo- α -pyrone derivative (\rightarrow mycotoxins) produced by \rightarrow Alternaria spp. (see Figure Alternariol methyl ether).

CHEMICAL DATA Empirical formula: $C_{15}H_{12}O_5$, molecular weight: 272

FUNGAL SOURCES

 \rightarrow Alternaria alternata (Fr.) Keissler, A. cucumerina, A. dauci, A. kikuchiana, A. solani

NATURAL OCCURRENCE

 \rightarrow apples, \rightarrow barley, \rightarrow mandarin fruits,

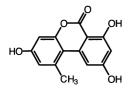
 \rightarrow melon, \rightarrow oats, \rightarrow olives, \rightarrow pecans,

 \rightarrow pepper, \rightarrow ragi, \rightarrow rye, \rightarrow sorghum,

 \rightarrow sunflower seeds, \rightarrow tomatoes, \rightarrow triticale, \rightarrow wheat

TOXICITY

necrotic (viscera), fetotoxic, \rightarrow teratogenic, \rightarrow mutagenic, and carcinogenic (?) AME possesses a very weak acute toxicity.



Alternariol

A synergistic effect between AME and \rightarrow alternariol could be shown. Dosage (ip): 400 mg AME / kg bw mice (1 of 10 mice died)

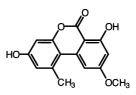
Detection

GC, HPLC, TLC

Thin-layer chromatograms and fluorescens of AME and zearalenone are similar, misidentification is possible.

Altertoxin I-III (Abbr.: ATX I, ATX II, ATX III) are 4,9-dihydroxyperylene-3,10quinons (ATX I = 1,2,11,12,12a,12b-hexahydro-1,4,9,12a-tetrahydroxy-3,10-perylenedione; ATX II = 7a,8a,8b,8c,9,10-hexahydro-1,6,8c-trihydroxy-perylo[1,2-b]oxirene-7,11-dione; ATX III = 1a,1b,5a,6a,6b,10a-hexahydro-4,9-dihydroxy-perylo[1,2-b:7,8-b']bisoxirene-5,10-dione) produced by \rightarrow Alternaria spp. (\rightarrow mycotoxins). Although altertoxins were isolated in 1973 their correct molecular structure was not elucidated until 1986 (see Figure Altertoxin I-III). Since the altertoxins possess a high toxicity their significance in food may be comparable to that of \rightarrow tenuazonic acid. The altertoxins are mainly responsible for the mutagenic activity of \rightarrow Alternaria mycotoxins. Compared to the other Alternaria mycotoxins the altertoxins are usually produced in small quantities by \rightarrow Alternaria alternata (Fr.) Keissler. This means a somewhat ameliorated risks for consumers.

CHEMICAL DATA Empirical formula: $C_{20}H_{16}O_6$, molecular weight: 352 (ATX I)



Alternariol methyl ether

Empirical formula: $C_{20}H_{14}O_6$, molecular weight: 350 (ATX II)

Empirical formula: $C_{20}H_{12}O_6$, molecular weight: 348 (ATX III)

FUNGAL SOURCES

Altertoxin I, II & III = A. alternata, A. mali, altertoxin I additionally A. tenuissima

NATURAL OCCURRENCE

 \rightarrow altertoxin I occurs in \rightarrow apples, \rightarrow sorghum

TOXICITY

cytotoxic, \rightarrow mutagenic

The altertoxins are very weak acute acting toxins, with an LD_{50} of 150 mg/kg bw mice. ATX-I and ATX-II were lethal to mice at the dose of 200 mg/kg bw. Treated animals showed inactivity, subendocarcial and subarachnoid hemorrhages, and blood in the cerebral ventricles. The mutagenic activity of ATX-III is approximately one tenth of that of \rightarrow aflatoxin B₁. ATX-I and ATX-II possessed a lower mutagenicity.

Detection

see \rightarrow Alternaria mycotoxins

Ammi (*Trachyspermum ammi* (Linn.) Sprague)

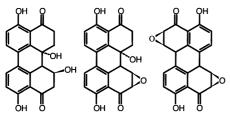
may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B_1

incidence: 1/7, conc.: 60 μg/kg, country: India

 \rightarrow aflatoxin B₂

incidence: 1/7, conc.: 34 µg/kg, country: India

 \rightarrow aflatoxin G₁



Altertoxin I-III

incidence: 1/7, conc.: 32 μg/kg, country: India

Ammoniation process Ammoniation greatly altered the biological activity of \rightarrow aflatoxin B₁ (\rightarrow aflatoxins) (450-fold decrease in mutagenicity).

Anemia A below average number of ery-throcytes.

Angina Any disease characterized by attacks of choking or suffocation.

Anguidine -	→ diacetoxyscirpenol
Anorexia Lo	oss of appetite
Antimycin –	→ citrinin
Apiospora –	→ Lasiosphaeriaceae
Aplastic aleuk kia	ia \rightarrow Alimentary toxic aleu-
Apple beveraging \rightarrow mycot	ges may contain the follow- oxins:

ing → mycotoxins: → patulin incidence: 29/66, conc. range: 5-54 µg/l, country: Sweden

Apple butter may contain the following \rightarrow mycotoxins: \rightarrow patulin incidence: 1/1, conc.: 1390 µg/kg, country: Finland **Apple flavor** may contain the following → mycotoxins: → patulin

incidence: 3/14, conc. range: 6-1770 μ g/kg, Ø conc.: 607 μ g/kg, country: Finland

Apple jam The ready solubility of \rightarrow patulin in water and the microchannels present in jams facilitate diffusion of this mycotoxin in this substrate. In domestic consumption, often only the top moldy layer of a mold-contaminated jar is discarded which is not sufficient in the case of patulin contamination. Apple jam (apple butter) may contain the following \rightarrow mycotoxins: patulin incidence: 1/1, conc.: 1390 µg/kg, country: Finland

Apple juice In commercial practise unsound, \rightarrow Penicillium expansum Link infected \rightarrow apples may partly be used for juice production. However, their portion is limited by causing off-flavors in the juice. Furthermore, the inclusion of infected apples will result in \rightarrow patulin contamination. Substantial toxin reduction (90%) is achieved by simple trimming of moldy apple tissues. During juice processing patulin content is reduced by only about 20%, mainly in the concentration step (vacuum distillation at 35-40 °C) (see Table Apple juice). Mycotoxin concentration in the juice is a good quality indicator for soundness of fruits used in the process.

Although commercially processed apple juices may be contaminated by patulin, levels are usually below 100 μ g/l. Juice directly made from fresh apples contained higher levels of patulin than juice prepared from concentrate. Thermal processing is not sufficient to insure a patulin free juice. Heat treatment for 10 or 20 min at 80 °C did not cause any destruction of patulin, 10s at 90 °C

(HTST) caused an almost 20% reduction. Only little decrease in patulin content could be observed after storage for up 3-4 weeks at 22 °C (10% after 2 weeks). However, addition of \rightarrow ascorbic acid, filtration or agitation with charcoal and fermentation to apple cider are highly effective in reducing patulin levels down to zero. Apple juice may contain the following \rightarrow mycotoxins: patulin incidence: 140/241*, conc. range: 5-50 μ g / kg (69 samples), 51- \leq 1130 μ g / kg, (71 sa) country: Australia, *apple and mixed \rightarrow fruit juices incidence: 1/30, conc.: 17 μ g/kg, country: Brazil incidence: 1/1, conc.: 1000 µg/l, country: Canada incidence: 5/11, conc. range: 20-120 µg/l, country: Canada incidence: 45/72, conc. range: $\leq 115 \,\mu g / l$, \emptyset conc.: 56.5 μ g/kg, country: Canada incidence: 28/61, conc. range: 20-17,700 μg/l, country: Canada incidence: 10/51, conc. range: 5-72 µg/l, country: Finland incidence: 8/20*, conc. range: \leq 65 µg/l, country: Finland, *home-made incidence: 9/13, conc. range: 100-300 μg/l, country: France incidence: 41/66, conc. range: 2-50 µg/l, country: Germany incidence: 4/17, conc. range: > 5.0-42.5 µg / l, country: Germany incidence: 7/36, conc. range: 20-300 μ g/l, country: Germany incidence: 5/10*, conc. range: 60-50,000 μg/l, country: Germany, *moldy incidence: 445/609, conc. range: ≤ 20 μ g/l (286 samples), $\leq 100 \mu$ g/l (122 sa), \leq 400 µg / l (37 sa) country: Germany incidence: 1/33, conc.: 52 μ g/kg, country: Germany incidence: 3/20, conc. range: 106-216 μ g/l, country: New Zealand incidence: nc/140, conc. range: < 1-220 μg / l, country: Norway

incidence: 80/165, conc. range: 20-253 μ g / l, Ø conc.: 30 μ g / l, country: Poland incidence: 82/100, conc. range: 0.5-170 μ g / l, Ø conc.: 13.8 μ g / kg, country: Spain incidence: 40/49, conc. range: \leq 70 µg/l, country: Sweden incidence: 29/66, conc. range: 2.5-27 μ g/l, country: Sweden incidence: 19/42, conc. range: 5-50 µg/l, country: Switzerland incidence: 20/21, conc. range: 5-20 µg/l $(5 \text{ samples}), > 20-50 \ \mu g/l \ (13 \ sa), > 50$ μ g/l (2 sa), country: Turkey incidence: $1/2^*$, conc.: 5-10 µg/l, country: UK, *long life incidence: $1/1^*$, conc.: 56 µg/l, country: UK, *organic incidence: 8/13*, conc. range: 5-10 µg/l (5 samples), 16-30 μ g/kg (3 sa), country: UK, *regular incidence: 24/45, conc. range: 1-56 µg/l, country: UK incidence: 14/20, conc. range: 1-38 µg/l, country: UK incidence: 23/40, conc. range: \approx 10-350 $\mu g/l$, Ø conc.: 51 $\mu g/l$, country: USA incidence: 8/13, conc. range: 44-309 µg/l, country: USA incidence: 5/5, conc. range: 244-3993 μ g/l, Ø conc.: 1902 μ g/l, country: USA incidence: 50/136, conc. range: 40-440 μ g/l, country: USA incidence: 9/40, conc. range: 20,000-45,000 μg/l, country: USA \rightarrow breakfast drinks , \rightarrow cider, fruit juice, \rightarrow grape juice, \rightarrow soft drinks Apple juice concentrate may contain the following \rightarrow mycotoxins: \rightarrow patulin incidence: 8/16, conc. range: 5-50 μ g/l (6 samples), > 50-646 μ g/l (2 sa), country: Australia

incidence: 15/71, conc. range: \leq 1450

μg/l, country: Finland

incidence: 27/27, conc. range: 55-610

μg/l, country: France

incidence: 79/165, Ø conc.: 30 μg/l, country: Poland incidence: 215/215, conc. range: 7-376 μg/l, country: Turkey

Apple products (no specification) may contain the following \rightarrow mycotoxins: \rightarrow patulin incidence: 7/105, conc. range: 11-50 µg/kg, country: Germany

Apples Patulin is the most important mycotoxin (\rightarrow mycotoxins) in apple and \rightarrow apple products. It is produced by the most common \rightarrow patulin-producing pathogen of apples (and pears), \rightarrow Penicillium expansum Link. Fruit infection is significantly favored by surface damage. Maximum patulin levels occurred 13-14 days after inoculation with P. expansum. Apples and pears are usually stored at low temperatures (-1 to 4 °C) and/or modified atmosphere (1 to 5% CO₂ and 1 to $3\% O_2$). These precautions delay senescence and suppress postharvest decay. However, even with these common postharvest technologies, P. expansum can grow and produce patulin. Fungal strain as well as the fruit cultivar are decisive for the patulin rate and the amounts produced.

The mycotoxin is primarily located in areas of the spoiled apple tissue although patulin contamination in visibly healthy fruit is known. Also, penetration up to

Apple juice. Relative decrease in patulin contamination in the course of apple juice processing (Kubacki 1986, modified)

rocessing step	losses (%)
asteurization I	3.4
epectinization	1.6
iltration	-
asteurization II	0.6
oncentration	18.4
acuum distillation (35–40 °C)	
otal losses	24
otal losses	

approximately 1 cm into the surrounding healthy tissue is possible. In consequence, removal of fungally decayed and surrounding tissues from apple prior to further processing significantly reduces patulin concentration in apple products. Concentration of patulin found in natural apple rots have been high as 136,000 μ g / kg of fruit. Apples may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/15*, conc.: 35 µg/kg, country: Germany, *moldy \rightarrow altenuene incidence: 5/8, conc. range: < 100-500 μ g / kg, Ø conc.: 100 μ g / kg, country: USA \rightarrow alternariol incidence: 1/20*, conc.: 160 µg/kg, country: Germany, *visibly moldy, different fruits incidence: 7/8, conc. range.: < 100-58,800 μ g / kg , Ø conc.: 7800 μ g / kg, country: USA \rightarrow alternariol methyl ether incidence: 1/20*, conc.: 250 µg/kg, country: Germany, *visibly moldy, different fruits incidence: 8/8, conc. range: < 100-2300 μ g / kg, Ø conc.: 1000 μ g / kg, country: USA \rightarrow altertoxin I incidence: 5/8, conc. range: nc, country: USA patulin incidence: 28/61*, conc. range: 20-17,700 μ g / apple, country: Canada, *rotted incidence: 5/12*, conc. range: 300-42,000 μ g/kg, country: Germany, *with rotten spots incidence: 1/16*, conc.: 2.6 µg/kg, country: Germany, *stewed incidence: 54/104, conc. range: 1-250 μg/kg, country: Spain → penicillic acid incidence: 1/6, conc.: nc, country: India \rightarrow tenuazonic acid

Apples

incidence 8/8, conc. range: 100-500 μ g/kg, country: USA \rightarrow fruits

Apricot seed paste → persipan

Arthrinium anamorphic \rightarrow Lasiosphaeriaceae, teleomorph \rightarrow Apiospora

Arthrinium sugarcane poisoning In China this disease is most prevalent from February to April. It caused 84 deaths in 847 cases between 1972 and 1988. A malfunction of the nervous system occurred after consumption of deteriorated sugarcane which may be contaminated by toxic fungal metabolites. The disease results in torsion spasms and may leave the victim permanently dislabled.

Besides \rightarrow Fusarium moniliforme Sheldon, \rightarrow Fusarium poae (Peck) Wollenw.,

 \rightarrow Penicillium aurantiogriseum Dierckx, and \rightarrow Cladosporium spp. certain

 \rightarrow Arthrinium species (A. sacchari, A. saccharicola, and A. phaeospermum) are discussed as the etiological fungi. From poisonous sugarcane samples 44% of the isolated fungi belonged to the latter genus.

Mice fed with Arthrinium culture material moved in circles and showed \rightarrow paralysis of limbs. Death occurred within 3 h. The only affected organ was the brain (encephaledema). A toxic fraction of Arthrinium was identified as $\rightarrow \beta$ nitropropionic acid. Juices of poisonous sugarcane contained this mycotoxin at levels as high as 1600 mg/kg. Such amount might be sufficient to cause human food poisoning outbreaks. In other studies F. moniliforme (\rightarrow fumonisins) and P. aurantiogriseum (various toxic factors) predominated on mildewed sugarcane. Feeding experiments with extracts of the spoiled sugarcane or both of the fungi caused nervous disorders and death.

Ascites Accumulation of serous fluid in the abdomen.

Ascomycota → Fungi

Ascorbic acid Addition of ascorbic acid to \rightarrow patulin-contaminated \rightarrow apple juice removed the toxin within 3 weeks.

Asparagus The vascular and epidermal tissue of asparagus is susceptible to by → Fusarium proliferatum (Matsushima) Nirenberg, alone or together with *F. oxy-sporum* f. sp. *asparagi* (→ Fusarium oxy-sporum Schlecht. emend. Snyd. & Hansen), causing crown and root rot. Fumonisin contamination (→ fumonisins) has been reported. Asparagus may contain the following → mycotoxins:

 \rightarrow fumonisin B₁

incidence: nc/25, conc. range: \leq 7400 µg*/ kg, 460 µg**/kg, country: Italy, *crown, **stem \rightarrow fumonisin B₂ incidence: nc/25, conc. range: \leq 830 µg*/kg, 60 µg**/kg, country: Italy,

*crown, **stem

Aspergillus anamorphic \rightarrow Trichocomaceae, teleomorphs \rightarrow Eurotium, \rightarrow Neosartorya, \rightarrow Emericella.

The genus is of ubiquitous distribution, but tends to predominate in tropical climates. Growth and metabolism of many species (e.g. \rightarrow Aspergillus versicolor (Vuill.) Tiraboshi, \rightarrow Aspergillus candidus Link) take place at low to very low water activities (\rightarrow Aspergillus restrictus G. Sm.). Therefore, *Aspergillus* spp. are the characteristic colonizers of stored products (see Figure *Aspergillus*). They are good indicators of previous storage conditions since each single species has its distinct minimum \rightarrow a_w value. *Aspergillus* spp. is further characterized by the production of numerous toxic metabolites (\rightarrow mycotoxins). Mycotoxin production starts at a_w levels between 0.80-0.83.

Some species are able to grow in the animal body (e.g. \rightarrow Aspergillus fumigatus Fres.) and may be associated with pathogenicity.

Important mycotoxin producers are: \rightarrow Aspergillus flavus Link, \rightarrow Aspergillus parasticus Speare, \rightarrow Aspergillus ochraceus group, *A. versicolor*. Important mycotoxins are: \rightarrow aflatoxins, \rightarrow citrinin, \rightarrow cyclopiazonic acid, \rightarrow ochratoxin A, \rightarrow sterigmatocystin

Aspergillus alutaceus var. alutaceus Berkely

& Curtis (formerly *A. ochraceus* K. Wilh.)

It is suggested that this is an important mycotoxin-producing fungus in cereals and the most important \rightarrow ochratoxin A producer within the genus *Aspergillus*. \rightarrow Peanuts and \rightarrow soybeans are the main substrates. The minimum $\rightarrow a_w$ of *A. alutaceus* for OTA and \rightarrow penicillic acid production is a_w 0.97-0.99 and a_w 0.85, respectively. Optimum OTA production occurs at $a_w > 0.97$. *A. alutaceus* may produce \rightarrow mycotoxins

A. *unitateus* may produce \rightarrow mycotoxins such as emodin, kojic acids (\rightarrow kojic acid), neoaspergillic acids, \rightarrow ochratoxins, \rightarrow penicillic acid, secalonic acid A (\rightarrow secalonic acids), \rightarrow viomellein, \rightarrow xanthomegnin.

Possible Mycotoxicosis \rightarrow Balkan endemic nephropathy

Aspergillus candidus Link is a frequent storage fungus. It is often found on \rightarrow cereals in silos where it contributes to the process of self-heating. In addition, it frequently occurs in cereals stored under a controlled atmosphere. A. candidus is the dominating fungus in flours (\rightarrow flour) and other \rightarrow cereal products (see Figure Aspergillus candidus Link).

A. candidus may produce \rightarrow mycotoxins such as candidulin, \rightarrow kojic acid, $\rightarrow \beta$ -

Aspergillus. Aspergillus flavus Link

nitropropionic acid, terphenyllins, xanthoascin.

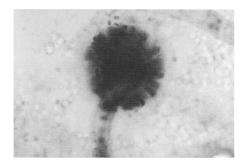
Aspergillus clavatus Desm. prefers the humid and high temperature conditions during malting and is therefore an important fungus in malt (see Figure Aspergillus clavatus Desm.). It causes the "malt worker's lung disease" (an allergic respiratory disease). Carbohydrates like glucose (better than sucrose, dextrin or lactose) may be essential for \rightarrow patulin production, but no formation will occur below 12 °C. The minimum water activity for patulin production of this fungus is \rightarrow a_w 0.99. During malting (\rightarrow malt) of \rightarrow barley and \rightarrow wheat. A. clavatus produces not only patulin but also cytochalasin E.

A. clavatus may produce \rightarrow mycotoxins such as ascladiol, cytochalasin E and "K", \rightarrow kojic acid (?), patulin, tryptoquivalins and tryptoquivalons (\rightarrow tremorgenic mycotoxins)

Possible Mycotoxicosis Ascladiol and patulin should be involved in mycotoxicosis.

Aspergillus flavus Link is a frequent mold in temperate climates. A. flavus has been isolated from various kinds of foodstuff but it is very common on cereal \rightarrow grains and their products as well as on \rightarrow spices (see Figure Aspergillus flavus

Aspergillus flavus Link



Aspergillus candidus Link

Link). Drought stress and insect damage favor the growth of this most toxic of all \rightarrow Aspergillus species prior to harvesting, especially in \rightarrow maize, \rightarrow peanuts, and cottonseed. However, healthy plant tissue may also be invaded.

FURTHER COMMENTS

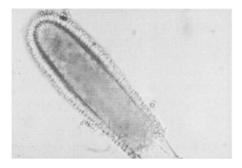
Although the toxicity of this fungus was described as early as 1910 by Kühl, it was not until 1960 that the \rightarrow aflatoxins could be identified in peanut meal as being highly toxic metabolites of *A. flavus*. *A. flavus* may accumulate AFB₁ and AFG₁ in conidia up to 84 mg/kg and 566 mg/kg, respectively. Sclerotia may contain 135 mg AFB₁/kg and 968 mg AFG₁/kg. Significant variation in total aflatoxin content in conidia and sclerotia within (intrafungal) and between strains could be established.

Production: Aflatoxin production starts with the formation of conidia and is usually proportional to the weight of the mycelium produced. Using a reduced amount of inoculum resulted in a three-to 12-fold increase of aflatoxin formation. Maximum rates occur until the period of intense sporulation (\approx sixth day) when the biomass production reaches its optimal value. Subsequently and similarly to the autolysis of the mycelium a rapid decrease in aflatoxin concentration due to degradation begins.

Toxigenic potential of this fungus is influenced by the geographical origin and by the substrate from which the fungus has been isolated. 60% of the isolates (n = 1390) coming from six different countries were toxigenic. It seems that strains (n = 427) isolated in tropical regions possess a higher degree of toxicity than strains from temperate origins (46% / 15%).

Various (competing) microorganisms like *Penicillium* spp., *Aspergillus niger*, or *Trichoderma viride* inhibited aflatoxin formation but their general effects on mycotoxin production are unpredictable. Sublethal concentrations of propionic acid may stimulate aflatoxin synthesis by *A. flavus*.

Substrate: A large number of toxigenic isolates has been found on U.S. American \rightarrow rice (94%) and peanuts (86%) as well as on groundnut kernels from Israel



Aspergillus clavatus Desm.



Aspergillus flavus Link

(71%). Generally A. flavus strains isolated from \rightarrow oil seeds - especially peanuts and peanut products - showed a higher proportion of aflatoxin producers than isolates contaminating \rightarrow cereals and their products. Instead of \rightarrow spices - ca. 30% of the isolated A. flavus strains were toxic - \rightarrow meat, cheeses (\rightarrow cheese), \rightarrow bread or \rightarrow pasta seem to be unsuitable substrates for toxigenic isolates. Carbon sources such as glucose, mannose, sucrose, and fructose as well as glyceraldehyde all favor aflatoxin production. The same is true for nitrogen sources like ammoniacal nitrogen, glutamic acid, or uric acid. In addition, yeast extract, peptone or certain amino acids (glycine, glutamate, proline) contribute to a higher aflatoxin formation. Vitamins of the B group, e.g. thiamine, cause stimulation of aflatoxin synthesis just as cadmium, iron, magnesium and zinc do. The wheat embryo, most probably because rich in diverse nutrients, allowed the production of much higher aflatoxin levels than the testa. Yields in the range from 100,000-2,000,000 µg aflatoxin / kg substrate, depending on the used culture, are known. The largest amounts have been reported for coconut flesh (8,000,000 $\mu g / kg$).

Moisture, temperature: A moisture content of 18.3-18.5% in cereal grains and 9-10% in seeds with a high oil content like \rightarrow nuts, \rightarrow copra, safflower and \rightarrow sunflower seeds may enable mycotoxin production. Below these values commodities are usually resistant to contamination. Temperatures between 24-28 °C are the optimum for \rightarrow aflatoxin B₁ production, 30 °C favor the formation of \rightarrow aflatoxin G₁. A constant temperature of 25 °C resulted in higher aflatoxin concentrations than fluctuating temperatures with a mean of 25 °C which are common in nature. 7.5 °C seems to be the lowest temperature enabling aflatoxin production whereas synthesis drops off sharply above 35 $^\circ\mathrm{C}.$

Atmosphere: Oxygen concentration as low as 1% in combination with 99% N_2 and 1% O_2 , 79% N_2 , and 20% CO_2 , respectively, allowed aflatoxin production. However, an atmosphere consisting of 1% O_2 , 19% N_2 , and 80% CO_2 prevented the synthesis of aflatoxin.

A. flavus may produce \rightarrow mycotoxins such as aflatoxins B₁, B₂, G₁, G₂ (although AFG₁ and AFG₂ are not generally produced), \rightarrow aflatrem, aspergillic acids, aspergillomarasmins, cyclopiazonic acids (\rightarrow cyclopiazonic acid), koji acids (\rightarrow kojic acid), maltoryzin, \rightarrow β -nitropropionic acid, paspalicin, paspalinine, \rightarrow sterigmatocystin.

Aspergillus fumigatus Fres. is an ubiquitous species which contaminates different kinds of food like → cereals (wet stored), → peanuts, → pecans, → tomatoes (see Figure Aspergillus fumigatus Fres.). It frequently occurs in cereals that are in advanced state of spoilage. Low oxygen tensions are tolerated. Due to its thermophilic nature, growth is adapted to high temperatures (\leq 55 °C). A. fumigatus may produce → mycotoxins such as fumagillin, fumigatins, fumiga-clavines, → fumitremorgins A & B, gliotoxin, → kojic acid (?),→ ochratoxin A, tryptoquivalins, verruculogen.

Aspergillus glaucus group \rightarrow Eurotium spp.

Aspergillus niger van Tieghem This fungus is a contaminant of various substrates of plant origin, e.g. \rightarrow cereals, but it usually does not predominate in spoiled cereal grain. \rightarrow Mycotoxins of the *A. niger* group (Section Nigri) have not yet been detected naturally in cereals. *A. niger* may produce mycotoxins such as aspergillins, \rightarrow kojic acid (?), malformins, naphthopyrones, \rightarrow ochratoxin A Aspergillus nomius Kurtzman et al. is not so common in \rightarrow foods as the very important mycotoxin producers \rightarrow Aspergillus flavus Link and \rightarrow Aspergillus parasiticus Speare.

A. nomius may produce \rightarrow mycotoxins such as \rightarrow aflatoxins B₁, B₂, G₁, G₂ (consistently produced), aspergillic acids, kojic acids (\rightarrow koji acid), nominine, \rightarrow tenuazonic acid.

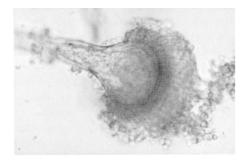
Aspergillus ochraceus group (= Section *Circumdati*)

included are \rightarrow Aspergillus alutaceus Berkley & Curtis, A. fresenii, A. ostianus, A. petrakii, A. quercinus, A. sclerotiorum. Fungi of this group / section produce \rightarrow ochratoxin A but they are considered to be rare on grain. These fungi do not produce ochratoxin A and \rightarrow penicillic acid below 12 °C.

Fungi of the *A. ochraceus* group may produce \rightarrow mycotoxins such as ochratoxin A, penicillic acid, \rightarrow secalonic acids, \rightarrow viomellein, vioxanthin, \rightarrow xanthomegnin.

Aspergillus ochraceus K. Wilh. (Syn.: → Aspergillus alutaceus Berkely & Curtis)

Aspergillus oryzae (Ahlburg) Cohn is often used for fermentating different kinds of foodstuff (e.g. koji, \rightarrow miso, soya sauce, saké alcohol) in Asian countries. Although it belongs to the *A. flavus* group and shows a high similarity with



Aspergillus fumigatus Fres.

the mycotoxin producer \rightarrow Aspergillus flavus Link \rightarrow aflatoxins are not synthesized. A. oryzae may produce \rightarrow mycotoxins such as aspergillomarasmin, \rightarrow cyclopiazonic acid, \rightarrow koji acid, maltoryzin, $\rightarrow \beta$ nitropropionic acid

Aspergillus parasiticus Speare In contrast to \rightarrow Aspergillus flavus Link A. parasiticus predominates in warmer climates (tropical and subtropical regions). Since this mold is most often associated with soil, \rightarrow peanuts are mainly infected by this Aspergillus species. Instead of this, A. flavus invasion is more common in \rightarrow maize. Nearly all strains of A. parasiticus are toxigenic while aflatoxin production is enhanced by the amino acid proline. \rightarrow Aflatoxin B₁ production starts at a water activity of 0.87. Maximum aflatoxin production on sterilized \rightarrow maize was observed at an $\rightarrow a_w$ of 0.90. A. parasiticus may produce \rightarrow mycotoxins such as \rightarrow aflatoxins B₁, B₂, G₁, G₂ (consistently produced), aspergillic acids, koji acids (\rightarrow kojic acid), $\rightarrow \beta$ -nitropropionic acid, \rightarrow sterigmatocystin

Aspergillus restrictus G. Sm. belongs to the important storage fungi in cereal \rightarrow grains. Besides \rightarrow Eurotium *halophilicum* it is the first growing fungus in \rightarrow cereals stored at moisture contents that are just a little too high for safe storage (\approx 14%). This slowly growing fungus does not cause any significant rise in grain temperature. The metabolic water of this primary colonizer enables the growth of mycotoxin-producing fungi like \rightarrow Aspergillus flavus Link. A. restrictus is often associated with storage insects such as the granary and the rice weevil which contribute to its distribution. A. restrictus may produce \rightarrow mycotoxins such as mitgilliin.

Aspergillus terreus Thom predominates in \rightarrow cereals stored under airtight condi-

Azotemia

tions (see Figure Aspergillus terreus Thom). Although it produces a wide range of \rightarrow mycotoxins it is not known whether they do naturally occur in cereals.

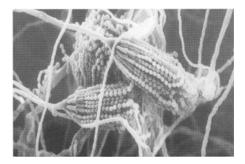
A. terreus may produce mycotoxins such as \rightarrow citreoviridin, \rightarrow citrinin, cytochalasin E (\rightarrow cytochalasins), flavipin?, gliotoxin, \rightarrow patulin, terreic acid, teritrem A, B, C, A', and B'.

Aspergillus versicolor (Vuill.) Tiraboshi has frequently been isolated from moldy seeds and their products and from oil seed products (\rightarrow oil seeds). Under certain environmental conditions, e.g. airtight storage, it may predominate (see Figure Aspergillus versicolor (Vuill.) Tiraboshi). A. versicolor is the most important producer of \rightarrow sterigmatocystin. Sterigmatocystin is produced in \rightarrow cheese ripened at 6 °C. Toxicoses which involve A. versicolor are probably due to sterigmatocystin and related metabolites. A. versicolor may produce \rightarrow mycotoxins such as aspertoxin, nidulotoxin, \rightarrow ochratoxin A, sterigmatocystins, versicolorins.

ATA \rightarrow Alimentary toxic aleukie

Ataxia Loss of muscle coordination

Atmosphere It seems that mycotoxin production (\rightarrow mycotoxins) is more sensitive to the concentration of atmospheric gases than fungal growth. In most cases mold development and mycotoxin formation is inhibited by low O₂ concentration (< 1%) and/or elevated levels of CO₂. High CO₂ levels appeared to be more effective in controlling fungal growth and mycotoxin formation than high N₂ and low O₂ concentrations. Since fungal growth has been reported in \rightarrow beer high levels of CO₂ may not be sufficient to prevent mold development and subsequent mycotoxin formation in all cases.

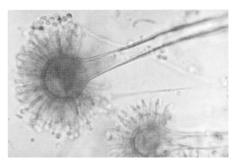


Aspergillus terreus Thom

Temperature and $\rightarrow a_w$ influence the tolerance to specific gas compositions.

The water activity (a_w) of a substrate a" is defined as the ratio of the water vapor pressure of the food substrate (p) to the vapor pressure of pure water (p_0) at the same temperature and pressure: $a_w = p/$ p_0 . The a_w expresses the moisture content of a substrate (e.g. food) as an index of water available for the growth of the microorganisms. Since each fungal species/strain has minimum water requirements for growth at a given temperature and on a distinct substrate the same is true for mycotoxin production (\rightarrow mycotoxins) of the different fungi (see Table a_w). Growth and mycotoxin production in different substrates are only comparable in terms of their a_w not their water contents.

Azotemia increase of nitrogen levels in the blood



Aspergillus versicolor (Vuill.) Tiraboshi

Azotemia

Mold	Mycotoxin	Minimum a _w - growth -	Minimum a _w - toxin production -
Aspergillus ochraceus	penicillic acid	0.76-0.83	0.80-0.88
A. flavus	aflatoxin	0.78-0.84	0.83-0.87
A. ochraceus	ochratoxin A	0.76-0.83	0.83-0.87
Penicillium verrucosum	ochratoxin A	0.81-0.83	0.83-0.90
P. griseofulvum	patulin	0.81-0.85	0.85-0.95
A. parasiticus	aflatoxin	0.78-0.82	0.87
P. aurantiogriseum	ochratoxin A	0.79-0.85	0.87-0.90
P. patulum	patulin	0.81-0.85	0.95
P. aurantiogriseum	penicillic acid	0.79-0.85	0.97-0.99
P. expansum	patulin	0.82-0.85	0.99
A. clavatus	patulin	0.85	0.99

aw. Minimum aw for growth and mycotoxin production by selected molds

B

Baby cereals (no specification) may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 30 products analysed, \emptyset conc.: 43 µg/kg, country: Canada ergocristine (\rightarrow ergot alkaloids) incidence: 1/1*, conc.: 0.4 µg/kg, country: Canada, *mixed \rightarrow cereals

Baby food may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: nc*, conc. range: 4-66 µg/kg, country: France, * \rightarrow meat/vegetable preparation (\rightarrow vegetables) incidence: nc*, conc. range: 2-26 µg/kg, country: France, *carrot preparation \rightarrow deoxynivalenol incidence: 14/39, conc. range: tr-90 µg/kg, country: USA \rightarrow ochratoxin A incidence: 2/34, conc. range: \leq 0.2 µg/kg, country: Germany

Bacon may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 2*/10, conc. range: 1000-5000 μ g / kg, Ø conc.: 3000 μ g / kg, country: Germany, *moldy \rightarrow ochratoxin A incidence: 39/206*, conc. range: 37-200 μ g / kg, country: Yugoslavia, *total of smoked \rightarrow meat products \rightarrow sausages

Bakery products (no specification) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins (no specification) incidence: 2/8, conc. range: 1-14 µg/kg, country: UK \rightarrow citrinin incidence: 1/2, conc.: < 150 µg/kg, country: UK → ochratoxin A incidence: 3/8, conc. range: 0-80 μ g / kg, country: UK → cereals

Baking Since most \rightarrow mycotoxins are heat-stable no significant reduction in their concentration will occur during baking (see Table Influence of baking and other heat processing on stability of trichothecenes). For details see each single mycotoxin.

 \rightarrow cereals

Balkan endemic nephropathy (Abbr.: BEN) A chronic kidney disease which was first described in the 1950s in the Vratza District (Bulgaria). Now the disease mainly occurs in different rural areas of Bulgaria, Romania, and Yugoslavia located within the Danube Basin. Some 20,000 people mainly of the rural population should be affected. In endemic areas up to 12% of the inhabitants suffer from this disease. Mortality rates of up to 40% have been reported. Resulting from prolonged exposure to a causal agent the affected individuals are almost exclusively between (30)-35 and (50)-55 years old. More females than males were affected.

While the onset of the disease remains unnoticed, in a progressed stage impairment of the kidney function becomes obvious. Severe \rightarrow nephropathy often accompanied by urinary tract tumors are the major symptons. Tumor rate of this rarely occurring kind of cancer is nearly 100 times higher in the endemic area compared to the non-endemic ones. Histologically, this \rightarrow renal disease is characterized by tubular degeneration and interstitial \rightarrow fibrosis. In the more superficial part of the \rightarrow cortex hyalization of the glomeruli appeared. The size of the kidneys affected is greatly reduced. A prominent and early indication of the

disease is the impairment of tubular function. The insidious onset of a normocytic, normochronic \rightarrow anemia, \rightarrow azotemia, and persistent \rightarrow proteinuria as well as renal failure are the leading clinical symptoms. This incurable disease is treated only symptomatically which includes hemodialysis. Within 5 to 10 years BEN progresses slowly up to death. The etiology of BEN is still unknown. A positive correlation between heavy rainfall (late summer and autumn), possibly favoring growth and toxigenicity of fungi in endemic areas, and the number of people who died of nephropathy during the succeeding 2 years could be established. Therefore, involvement of fungi (e.g. \rightarrow Penicillium and \rightarrow Aspergillus) and certain of their \rightarrow mycotoxins is discussed. The mammalian carcinogen \rightarrow ochratoxin A might be the main causal agent, especially because similarities with the \rightarrow mycotoxic porcine nephropathy due to this mycotoxin in Scandinavia do exist. This is corroborated by the fact that different foodstuffs, e.g. cereal \rightarrow grains, produced in the endemic areas of Yugoslavia showed a significantly higher OTA contamination than products from nonendemic areas. Consequently, residues of OTA could be detected more frequently and at higher concentrations in the blood of inhabitants as well as in the \rightarrow pig kidneys, \rightarrow pig liver, and \rightarrow pig blood of endemic areas. Besides \rightarrow citrinin $(\rightarrow$ nephrotoxin), which is also found in greater proportion and greater degree in the staple foods of affected families, a novel Penicillium mycotoxin, possibly a glycopeptide (molecular weight ca. 1500), might be involved in the etioloy of BEN. However, although substantial OTA contamination of food- and feedstuff has been reported no mycotoxic porcine nephropathy occurred in the endemic areas of BEN. Furthermore, data about OTA contamination of foodstuffs and the blood of BEN patients are not sufficient

to prove a quantitative relationship between the degree of OTA exposure and the severity of human nephropathy. The involvement of heavy metals and / or viruses is also under discussion showing that the significance of mycotoxins in BEN still remains unresolved.

Bananas may contain the following \rightarrow mycotoxins: \rightarrow zearalenone Incidence: 1/1, conc.: 17 µg/kg, country: India \rightarrow fruits

Barley In years of moist weather, seeds of barley may be relatively heavily invaded by more than a dozen species of \rightarrow Fusarium spp. during time of maturation. Severe invasion will result in a reddish discoloration of a portion of the kernels. In consequence, seeds fail to develop and shrivel, or the partly developed kernel deteriorates. This disease is called "scab" or "blight". Mycotoxin contamination of the kernels is possible. Barley may contain the following \rightarrow mycotoxins: → 3-acetyldeoxynivalenol incidence: 24/40, conc. range: \leq 350 $\mu g/kg$, Ø conc.: 40 $\mu g/kg$, country: Canada incidence: 1/6, conc.: $< 200 \mu g / kg$, country: Finland incidence: 10/30, conc. range: 24-96 μ g/kg, Ø conc.: 46 μ g/kg, country: Finland incidence: 1/30, conc.: 7 µg/kg, country: Korea → 15-acetyldeoxynivalenol incidence: 24/77, conc. range: ≤ 400 μ g / kg, Ø conc.: 40 μ g / kg, country: Canada incidence: 39/40, conc. range: 1240 μ g/kg, Ø conc.: 210 μ g/kg, country: Canada

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 \rightarrow aflatoxin B₁ incidence: 3/376, conc. range: < 10-2000 μ g / kg, country: UK aflatoxin $B_1 \& \rightarrow$ aflatoxin B_2 incidence: $1^{*}/94^{**}$, conc.: $\approx 10 \, \mu g / kg$, country: Japan, *moldy, **barley and pressed barley aflatoxin (no specification) incidence: 13/37, conc. range: 1-5 µg/kg (9 samples), $5-20 \,\mu g / kg$ (3 sa), $31 \,\mu g / kg$ (1 sa), country: Germany incidence: 12/137*, conc. range: 2-20 μ g / kg (10 samples), > 20 μ g / kg (2 sa), country: Uruguay, *and malt \rightarrow alternariol incidence: 1/179, conc.: 15 µg/kg, country: Germany incidence: 1/3, conc.: 116 µg/kg, country: Poland \rightarrow alternariol methyl ether incidence: 12/254, conc. range: 4-25 μ g / kg, Ø conc.: 10.2 μ g / kg, country: Germany incidence: 1/3, conc.: 58 μ g/kg, country: Poland \rightarrow citrinin incidence: 4/269, conc. range: 30-480 μg/kg, country: Sweden incidence: 4/4*, conc. range: tr-1600 μg / kg, country: UK, *moldy → deoxynivalenol incidence: 18/20, \emptyset conc.: 237 µg/kg, country: Argentina incidence: 16/90, conc. range: 7-1670 μ g / kg, Ø conc.: 270 μ g / kg, country: Canada incidence: 77/77, conc. range: 100-15,100 μ g / kg, Ø conc.: 2650 μ g / kg, country: Canada incidence: 40/40, conc. range: 30-15,790 μ g / kg, Ø conc.: 2690 μ g / kg, country: Canada incidence: 4/4, conc. range: 21-164 µg/kg, \emptyset conc.: 83 µg/kg, country: China incidence: 3/5, conc. range: 10-20 µg/kg, country: Denmark incidence: 1/nc, conc.: 1000 µg/kg, country: Denmark

incidence: 3/6, conc. range: 1-6300 µg / kg, country: Finland incidence: 27/30, conc. range: 10-202 μ g / kg, Ø conc.: 78 μ g / kg, country: Finland incidence: 3/3*, conc. range: 27-46 µg/kg, \emptyset conc.: 38 µg/kg, country: Finland, *imported from Canada, Sweden incidence: 1/4, conc.: 10 μ g/kg, country: France incidence: 2/10, Ø conc.: 190 µg/kg, country: Germany incidence: 6/25, conc. range: 150-1000 μg/kg, country: Germany incidence: 2/13, Ø conc.: 190 µg/kg, country: Germany incidence: 6/123, conc. range: 10-100 μg/kg, country: Germany incidence: 1/1, conc.: 46 µg/kg, country: Germany incidence: 31/46*, conc. range: 20-2140 μg/kg, country: Germany, *visibly damaged incidence: 6/7*, conc. range: 34-440 μ g/kg, Ø conc.: 137.8 μ g/kg, country: Germany, *organic produce incidence: 2/5, \emptyset conc.: 195 µg/kg, country: Italy incidence: 95/101, conc. range: 50-49,600 μ g/kg, Ø conc.: 1506 μ g/kg, country: Japan incidence: 5/5, \emptyset conc.: 249 µg/kg, country: Japan incidence: 18/20, conc. range: 0-4600 μ g / kg, Ø conc.: 900 μ g / kg, country: Japan incidence: 12/14, conc. range: 50-7840 μ g/kg, Ø conc.: 2010 μ g/kg, country. Japan incidence: 2/14, conc. range: 100-510 μ g/kg Ø conc.: 305 μ g/kg, country: Japan incidence: 12/12, conc. range: 90-1700 μ g/kg, Ø conc.: 580 μ g/kg, country: Japan incidence: $8/12^*$, Ø conc: 176 µg/kg, country: Japan, *naked

Barley

incidence: 6 products* analysed, conc. range: 27-85 µg/kg, country: Japan, *parched incidence: $1/1^*$, conc.: 48 µg/kg, country: Japan, *pearled incidence: 10/14*, conc. range: 3-50 µg/kg, country: Japan, *pressed incidence: 31/31, conc. range: \leq 900 μ g/kg, Ø conc.: 124 μ g/kg, country: Korea incidence: 20/30, conc. range: 5-361 μ g/kg, Ø conc.: 106 μ g/kg, country: Korea incidence: 26/28*, conc. range: 4-508 μ g/kg, Ø conc.: 117 μ g/kg, country: Korea, *unpolished incidence: 24/27*, conc. range: 38-645 μ g / kg, Ø conc.: 213 μ g / kg, country: Korea, *naked incidence: 9/10*, conc. range: 29-677 μ g/kg, Ø conc.: 263 μ g/kg, country: Korea, *husked incidence: 3/11, conc. range: 168-506 μ g/kg, Ø conc.: 297 μ g/kg, country: Korea incidence: 26/44, conc. range: ≤ 1000 μg/kg, country: New Zealand incidence: 1/6, Ø conc.: 390 µg/kg, country: Poland incidence: 5/8, conc. range: 10-81 µg/kg, \emptyset conc.: 42 µg/kg, country: Scotland incidence: 2/14, conc. range: 80-160 μ g / kg, Ø conc. 120 μ g / kg, country: Sweden incidence: 4/32, conc. range: 60-150 μ g / kg, Ø conc.: 90 μ g / kg, country: Sweden incidence: 1/6, conc.: 50 µg/kg, country: Sweden incidence: 7/52, conc. range: 50-200 μ g / kg, Ø conc.: 90 μ g / kg, country: Sweden incidence: 4/4, Ø conc.: 83 µg/kg, country: Taiwan incidence: 2/5, conc. range: 10-30 µg/kg, \emptyset conc.: 20 µg/kg, country: The Netherlands

incidence: 5/6, conc. range: 4-152 µg/kg, \emptyset conc.: 58 µg/kg, country: The Netherlands incidence: 3/3*, conc. range: 56-147 μ g / kg, Ø conc.: 110 μ g / kg, country: The Netherlands, *pearled incidence: 28/92, conc. range: 20-500 μg/kg, country: UK incidence: 22/49, conc. range: 20-100 μ g/kg, country: UK incidence: 108/147, conc. range: 500-26,000 μg/kg, Ø conc.: 4200 μg/kg, country: USA incidence: nc/204, conc. range: tr-22,000 μ g / kg, country: USA incidence: 2/3, Ø conc.: 19 µg/kg, country: Yemen \rightarrow diacetoxyscirpenol incidence: 16/134, conc. range: 200-17,000 µg/kg, country: Germany incidence: 1/86, conc.: 100 µg/kg, country: USSR 3,15-diacetyldeoxynivalenol incidence: 25/40, conc. range: \leq 400 μ g/kg, Ø conc.: 60 μ g/kg, country: Canada \rightarrow HT-2 toxin incidence: 37/94, conc. range: 100-10,000 μg/kg, country: Germany incidence: 5/24*, conc. range: 210-370 μ g / kg, Ø conc.: 230 μ g / kg, country: Poland, *spring barley → neosolaniol incidence: 1/86, conc.: 100 µg/kg, country: USSR \rightarrow nivalenol incidence: 15/20, \emptyset conc.: 25 µg/kg, country: Argentina incidence: 1/6, conc.: $< 100 \mu g / kg$, country: Finland incidence: 4/30, conc. range: 38-59 μ g / kg, Ø conc.: 46 μ g / kg, country: Finland incidence: 1/3, conc.: 44 μ g/kg, country: Germany incidence: 1/13, conc.: 40 µg/kg, country: Germany

incidence: 1/5, conc.: 23 μ g/kg, country: Italy incidence: 95/101, conc. range: 23-22,900 μ g / kg, Ø conc.: 1020 μ g / kg, country: Japan incidence: 5/7, conc. range: 90-640 µg/kg, country: Japan incidence: 5/5, \emptyset conc.: 708 μ g/kg, country: Japan incidence: 12/12, conc. range: 60-1500 μ g/kg, Ø conc.: 480 μ g/kg, country: Japan incidence: 18/20, conc. range: 0-2900 μ g / kg, Ø conc.: 700 μ g / kg, country: Japan incidence: 12/14, conc. range: 0-2320 μ g / kg, Ø conc.: 430 μ g / kg, country: Japan incidence: 2/14, conc. range: 0-270 μ g/kg, Ø conc.: 140 μ g/kg, country: Japan incidence: 12/12*, Ø conc.: 342 µg/kg, country: Japan, *naked incidence: 1/1*, conc. range: 220 µg/kg, country: Japan, *pearled incidence: 13/14*, conc. range: 8-380 μg/kg, country: Japan, *pressed incidence: 31/31, conc. range: \leq 1100 μ g/kg, Ø conc.: 489 μ g/kg, country: Korea incidence: 28/30, conc. range: 40-2038 μ g / kg, Ø conc.: 390 μ g / kg, country: Korea incidence: 28/28*, conc. range: 17-3002 μ g / kg, Ø conc.: 546 μ g / kg, country: Korea, *unpolished incidence: 27/27*, conc. range: 85-4569 μ g/kg, Ø conc.: 1110 μ g/kg, country: Korea, *naked incidence: 10/10*, conc. range: 114-1546 μ g / kg, Ø conc.: 742 μ g / kg, country: Korea, *husked incidence: 2/11, conc. range: 189-324 μ g/kg, Ø conc.: 257 μ g/kg, country: Korea incidence: 1/4, conc.: 21 μ g/kg, country: Nepal

incidence: 33/44, conc. range: \leq 530 µg/kg, country: New Zealand incidence: 3/6, Ø conc.: 78 µg/kg, country: Poland incidence: 3/8, conc. range: 7-1140 µg/kg, \emptyset conc.: 391 µg/kg, country: Scotland incidence: 4/4, conc. range: 290-976 μ g / kg, Ø conc.: 634 μ g / kg, country: Taiwan incidence: 4/6, conc. range: 30-145 μ g / kg, Ø conc.: 85 μ g / kg, country: The Netherlands incidence: 3/3*, conc. range: 17-39 μ g / kg, Ø conc.: 27 μ g / kg, country: The Netherlands, *pearled incidence: 2/3, Ø conc.: 13 µg/kg, country: Yemen \rightarrow ochratoxin A incidence: 3/27, conc. range: 5-1000 μ g / kg, country: Austria incidence: 1/48, conc.: 3800 µg/kg, country: Czechoslovakia incidence: 11/41*, conc. range: 0.05-4.9 μ g / kg (8 samples), 5-14 μ g / kg (3 sa), country: Denmark, *conventional incidence: 6/20*, conc. range: 0.05-4.9 μ g/kg (4 samples), 5-13 μ g/kg (2 sa), country: Denmark, *ecological incidence: 17/17, conc. range: 9-27,520 μg/kg, country: Denmark incidence: 3/50, conc. range: 9-189 μ g/kg, Ø conc.: 80.7 μ g/kg, country: Denmark incidence: 10/68, conc. range: 0.1-206 μ g / kg, Ø conc.: 58.8 μ g / kg, country: Germany incidence: 11/165, conc. range: 100-1800 μ g/kg, Ø conc.: 634 μ g/kg, country: Poland incidence: 54/616, conc. range: 5-1200 μ g / kg, country: Poland incidence: 17*/269, conc. range: 2-20 μ g / kg, country: Sweden, *14 only traces incidence: 21/21*, conc. range: 0.1-8652 μ g / kg, country: Tunesia, *and derived foods incidence: 9/52, conc. range: \leq 4.9-45 μ g / kg, country: UK

Barley

incidence: 10/50, conc. range: \leq 4.9-13.7 μ g/kg, country: UK incidence: 7/150, conc. range: $\leq 4.9-33.4$ μ g / kg, country: UK incidence: 51/376, conc. range: < 25-5000 μg/kg, country: UK incidence: 4/4*, conc. range: 75-11,000 μ g/kg, Ø conc.: 3038 μ g/kg, country: UK, *moldy incidence: 18/127, conc. range: tr-38 μg/kg, country: USA incidence: 11/848, conc. range: < 15-116 µg/kg, country: USA incidence: 23/182, conc. range: 10-29 μg/kg, country: USA incidence: 22*/159, conc. range: \leq 29 μ g/kg, country: USA, *11 contained less than 10 µg/kg incidence: 23/164, conc. range: < 10-29 μg/kg, country: USA incidence: 11/103, conc. range: 0.03-17,000 μg/kg, Ø conc.: 6.87 μg/kg, country: USA incidence: 1/48, conc.: 3800 µg/kg, country: USSR incidence: 8/64*, conc. range: 14-27 μ g / kg, country: Yugoslavia, *area with endemic nephropathy incidence: 1/23, conc.: 5 µg/kg, country: Yugoslavia → penicillic acid incidence: 1/165, conc.: 800 µg/kg, country: Poland → sterigmatocystin incidence: 2/4*, conc. range: traces, country: UK, *moldy \rightarrow T-2 toxin incidence: 2/6, conc. range: 50-600 μ g/kg, Ø conc.: 325 μ g/kg, country: Finland incidence: 1/18, conc.: 160 µg/kg, country: Finland incidence: 1/?, conc.: 20 µg / kg, country: France incidence: 1/78, conc.: 90 µg/kg, country: Germany

incidence: 2/12, conc. range: 105-165 μ g/kg, Ø conc.: 135 μ g/kg, country: Germany incidence: 6/132, conc. range: 200-14,000 μg/kg, country: Germany incidence: 5/49, conc. range: 2-22 µg/kg, country: Norway incidence: 12/24*, conc. range: 20-2400 μ g/kg, Ø conc.: 450 μ g/kg, country: Poland, *spring barley incidence: 1/86, conc.: 600 µg/kg, country: USSR T-2 tetraol incidence: 2/24*, conc. range: 10-210 μ g/kg, Ø conc.: 110 μ g/kg, country: Poland, *spring barley T-2 triol incidence: 5/94, conc. range: 100-300 μg/kg, country: Germany \rightarrow viomellein incidence: 3/4*, conc. range: tr-600 μg/kg, country: UK, *moldy vioxanthin incidence: 3/4*, conc. range: 10-90 μ g/kg, Ø conc.: 40 μ g/kg, country: UK, *moldy \rightarrow xanthomegnin incidence: 3/4*, conc. range: tr-450 μ g / kg, country: UK, *moldy \rightarrow zearalenone incidence: 13/20, \emptyset conc.: 5 µg/kg, country: Argentina incidence: 2/30, conc. range: 21-30 μ g/kg, Ø conc.: 26 μ g/kg, country: Finland incidence: 2/10, Ø conc.: 16 µg/kg, country: Germany incidence: 3/3, Ø conc.: $3 \mu g / kg$, country: Germany incidence: 21/92, conc. range: 1-1730 μ g/kg, Ø conc.: 60 μ g/kg, country: Germany incidence: 5/13, Ø conc.: 10 µg/kg, country: Germany incidence: 5/40, conc. range: 10-20 μg/kg, country: Germany

incidence: 24/46*, conc. range: \leq 320 μ g / kg, Ø conc.: 24 μ g / kg, country: Germany, *damaged kernels incidence: 1/5, conc.: 56 µg/kg, country: Italy incidence: 3/5, Ø conc.: 9 µg/kg, country: Japan incidence: $10/12^*$, Ø conc.: 4 µg/kg, country: Japan,*naked incidence: $1/1^*$, conc.: 4 µg/kg, country: Japan, *pearled incidence: 1/13*, conc.: 6 µg/kg, country: Japan, *pressed incidence: $1/3^*$, conc.: 6 µg/kg, country: Japan, *polished incidence: 29/31, \emptyset conc.: 24 μ g/kg, country: Korea incidence: 21/28*, conc. range: 3-1581 μ g / kg, Ø conc.: 110 μ g / kg, country: Korea, *unpolished incidence: 6/27*, conc. range: 40-1081, \emptyset conc.: 579 µg/kg, country: Korea, *naked incidence: 4/10*, conc. range: 183-1416 μ g / kg, Ø conc.: 552 μ g / kg, country: Korea, *husked incidence: 4/4, Ø conc.: 18 µg/kg, country: Nepal incidence: 15/85, conc. range: \leq 170 μg/kg, country: New Zealand incidence: 3/584, conc. range: 200-1200 μ g / kg, Ø conc.: 700 μ g / kg, country: Poland incidence: 8/8, conc. range: 3-33 µg/kg, \emptyset conc.: 10 µg/kg, country: Scotland incidence: 2/4, conc. range: 17-22 µg/kg, \emptyset conc.: 19 µg/kg, country: Taiwan incidence: 6/6, conc. range: 4-9 µg/kg, \emptyset conc.: 7 µg/kg, country: The Netherlands incidence: 3/3*, conc. range: 16-29 μ g / kg, Ø conc.: 22 μ g / kg, country: The Netherlands, *pearled incidence: 20/137*, conc. range: 100-200 μ g / kg (12 samples), > 200 μ g / kg (8 sa), country: Uruguay, *and malt incidence: 3/3, Ø conc.: 43 µg/kg, country: Yemen

 \rightarrow cereals

Barley flour may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 1/1*, conc.: 32 µg/kg, country: Germany, *whole meal incidence: 3/6, conc. range: 8-39 µg/kg, country: Japan \rightarrow nivalenol incidence: 6/6, conc. range: 13-41 µg/kg, country: Japan \rightarrow zearalenone incidence: 6/6, conc. range: 1-4 µg/kg, country: Japan \rightarrow flour

Barley grits may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 1/1, conc.: 36 µg/kg, country: Germany \rightarrow maize grits, \rightarrow rye grits, \rightarrow wheat grits

Barley malt may contain the following \rightarrow mycotoxins: aflatoxin (no specification) (\rightarrow aflatoxins) incidence: 9/42, conc. range: 1-5 µg/kg (7 samples), $5- \le 14 \ \mu g / kg$ (2 sa), country: Germany \rightarrow deoxynivalenol incidence: 1/8, conc.: 70 µg/kg, country: Canada incidence: 4/5, Ø conc.: $40 \mu g/kg$, country: Canada incidence: 4/4, conc. range: 22-5840 μ g / kg, Ø conc.: 1595 μ g / kg, country: Korea incidence: 13/42, conc. range: 10-20 μ g/kg (5 samples), 20-100 μ g/kg (8 sa), country: UK \rightarrow nivalenol incidence: 4/4, conc. range: 122-436 μ g / kg, Ø conc.: 243 μ g / kg, country: Korea → ochratoxin A

Barley malt

incidence: 3/50, conc. range: 9-189 μ g/kg, country: Denmark \rightarrow zearalenone incidence: 4/4, conc. range: 2-36 μ g/kg, \emptyset conc.: 19 μ g/kg, country: Korea \rightarrow beer, \rightarrow malt

Bay leaf may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/1, conc.: 5.1 µg/kg, country: The Netherlands \rightarrow spices

Bean hull poisoning Sporadically dried bean hulls (\rightarrow beans) and plants used as feed caused a high incidence of poisoning of horses in Japan (Hokkaido). The affected animals showed circular movement, motor irritation, cronic musclespasm and \rightarrow tachycardia. \rightarrow Jaundice, \rightarrow hemorrhage of nerve cells and \rightarrow renal tubular epithelium also occurred. One of the isolated \rightarrow Fusarium strains (F. sporotrichioides M-1-1) produced \rightarrow T-2 toxin, \rightarrow neosolaniol and related \rightarrow trichothecences. Because the purified T-2 toxin did not cause such nervous symptoms, it was concluded that some other toxin(s) may be involved.

Bean jam may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/24*, conc.: 0.8 µg/kg, country: Japan, *red incidence: 5/41*, conc. range: 0.1-0.7 µg/kg, country: Japan, *white

Beans (no specification)

Although we do not have much information about mycotoxin contamination of beans in comparison to cereal \rightarrow grains several \rightarrow mycotoxins such as \rightarrow aflatoxins, \rightarrow deoxynivalenol, \rightarrow diacetoxyscirpenol, \rightarrow fumonisin B₁, \rightarrow ochratoxin A,

 \rightarrow penicillic acid, \rightarrow T2-toxin and \rightarrow zearalenone have been detected. Cooking of beans (*Phaseolus vulgaris* L.) naturally contaminated with OTA did not result in a total destruction of this mycotoxin. About 16-60% of the original toxin was detected after processing. Losses amounted to between 80 and 95% of the original OTA levels if soaking was included. Extending the autoclaving period did not significantly contribute to a reduction of the OTA level. In the case of Faba beans (Vicia faba L.), cooking under pressure (115 °C, 2 h) caused a 20% reduction in OTA concentration. Losses in the range of 20-76% occurred in artificially contaminated beans after cooking for 20 min at 121 °C in an autoclave. Beans may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/10*, conc.: 39 µg/kg, country: Brazil, *Carioquinha, dried incidence: $1/3^*$, conc.: 52 µg/kg, country: Brazil, *Mulatinho, dried incidence: 1/6*, conc.: 1.7 µg/kg, country: Egypt, *Vicia faba L. incidence: 4/381*, conc. range: 1.4-254 μ g/kg, country: Japan, *for bean jam incidence: 5/99*, conc. range: 1.5-12.0 μ g/kg, Ø conc.: 4.5 μ g/kg, country: Japan, *butter incidence: 2/2, conc. range: 6.3-26.9 μ g/kg, Ø conc.: 16.6 μ g/kg, country: Japan, *red incidence: 1/231*, conc.: 1.4 µg/kg, country: Japan, *small red incidence: 3/37*, conc. range: 1.3-11 μ g / kg, Ø conc.: 4.56 μ g / kg, country: Japan, *Saltani-Saltapaya incidence: 10*/322, Ø conc.: 213 µg/kg, country: Thailand, *total: Ø conc.: 1620 μ g / kg AFB₁, AFB₂, AFG₁, AFG₂ incidence: 7*/140**, Ø conc.: 16 μg/kg, country: Thailand, *total Ø conc.: 112 μg/kg AFB₁, AFB₂, AFG₁, AFG₂, **mung

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 \rightarrow aflatoxin B₂ incidence: 1/6, conc.: 1.5 µg/kg, country: Egypt, *Vicia faba L. incidence: 4/381*, conc. range: 1.2-8.5 μ g / kg, country: Japan, *for bean jam incidence: 5/99, conc. range: 0.5-2.2 µg/ kg, Ø conc.: 1.24 μ g/kg, country: Japan, *butter incidence: 2/2, conc. range: 3.5-6.9 μ g/kg, Ø conc.: 5.2 μ g/kg, country: Japan, *red incidence: 1/231, conc.: 0.4 µg/kg, country: Japan, *small red incidence: 3/37*, conc. range: 0.4-3 μ g / kg, Ø conc.: 1.33 μ g / kg, country: Japan, *Saltani-Saltapaya \rightarrow aflatoxin G₁ incidence: 1/10*, conc.: 21 µg/kg, country: Brazil, *Carioquinha, dried incidence: 1/3*, conc.: 31 µg/kg, country: Brazil, *Mulatinho, dried \rightarrow aflatoxin G₂ incidence: $1/10^*$, conc.: 4 μ g/kg, country: Brazil, *Carioquinha, dried incidence: 1/3*, conc.: 8 µg/kg, country: Brazil, *Mulatinho, dried aflatoxin (no specification) incidence: 18/20*, conc. range: \leq 222 μ g / kg, Ø conc.: 63 μ g / kg, country: Philippines, *brown kidney incidence: $4/7^*$, conc. range: ≤ 118 μ g / kg, Ø conc.: 58 μ g / kg, country: Philippines, *lima (*Phaseolus lunatus* L.) incidence: 29/33*, conc. range: ≤ 46 $\mu g / kg$, Ø conc.: 13 $\mu g / kg$, country: Philippines, *mung (Phaseolus aureus Roxb.) aflatoxins (no specification) incidence: 9/68, conc. range: nc, country: Hong Kong incidence: 1/12*, conc. range: nc, country: Hong Kong, *mung incidence: 11/610, conc. range: 2-36 μg/kg, country: Japan incidence: 2/29, conc. range: > 30 - \leq 86 μg/kg, country: Philippines incidence: 46*/64, conc. range: 1-100 μ g / kg (30 samples), 100-1000 μ g / kg (11 sa), > 1000 μ g/kg (5 sa), country: Uganda

15 samples contained AFB₁ (\emptyset conc.: 500 µg/kg), 42 AFB₂, 11 AFG₁, 1 AFG₂ \rightarrow deoxynivalenol incidence: 2/3, conc. range: 3100-6500 μ g / kg, Ø conc.: 4800 μ g / kg, country: Taiwan, *grey and/ or pink discoloration, navy bean (Phaseolus vulgaris L.) → diacetoxyscirpenol incidence: 2*/3, conc. range: 3300-9200 μ g / kg, Ø conc.: 6250 μ g / kg, country: Taiwan, *grey and/ or pink discoloration, navy bean (Phaseolus vulgaris L.) \rightarrow fumonisin B₁ incidence: 2*/3, conc. range: 500-1100 μ g/kg, Ø conc.: 800 μ g/kg, country: Taiwan, *grey and/ or pink discoloration, navy bean (Phaseolus vulgaris L.) \rightarrow ochratoxin A incidence: 1/10*, conc.: 94 µg/kg, country: Brazil, *Carioquinha, dried incidence: 1/3*, conc.: 160 µg/kg, country: Brazil, *Rosinha, dried incidence: 4/24*, conc. range: 25-27 μ g / kg, country: Bulgaria, *area with endemic nephropathy incidence: 2/28*, conc. range: 25-50 μ g / kg, country: Bulgaria, *area with endemic nephropathy incidence: 75/157*, conc. range: 0.05-260 μ g / kg, country: Bulgaria, *area with endemic nephropathy incidence: 31/113, conc. range: 0.2-285 μg/kg, country: Bulgaria incidence: 1/84*, conc.: 20 µg/kg, country: Canada, *beans & peas incidence: 3/4*, conc. range: 40-2000 μ g/kg, Ø conc.: 766 μ g/kg, country: Canada, *Phaseolus vulgaris L., dried incidence: 1/3*, conc.: 7 μg/kg, country: Egypt, *horse bean incidence: 2/8, conc. range: 25-50 µg/kg, \emptyset conc.: 37.5 µg/kg, country: USA, *red incidence: 6/71, conc. range: 10-442 μg/kg, country: Sweden incidence: 9/127*, conc. range: 10-442 μg/kg, country: Sweden, *brown kidney incidence: 2/8*, conc. range; 35 µg/kg, country: USA, *black turtle soup

Beans

incidence: 6/8*, conc. range: 20-100 μ g/kg, Ø conc.: 40 μ g/kg, country: USA, *great northern incidence: 6/8*, conc. range: 25-100 $\mu g / kg$, \emptyset conc.: 50 $\mu g / kg$, country: USA, *navy (Phaseolus vulgaris L.) incidence: 3/8*, conc. range: 50- >1000 μg/kg, country: USA, *pinto incidence: 2/8*, conc. range: 25 µg/kg, \emptyset conc. 25 μ g/kg, country: USA, *pinto \rightarrow penicillic acid incidence: 3/8 conc. range: 300-500 μ g / kg, Ø conc.: 550 μ g / kg, country: USA, *red incidence: 5/20, conc. range: 11-179 $\mu g / kg$, Ø conc.: 82 $\mu g / kg$, country: USA \rightarrow T2-toxin incidence: 2*/3, conc. range: 5500-13,500 μ g/kg, Ø conc.: 9500 μ g/kg, country: Taiwan, *grey and/ or pink discoloration, navy bean (Phaseolus vulgaris L.) \rightarrow zearalenone incidence: 1/150, conc.: 160 µg/kg, country: Yugoslavia \rightarrow cabbage, \rightarrow cowpeas, \rightarrow lentils, \rightarrow peas, \rightarrow pigeon peas, \rightarrow soybeans,

 \rightarrow vegetables

Beauvericin (Abbr.: BEA) This cyclic lactone trimer (\rightarrow mycotoxins) with an alternating sequence at three *N*-methyl L-phenylalanyl and three D- α -hydroxyisoa-leryl residues is synthesized by several \rightarrow Fusarium species (see Figure Beauvericin).

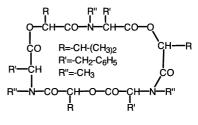
CHEMICAL DATA Empirical formula: $C_{45}H_{47}N_3O_9$, molecular weight: 725

Fungal Sources

Fusarium semitectum, F. subglutinans

NATURAL OCCURRENCE

It was first detected in Polish \rightarrow maize but natural contamination of Italian and US maize has also been reported.



Beauvericin

TOXICITY

BEA is highly toxic for insects, as well as for murine and human cells, in which it induces apoptosis.

Detection HPLC

Beefburger Detection of \rightarrow aflatoxins in beefburgers results from the use of mycotoxin-contaminated \rightarrow spices and / or the incorporation of aflatoxin producers. Beefburgers may contain the following \rightarrow mycotoxins:

 \rightarrow aflatoxin B₁

incidence: 5/25, \emptyset conc.: 8 µg / kg, country: Egypt \rightarrow meat

Beer Beer may be contaminated with different \rightarrow mycotoxins.

Some special beer types, e.g. strong beer, might be important contributors to the daily \rightarrow ochratoxin A intake. The degree of contamination depends upon the quality of the initial \rightarrow barley, storage conditions (< 16% mc barley malt) and the fate of OTA during malting (\rightarrow malt), brewing, and fermentation.

Although OTA does not survive malting, even if very highly contaminated barley malt is used, the addition of this mycotoxin to the mash or before fermentation (simulating use of adjuncts which are usually added at the beginning of the brewing process) revealed a possible transmission into the beer. In consequence, adjuncts such as \rightarrow maize products (e.g. maize syrup and grits),

 \rightarrow rice, barley and \rightarrow wheat would be expected to be the source of any OTA contamination in commercial beer. The contamination of commercial beers with \rightarrow fumonisins may be attributed to the use of contaminated barley but more likely, corn-based brewing adjuncts. Often a portion of barley is replaced by adjuncts, e.g. \rightarrow maize grits, which are most frequently used by the brewing factories. It could be shown that fumonisins (FB₁, FB₂) are appreciably stable towards the yeast fermentation of maize and \rightarrow wort. Calculated from Canadian and imported beers the daily intake estimates for adult beer drinkers were in the range of 0.010-0.049 μ g fumonisin B₁ / kg bw, assuming an average intake of 643 ml beer / day. Consumption of 950 ml beer / day (heavy beer drinkers) containing the maximum fumonisin level found will result in a daily FB₁ intake in the range of 0.24-0.60 µg/kg bw. \rightarrow Deoxynivalenol and \rightarrow nivalenol may occur in beer since the process for cleaning \rightarrow grains (e.g. barley) destined for brewing is ineffecient. Contaminated kernels with near-normal size and weight cannot be selectively removed. In addition, if moldy grains such as maize, especially in developing countries, are used for local beer production, humans may be exposed to elevated levels of a number of \rightarrow Fusarium mycotoxins via consumption.

Beer may contain the following mycotoxins:

→ aflatoxin B₁ incidence: 3/3, conc. range: 0.006-0.059 μ g / kg, country: Mexico → aflatoxin B₁ & → aflatoxin B₂ incidence: 16/304*, conc. range: 1.0-2.5 μ g / l, country: Kenya, *local, home brewed → deoxynivalenol incidence: 28/50*, conc. range: 0.3-50.3 μ g / l, Ø conc.: 5.7 μ g / kg, country: Canada, *28 Canadian and 22 imported beers

incidence: 1/49, conc.: 20 µg/l, country: France incidence: 18/18, conc. range: \leq 9 µg/l, Ø conc.: $\approx 5 \,\mu g/l$, country: Germany incidence: 8/38, conc. range: 1-5.3 µg/l, \emptyset conc.: 3.1 µg/l, country: Korea incidence: 2/5*, conc. range: 3.8-10 µg/l, \emptyset conc.: 6.9 μ g/l, country: Korea, *imported beers \rightarrow diacetoxyscirpenol incidence: 5/49, conc.: ca. 10-35 µg/l, country: France \rightarrow fumonisin B₁ incidence: 11/41, conc. range: 0.42-59 μ g/l, country: Canada incidence: 20/46, conc. range: 0.2-58.2 μ g/l, country: Canada \rightarrow fumonisin B₂ incidence: nc, conc. range: 0.45-9.2 µg/l, country: Canada incidence: 7/46, conc. range: 0.4-11.5 μ g/l, country: Canada \rightarrow fumonisins incidence: 14/32, conc. range: 4.8-85.5 μ g/kg, Ø conc.: 25.8 μ g/l, country: Spain → nivalenol incidence: 3/50*, conc. range: 0.1-0.84 μ g/kg, Ø conc.: 0.4 μ g/l, country: Canada, *28 Canadian and 22 imported beers incidence: 32/38, conc. range: < 1-20 μ g/l, country: Korea incidence. $3/5^*$, conc. range: 1.3-2.5 µg/l, \emptyset conc.: 1.93 µg/l, country: Korea, *imported beers \rightarrow ochratoxin A incidence: 26/41*, conc. range: tr-0.2 μ g/l, Ø conc.: 0.061 μ g/l, country: Canada, *Canadian and imported beers (11)incidence: 21/21, conc. range: ≤ 0.16 μ g/kg, Ø conc.: 0.049 μ g/l, country: Denmark incidence: 5/66, conc. range: $\leq 0.1 \ \mu g / l$, country: Germany

incidence: 80/160, conc. range: \leq 0.49 µg/l, country: Germany

Beer

incidence: 6/11, conc. range: 0.03-0.08 μg/kg, country: Germany incidence: 4/37, conc. range: 5-110 µg/l, Ø conc.: 56.3 µg/l, country: France incidence: 14/16, conc. range: 0.002-0.052 μ g/l, Ø conc.: 0.014 μ g/l, country: UK \rightarrow T-2 toxin incidence: 3/49, conc. range: ca. 10-42 μg/l, country: France \rightarrow zearalenone incidence: 1/49, conc.: 100 μ g/l, country: France incidence: 17/140, conc. range: 300-2000 μ g/l, country: Lesotho incidence: 2/23*, conc. range: 8000-53,000 µg/l, country: Swaziland, *and other fermented products incidence: 14-15/23, conc. range: < 90-4600 μg/l, Ø conc.: 1410-1500 μg/l, country: Zambia barley, \rightarrow barley malt, cereals, maize, \rightarrow sorghum, wheat

Beer (draft) may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 2/2*, conc. range: 6.3-8.8 µg/l, \emptyset conc.: 7.55 µg/l, country: Korea, *imported beers \rightarrow nivalenol incidence: 1/2*, conc.: 8.8 µg/l, country: Korea, *imported beers

Beer (light) may contain the following \rightarrow mycotoxins: \rightarrow nivalenol incidence: 3/3, conc. range: 24-38 µg/l, Ø conc.: 31.3 µg/l, country: Korea incidence: 3/3*, conc. range: 3.3-7 µg/l, Ø conc.: 4.6 µg/l, country: Korea, *imported beers

Beer (non-alcoholic) may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 2/2*, conc. range: 18-23 μ g/l, Ø conc.: 20.5 μ g/l, country: Korea, *imported beers \rightarrow nivalenol incidence: 1/1, conc.: 9 μ g/l, country: Korea

Beer (pale) may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A incidence: 1/28, conc. range: 0.3 µg/l, country: Germany incidence: 7/7, conc. range: 0.01-0.033 µg/l, country: Switzerland

Beer (strong) may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A incidence: 14/40, conc. $\leq 1.5 \ \mu g / l$, \emptyset conc. 0.28 $\mu g / l$, country: Germany incidence: 9/26, conc. range: 0.35-1.53 $\mu g / l$, \emptyset conc.: 1 $\mu g / l$, country: Germany incidence: 13/32, conc. range: 0.05-0.49 $\mu g / l$, country: Germany

Beer, barley may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 35/123, conc. range: \leq 478 µg/l, \emptyset conc.: 148 µg/l, country: Germany

Beer, burukutu is a Nigerian type of beverage made from guinea corn (*Sorghum* sp.) and \rightarrow millet (*Penissetum* sp.) while the malt is retained. In experimental studies it could be shown that there was a \rightarrow carry over of \rightarrow zearalenone into the finished product from 43-62%. Burukutu beer may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ & \rightarrow aflatoxin G₁ incidence: 2/2, conc. range: 253-262 µg/l, \emptyset conc.: 257.5 µg/l, country: Nigeria

Beer, joala The composition of joala beer varies with the proportions of the

ingredients, which are malted \rightarrow maize and / or \rightarrow sorghum, \rightarrow flour and hops and occasionally various \rightarrow fruits such as grapes and pineapples.

Joala may contain the following \rightarrow mycotoxins:

 \rightarrow zearalenone

incidence: 17/40, conc. range: 300-2000 μ g/l, country: Lesotho

Beer, millet may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 32/40*, conc. range: 1.7-138

μg/kg, Ø conc.: 64 μg/kg, country: Nigeria, *native

→ aflatoxin B₁ and other → aflatoxins incidence: 10/10, conc. range: 4- > 50 μ g/l, Ø conc.: 25 μ g/kg, country: Nigeria

Beer, opaque maize is a Zambian type of beverage brewed from \rightarrow maize, \rightarrow millet or red \rightarrow sorghum. A \rightarrow carry over of \rightarrow zearalenone in the range of 51.4% from starting zearalenone concentration in the finished product has been recorded in maize beer. Maize beer may contain the following \rightarrow mycotoxins: zearalenone incidence: nc/23, Ø conc.: 920 µg/l, country: Zambia

Beer, pito is a Nigerian type of beverage brewed from red guinea corn (*Sorghum* sp.) and \rightarrow millet (*Penissetum* sp.) or a mixture of both while the \rightarrow malt is filtered off.

Pito may contain the following \rightarrow mycotoxins:

 \rightarrow aflatoxin B₁ & \rightarrow aflatoxin G₁

incidence: 2/2, conc. range: 92-142 μg/l, Ø conc.: 117 μg/l, country: Nigeria → zearalenone incidence: 28/46, conc. range: 12.5-200 $\mu g/l$, Ø conc.: 81.8 $\mu g/l$, country: Nigeria

Beer, sorghum may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 8/150, conc. range: 0.05-0.13 $\mu g/l$, \emptyset conc.: 0.1 $\mu g/l$, country: South Africa

Beer, wheat may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 50/67, conc. range: \leq 569 µg/l, Ø conc.: 245 µg/l, country: Germany \rightarrow ochratoxin A incidence: 1/3, conc.: 0.3 µg/l, country: Germany

Bentonite A clay (montmorillonit) with adsorptive properties used for the removal of \rightarrow mycotoxins (e.g. \rightarrow aflatoxins, \rightarrow patulin) from \rightarrow milk, \rightarrow apple juice and other fluid products. \rightarrow decontamination

Bile duct Passages for conveyance of bile in and from the liver.

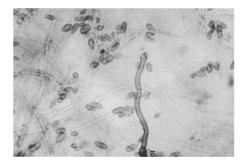
Bioassays This preliminary screening system enables toxicity test of extracts made from commodities which might show a mycotoxin contamination. Bacteria, yeasts, Tetrahymena pyriformis, Artemia salina as well as larvae of trout and other fish can be used for assaying \rightarrow mycotoxins while toxicity is expressed as a percentage of death in a given time. In addition, chick embryos and ducklings, as well as cell cultures (e.g. rat liver, baby hamster kidney, human epithelial), have also been used. However, lack of specificity due to other (toxic) substances coextracted with mycotoxins limits the application of bioassays.

Biscuits \rightarrow Ochratoxin A is partially (ca. 60%) destroyed or immobilized during biscuit making. This reduction in OTA concentration may be explained by the high dough temperature during \rightarrow baking, the low water content of these \rightarrow cereal products, and / or the presence of bicarbonate in the dough. Biscuits may contain the following \rightarrow mycotoxins: ochratoxin A incidence: 1/9, conc. range: 0.2-0.49 μg/kg, country: Germany incidence: 3/11*, conc. range: 0.1-1.49 μg/kg, country: Germany, *salted \rightarrow cereals, \rightarrow cookies

Black molds Molds like \rightarrow Alternaria spp., \rightarrow Cladosporium spp., *Epicoccum* spp. and *Helminthosporium* spp. grow on prematurely dead ears as a superficial dark mycelium and spore masses (see Figure Black molds). Spikelets with excreted honeydew are the preferred substrates. Among the black molds, especially species of the genus *Alternaria* belong to the important mycotoxin producers.

Blepharis edulis (medicinal seeds) may contain the following \rightarrow mycotoxins: aflatoxin B₁ incidence: nc/nc, conc. range: 10-1040

 μ g/kg, country: India



Black molds. Cladosporium herbarum

"Blind staggers" syndrome → Leucoencephalomalacia

Blue Castello cheese \rightarrow cheese, Blue Castello

Blue cheese \rightarrow cheese, Blue

Blue cheese dressing \rightarrow cheese dressing, blue

Blue Haverti cheese \rightarrow cheese, Blue Haverti;

Blueberries may contain the following \rightarrow mycotoxins: \rightarrow patulin incidence: 3/16, conc. range: 75-190 μ g/kg, country: Sweden incidence: 1/12, conc.: 21 μ g/kg, country: Sweden \rightarrow fruits

Bondakaledkai is an Indian peanut $(\rightarrow \text{ peanuts})$ based spiced snack which consists of whole seeds with an intact seed coat. Before deep-fat-frying the \rightarrow nuts are covered with salt, \rightarrow rice flour and a paste of red \rightarrow chilli powder. Aflatoxin contamination may be due to the use of uncleaned and unpicked whole seeds along with the seed coat. Bondakaledkai may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 22/54, conc. range: 3-1500 μg/kg, country: India \rightarrow aflatoxin B₂ incidence: 14/54, conc. range: 3-370 μg/kg, country: India \rightarrow congressbele, \rightarrow groundnut toffee **bovinocidin** $\rightarrow \beta$ -nitropropionic acid

Bran (no specifiaction) may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 14 products analysed, Ø conc.: 170 µg/kg, country: Canada \rightarrow ochratoxin A incidence: 1/41, conc.: 0.1 µg/kg, country: Germany incidence: 19/30, conc. range: 0.1-0.49 μ g / kg (12 samples), 0.5-1.49 μ g / kg (4 sa), 1.5-9.99 µg/kg (3 sa), country: Germany incidence: 9/84, \emptyset conc.: 6.8 μ g/kg, country: Germany incidence: 5/35, conc. range: $\leq 11 \ \mu g / kg$, \emptyset conc.: 4.5 μ g/kg, country: Italy incidence: 12/43, conc. range: \leq 4.9 µg/ kg, \emptyset conc.: 1.03 µg/kg, country: UK \rightarrow cereals, \rightarrow maize bran, \rightarrow milling, \rightarrow oat bran, \rightarrow rice bran, \rightarrow rye bran, \rightarrow wheat bran

Brazil nuts Brown and fluorescent kernels generally contain the main part of the \rightarrow aflatoxins. Since contaminated \rightarrow nuts are so obviously damaged, human consumption seems unlikely. Separation of aflatoxin positive nuts is based on the assumption that the moldy nuts are lighter than the good ones. The former are removed by an air blower and by means of gravity separation. Brazil nuts may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 33/302*, Ø conc.: 286 µg/kg, country: Germany, *in-shell incidence: 57/135, conc. range: $< 5 \ \mu g / kg$ (29 samples), 40-8000 µg/kg (28 sa) country: Germany incidence: 1/2, conc.: 3200 µg/kg, country: Germany incidence: 13/17, conc. range: < 5 µg/kg (12 samples), $5 \mu g / kg$ (1 sa), country: Germany incidence: 10/10*, conc. range: 8-47,000 μ g/kg, Ø conc.: 12,522 μ g/kg, country: Germany

incidence: 16/27*, conc. range: 3-4200 μ g / kg, Ø conc.: 500.2 μ g / kg, country: Norway *imported incidence: 5/23, conc. range: 0.5-5 µg/kg (4 samples), 33 μ g/kg (1 sa), country: UK \rightarrow aflatoxin B₂ incidence: 4/10*, conc. range: 0.6-883 μ g / kg, Ø conc.: 517.7 μ g / kg, country: Germany incidence: 16/27*, conc. range: tr-1600 μg/kg, country: Norway, *imported \rightarrow aflatoxin G₁ incidence: 9/10*, conc. range: 7-56,000 μ g / kg, Ø conc.: 18,457 μ g / kg, country: Germany incidence: 16/27*, conc. range: 2-3250 μ g / kg, Ø conc.: 478.2 μ g / kg, country: Norway *imported \rightarrow aflatoxin G₂ incidence: 3/10*, conc. range: 1.2-1000 μ g / kg, Ø conc.: 533.7 μ g / kg, country: Germany, * kernels visibly discolored incidence: 16/27*, conc. range: tr-600 μg/kg, country: Norway, *imported \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 31/69*, conc. range: 6-100 μg/kg (8 samples), 101-1000 μg/kg (7 sa), 1001-10,000 μg/kg (11 sa), > 10,000 μ g/kg (5 sa), country: Sweden, *imported; edible, possibly edible and inedible nuts aflatoxins (no specification) incidence: 62/234, conc. range: nc, country: Germany incidence: 33/302*, Ø conc.: 305 µg/kg, country: Germany incidence: 4/14*, conc. range: 2-129 μ g / kg, country: UK incidence: 6/38*, conc. range: nc, country: UK incidence: 5/23*, conc. range: 0.5-5 $\mu g \, / \, kg$ (3 samples), 6-10 $\mu g \, / \, kg$ (1 sa), 60 μ g/kg, country: UK *in-shell incidence: 4/18*, conc. range: 2-129 μ g/kg, country: UK

Brazil nuts

incidence: $6/12^*$, conc. range: ≤ 42 $\mu g / kg$, \emptyset conc.: 20 $\mu g / kg$, country: USA *shelled \rightarrow nuts

Bread (no specification) may be contaminated by different \rightarrow mycotoxins which are more or less stable during processing. If white \rightarrow flour is spiked with \rightarrow ochratoxin A no decomposition of the mycotoxin occurs after baking (220 °C, 25 min).

However, levels of \rightarrow aflatoxins in flour were significantly reduced during fermentation and \rightarrow baking as compared to that in the finished bread. Besides oxidation during kneading, especially fermentation and hydrothermal processes during cooking caused degradation of most of the \rightarrow aflatoxin B₁ (\approx 40-80%). Infection of bread with toxigenic isolates of Aspergillus flavus Link resulted in aflatoxin contamination several days later, although the wrapped bread restricted fungal growth due to lack of oxygen (see Figure Bread). The pH of the bread is a decisive factor while increased concentrations of vitamin B₁, protein and salt favor aflatoxin formation.

Although \rightarrow patulin contamination has been reported in spontaneously molded bread this mycotoxin reacts with sulfhydryl-containing amino acids or proteins and is therefore not stable in this substrate.

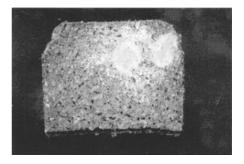
Japanese studies proved a 50% reduction in \rightarrow deoxynivalenol levels compared to the original concentration by baking. However, in Canadian experiments only a 20% reduction of this mycotoxin was observed and almost none during Egyptian bread making. In Japanese bread making losses of various trichothecenes (deoxynivalenol, \rightarrow nivalenol, \rightarrow diacetoxyscirpenol, \rightarrow neosolaniol, \rightarrow T-2 toxin

and \rightarrow fusarenon X) amounted to $\approx 50\%$ (artifical contamination). Bread may contain the following mycotoxins: → acetyldeoxynivalenol incidence: 4/24*, conc. range: 600-2400 μg/kg, country: India, *wheat \rightarrow aflatoxin B₁ incidence: 4*/18**, conc. range: 5-60 μg/kg, country: Germany, *moldy, **whole meal wheat incidence: 1*/14**, conc.: 10 µg/kg, country: Germany, *moldy, **German "Landbrot" (80% wheat and 20% rye flour) incidence: 2*/18**, conc. range: 20-25 μg/kg, country: Germany, *moldy, **white \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 1/4*, conc.: 3.3 µg/kg, country: UK \rightarrow citrinin incidence: 11*/110, conc. range: ≤ 5 μg/kg, country: Germany, *sliced packed bread, visible moldy deoxynivalenol incidence: 1/1*, conc.: 378 µg /kg, country: Argentina, *bran incidence: 4/4*, conc. range: 269-384 μ g/kg, Ø conc. 327 μ g/kg, country: Argentina, *homemade incidence: 10/12*, conc. range: 198-436 μ g/kg, Ø conc.: 263 μ g/kg, country: Argentina, *french incidence: 4 products analysed*, Ø conc.: 58 µg/kg, country: Canada, *rye bread incidence: 11/24*, conc. range: 340-8400 μg/kg, country: India, *wheat incidence: nc/4*, conc. range: 8-28 μg/kg, country: UK, *pitta \rightarrow fumonisin B₁ incidence: 1/2*, conc.: 80 µg/kg, country: The Netherlands \rightarrow fumonisins incidence: 4/4*, conc. range: 400-3450 μ g / kg, Ø conc.: 1285 μ g / kg, country: USA

incidence: 1/1*, conc.: 600 µg/kg, country: USA, *maize nivalenol incidence: 2/24*, conc. range: 30-100 μ g/kg, Ø conc.: 65 μ g/kg, country: India, *wheat incidence: $1/4^*$, conc.: 21 µg/kg, country: UK, *pitta incidence: 20/25, conc. range: ND-240 μg/kg, country: USA ochratoxin A incidence: $4^{*}/110$, conc. range: ≤ 80 μg/kg, country: Germany, *sliced packed bread, visibly moldy incidence: 4/57*, conc. range: 0.1-1 µg/kg, \emptyset conc.: 0.07 µg/kg, country: Germany, *wheat and rye bread (German "Mischbrot") incidence: 26/51, conc. range: \leq 1.49 µg / kg, \emptyset conc.: 0.17 µg / kg, country: Germany incidence: 4/36*, conc. range: 0.2-0.3 μg/kg, country: Germany, *whole meal bread incidence: 6/46*, conc. range: 0.2-0.9 µg/kg, country: Germany, *crisp incidence: 33/47*, conc range: 0.1-0.49 μg/kg (16 samples), 0.5-9.99 μg/kg (17 sa), country: Germany, *crisp incidence: 8/9*, conc. range: 0.05-0.49 μ g/kg (7 samples), 0.5-1.49 μ g/kg (1 sa), country: Germany, *Pumpernickel incidence: 6/8*, conc. range: 0.1-0.49 μ g / kg (3 samples), 0.5-1.49 μ g / kg (3 sa), country: Germany, *toast incidence: 1/2*, conc.: 80,000 µg/kg, country: Italy, *moldy, intended for animal feed incidence: 63/386, Ø conc.: 1360 µg/kg, country: Poland incidence: 11/26*, conc.: \leq 0.6 µg/kg, Ø conc.: 0.2 μ g / kg, country: Sweden, *crisp incidence: 1/2, conc.: 0.2 µg/kg, country: Switzerland incidence: 1/50*, conc.: 210 µg/kg, country: UK, *moldy incidence: 3/4*, conc.: 0.2-0.8 µg/kg, country: UK, *pitta

incidence: 6/32*, conc. range: nc, country: Yugoslavia, *wheat incidence: 1*/50**, conc.: 210 µg/kg, country: Yugoslavia, *moldy, **wheat \rightarrow ochratoxin B incidence: 1/2*, conc.: 9600 µg/kg, country: Italy, *moldy, intended for animal feed incidence: 6/32*, conc. range: nc, country: Yugoslavia, *wheat T-2 toxin incidence: 5/24*, conc. range: 550-4000 µg/kg, country: India, *wheat \rightarrow zearalenone incidence: $6^{*}/110$, conc. range: $\leq 5 \,\mu g / kg$, country: Germany, *sliced packed bread, visibly moldy incidence: 2/2*, conc. range: 250-750 μ g / kg, Ø conc.: 500 μ g / kg, country: Papua, New Guinea, *imported, wheat bread crumbs \rightarrow cereals

Breakfast cereals may be contaminated by various \rightarrow mycotoxins. This results from the fact that this kind of foodstuff is made from different kinds of \rightarrow cereals and \rightarrow cereal products which are often contaminated by \rightarrow Fusarium, \rightarrow Aspergillus and \rightarrow Penicillium mycotoxins. The detection of \rightarrow deoxynivalenol in breakfast cereals proves DON contamination of the grains and its survival through processing \rightarrow bread.



Bread. Aspergillus flavus Link on Pumpernickel

Breakfast cereals

Breakfast cereals may contain the following mycotoxins: \rightarrow aflatoxins (no specification) incidence: 2/6*, conc. range: 1-5 µg/kg, country: UK, $* \rightarrow$ bran-based incidence: 1/6*, conc. range: 1-5 µg/kg, country: UK, * \rightarrow maize-based incidence: 1/6*, conc. range: 1-5 µg/kg, country: UK, * \rightarrow oat-based incidence: 3/5*, conc. range: 1-5 µg/kg, country: UK, $* \rightarrow$ rice-based incidence: 1/14*, conc. range: 1-5 µg/kg, country: UK, $* \rightarrow$ wheat-based deoxynivalenol incidence: 36 products analysed, Ø conc.: 86 μ g/kg, country: Canada incidence: 7/7*, conc. range: 30-100 µg/ kg, country: UK, *bran-based incidence: 2/3*, conc. range: traces, country: UK, *maize-based incidence: 35/60, conc. range: ND-530 μg/kg, country: USA incidence: 36/60, \emptyset conc.: 100 µg/kg, country: USA \rightarrow fumonisin B₁ incidence: 11/52*, conc. range: < 100-320 µg/kg, country: Canada, *maize-based incidence: 9/17, conc. range: < 10-330 μ g/kg, Ø conc.: 130 μ g/kg, country: USA incidence: nc/3**, conc. range: 1060-3630 μg/kg, country: Zimbabwe \rightarrow fumonisin B₂ incidence: nc/17, conc. range: < 10-70 μ g / kg, Ø conc.: 30 μ g / kg, country: USA incidence: nc/3**, conc. range: 240-910 µg/kg, country: Zimbabwe \rightarrow fumonisin B₃ incidence: nc/3**, conc. range: 130-230 μg/kg, country: Zimbabwe **health breakfast cereal \rightarrow fumonisins (FB₁, FB₂, FB₃) incidence: 12/50, conc. range: 11-194 μ g / kg, Ø con.: 29 μ g / kg, country: UK → ochratoxin A incidence: 13/54, conc. range: \leq 4.9-9.8 μ g / kg, Ø conc.: 0.51 μ g / kg, country: Germany

incidence: 2/26, conc. range: $\leq 0.5~\mu\text{g}$ / kg, country: Germany incidence: $3/6^*$, conc. range: < 10 µg/kg, country: UK, *bran-based incidence: 3/6*, conc. range: < 10-20 μg/kg, country: UK, *maize-based incidence: $2/6^*$, conc. range: < 10 µg/kg, country: UK, *oat-based incidence: $1/5^*$, conc. range: < 10 µg/kg, country: UK, *rice-based incidence: 7/14*, conc. range: < 10-50 μg/kg, country: UK, *wheat-based incidence: 12/243, conc. range: 5-108 μg/kg, country: UK → sterigmatocystin incidence: 1/14 (wheat-based), conc.: ≤ 7 μg/kg, country: UK \rightarrow trichothecenes* (no specification) incidence: 5/6*, conc. range: nc, country: UK, *maize-based incidence: 4/6*, conc. range: nc, country: UK, *oat-based incidence: 5/13*, conc. range: nc, country: UK, *wheat-based * max. level: $\leq 5 \,\mu g / kg$ \rightarrow zearalenone incidence: $2/6^*$, conc. range: < 50 µg/kg, country: UK, *maize-based incidence: $3/14^*$, conc. range: $< 50 \ \mu g/$ kg, country: UK, *wheat-based incidence: 4/39, conc. range: 2.6-8.6 μ g / kg, Ø conc.: 4.6 μ g / kg, country: USA

Breakfast drinks may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A incidence: 2/2, conc. range: 0.1-0.3 μ g/kg, Ø conc.: 0.2 μ g/kg, country: Switzerland \rightarrow apple juice, \rightarrow fruit juice, \rightarrow grape juice, \rightarrow soft drinks

Brick cheese \rightarrow cheese, brick

Brie cheese \rightarrow cheese, Brie

Buckwheat may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 23/123, conc. range: 0.1-4.2 μg/kg, country: Japan \rightarrow aflatoxin B₂ incidence: 23/123, conc. range: 0.1-0.9 μg/kg, country: Japan \rightarrow aflatoxin G₁ incidence: 23/123, conc. range: 0.2-0.8 μg/kg, country: Japan \rightarrow aflatoxin G₂ incidence: 23/123, conc. range: tr-0.1 μg/kg, country: Japan \rightarrow ochratoxin A incidence: 1/34, conc.: 5 µg/kg, country: Germany \rightarrow cereals

Buckwheat flour may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ & \rightarrow aflatoxin B₂ incidence: 1*/37, conc.: \approx 10 µg/kg, country: Japan, *moldy \rightarrow flour

Bulla A large blister or skin vesicle filled with fluid.

Buns may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 21 products analysed, \emptyset conc.: 80 µg/kg, country: Canada \rightarrow cereals

Burukutu \rightarrow beer, burukutu

Butenolide is a 4-acetamido-4-hydroxy-2-butenoicacid χ -lactone (\rightarrow mycotoxins) derived from glutamic acid and associated with outbreaks of "fescue foot" of cattle in the US, Australia, and New Zealand (see Figure Butenolide).

CHEMICAL DATA Empirical formula: $C_6H_7NO_3$, molecular weight: 141 FUNGAL SOURCES

→ Aspergillus terreus Thom, *Fusarium* acuminatum Ellis & Everh. sensu Gordon?, → Fusarium avenaceum (Fr.) Sacc.?, → Fusarium graminearum Schwabe, → Fusarium poae (Peck) Wollenw.?, *F.* semitectum ?, → Fusarium sporotrichioides Sherb., → Fusarium equiseti (Corda) Sacc. sensu Gordon NATURAL OCCURRENCE

 \rightarrow wheat, \rightarrow barley (11 of 34 samples were contaminated between 10-430 μ g/kg)

TOXICITY

 LD_{50} : 43.6±1.24 mg/kg bw mice

FURTHER COMMENTS

Butenolide was occasionally detected in association with \rightarrow neosolaniol, \rightarrow T-2 toxin and \rightarrow diacetoxyscirpenol. Besides \rightarrow zearalenone and 12,13-epoxythrichothecene (\rightarrow trichothecenes) derivatives it belongs to the major toxic metabolites (mycotoxins) of \rightarrow Fusarium spp.

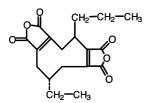
Butter Manufacturing butter from naturally contaminated cream (\rightarrow cream) 18-28% of the \rightarrow aflatoxin M₁ was found in the finished butter. However, the major portion occurred in the buttermilk (\rightarrow milk, butter-).

Buttermilk → milk, butter-

Byssochlamic acid belongs to the group of nonadrides characterized by the presence of anhydride groups attached to a nine membered carbocyclic ring (10ethyl-5,9,10,11-tetrahydro-4-propyl-1Hcyclonona[1,2-c:5,6-c']difuran-1,3,6,8(4H)-tetrone). Further members of this group are the glaucanic and glauconic acids (\rightarrow Penicillium *purpurogenum*)

NHCOCH

Butenolide



Byssochlamic acid

and the rubratoxins (*P. rubrum*) with a complex formulae (see Figure Byssochlamic acid).

CHEMICAL DATA

Empirical formula: $C_{18}H_{24}O_8$, molecular weight: 368

FUNGAL SOURCES

 \rightarrow Byssochlamys spp. (B. fulva, B. nivea),

 \rightarrow Paecilomyces variotii Bain

NATURAL OCCURRENCE Fruit juices may be contaminated.

Тохісіту

cytotoxic, hemorrhagic (\rightarrow hemorrhage) LD₅₀ > 2.5 g/kg bw but < 4.9 g/kg bw mice.

Byssochlamic acid is not as toxic as \rightarrow patulin.

DETECTION TLC

ILC

Further Comments

Up to now, no mycotoxicoses due to the consumption of contaminated \rightarrow foods have been reported. From the chemical structure it was concluded that only foods that contain fatty acids, with free glycerol present, are suitable for the production of byssochlamic acid. Therefore, byssochlamic acid is not a contaminant of margarine, \rightarrow olive oil or \rightarrow ham, whereas a metabolite very similar to

byssochlamic acid may be formed in \rightarrow butter. However, there are only few reports concerning the spoilage and contamination of foods with heat-resistant fungi like \rightarrow Byssochlamys spp., \rightarrow Paecilomyces variotii Bain, as well as byssochlamic acid.

Byssochlamys \rightarrow Trichocomaceae, anamorph \rightarrow Paecilomyces The only two food-relevant species: Byssochlamys fulva Olliver & Smith* and B. nivea Westl.** possess very heat-resistant ascospores and may cause degradation (pectinase activity) and spoilage (mycotoxin contamination) of processed (heated) fruit products as well as canned and bottled \rightarrow fruits. Reduced oxygen tensions (0.27% O₂) present in such commodities are tolerated. Adequate heat treatment for complete destruction of all inherent ascospores could impair the organoleptic quality of the product. Compared to B. fulva, strains of B. nivea possess a higher potential for \rightarrow patulin production. Although B. nivea forms patulin in \rightarrow apple juice under low O₂ levels (0.5-2%), the production of significant levels of patulin under commercial conditions is not anticipated. The minimum $\rightarrow a_w$ for patulin formation was a_w 0.950 at 37 °C after 10 days of incubation.

Byssochlamys spp. may produce \rightarrow mycotoxins such as \rightarrow byssochlamic acid* **, byssotoxin A*, malformins**, \rightarrow patulin* **, and variotin. Up to now, no mycotoxicosis due to the consumption of foods contaminated with byssochlamic acid has been reported.

C

Cabbage (fried with pork and garlic) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/1, conc.: 748 µg/kg, country: Thailand, *total: 1299 µg AFB₁, AFB₂, AFG₁, AFG₂/kg food \rightarrow beans, \rightarrow cowpeas, \rightarrow lentils, \rightarrow peas, \rightarrow pigeon peas, \rightarrow soybeans, \rightarrow vegetables

Caesalpinea digyna (medicinal seeds) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: nc/nc, conc. range: 80-1180 µg/kg, country: India

Carbon dioxide → atmosphere

Carcinoma A new growth or malignant tumor enclosing epithelial cells in connective tissue and tending to infiltrate and give rise to metastases. \rightarrow Aflatoxin B₁, \rightarrow aflatoxin G₁, \rightarrow aflatoxin M_1 , \rightarrow sterigmatocystin, versicolorin A, \rightarrow luteoskyrin and \rightarrow rugulosin are \rightarrow mycotoxins with a well-known carcinogenic potential. These toxic fungal metabolites are genotoxic and produce positive results in short-term genotoxicity assays (Ames mutagenicity test). Similar genotoxic porperties have been reported for \rightarrow fusarin C and emodin which are also likely to be carcinogenic. The genotoxicity of the \rightarrow trichothecenes, \rightarrow ochratoxin A and \rightarrow zearalenone is questionable or non-existent, but they definitely promote cancer like the \rightarrow fumonisins.

Cardamom (*Elettaria cardamomum* Linn.)

may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁

incidence: 1/6, conc.: 20 $\mu g\,/\,kg,$ country: India

→ aflatoxin B₂ incidence: 1/6, conc.: 15 µg/kg, country: India → aflatoxin G₁ incidence: 1/6, conc.: 12 µg/kg, country: India → citrinin incidence: 1/6, conc.: 25 µg/kg, country: India → spices

Cardamom, greater (Amomum subula-

tum Roxb.) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 3/6, conc. range: 18-129 μg/kg, country: India aflatoxin B₂ incidence: 3/6, conc. range: 14-108 μ g / kg, country: India \rightarrow aflatoxin G₁ incidence: 3/6, conc. range: 17-78 µg/kg, country: India \rightarrow aflatoxin G₂ incidence: 3/6, conc. range: 5-76 µg/kg, country: India \rightarrow ochratoxin A incidence: 1/6, conc.: nc, country: India \rightarrow spices

Cardiac beriberi → Acute cardiac beriberi

Carry over Edible tissues, \rightarrow milk, and eggs of food-producing animals in general are contaminated with only low levels of \rightarrow mycotoxins. This is due to the fact that only minor amounts of mycotoxins and/or their metabolites are transmitted from the feeds to this kind of foodstuff. For the extent of mycotoxin carry over and contamination the total dose level, not the duration of dose is mainly responsible. The main important "carry over" myco-

The main important "carry over" mycotoxins are \rightarrow aflatoxin B₁ / \rightarrow aflatoxin M₁ in \rightarrow milk and \rightarrow milk products and \rightarrow ochratoxin A in \rightarrow meat and meat products (e.g. \rightarrow sausages). To avoid aflatoxin contamination, many countries have a strict regulation for these mycotoxins at the feed level. OTA residues in meat are monitored by regulatory authorities especially in European countries. There are also carry over studies for \rightarrow trichothecenes, \rightarrow zearalenone, and \rightarrow fumonisins. However, it seems that these mycotoxins are only of minor importance concerning a carry over. In addition, analytical detection may be difficult especially because the identity and hazard of the metabolites are unknown.

Cashew nuts may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1*/6, conc.: 830 µg/kg, country: Germany, *moldy \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 23/120, conc. range: 5-24.9 µg/kg (19 samples), > 25 µg/kg (4 sa), country: Canada incidence: 1/3*, conc.: traces, country: Norway, *imported \rightarrow nuts

Cassava (raw)

may contain the following \rightarrow mycotoxins: aflatoxin (no specification) incidence: 1/1, conc.: 1700 µg/kg, country: Uganda incidence: 23/23, conc. range: > 20 µg/kg (13 samples), country: Philippines (very high concentrations have been detected but no data were presented) \rightarrow aflatoxins (no specification) incidence: 4*/34, conc. range: 100-1000 µg/kg (2 samples), > 1000 µg/kg (2 sa), country: Uganda, * 2 samples contained AFB₁, 4 sa AFB₂, 2 sa AFG₁

Cassava flour may contain the following

 \rightarrow mycotoxins:

 \rightarrow ochratoxin A

incidence: 2/2, conc. range: 32-65 μ g/kg, Ø conc.: 48.5 μ g/kg, country: Brazil

Cassava starch → Sago

Cassia fistula (medicinal seeds) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: nc/nc, conc. range: 80-1110 µg/kg, country: India \rightarrow citrinin incidence: nc/nc, conc. range: 10-690 µg/kg, country: India

Cattle are relatively resistant against \rightarrow Fusarium mycotoxins in their diet. However, elevated concentrations caused different symptons like feed refusal, gain losses, impaired \rightarrow milk production, diarrhea, decreased immune response, and abnormal estrous cycles. Feeds contaminated with 200 µg or 800 µg \rightarrow deoxynivalenol/kg dry matter lowered milk production with 0.45 kg and 2 kg/day, respectively. Furthermore, the higher concentration caused a delay in breeding (8 days).

 \rightarrow cattle liver, \rightarrow meat

Cattle liver may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 6/19, conc. range: 0.02-0.08 μ g/kg (5 samples), 6.6 μ g/kg, (1 sa) country: Germany \rightarrow aflatoxin B₂ incidence: 4/19, conc. range: 0.01-0.03 μ g/kg, Ø conc.: 0.02 μ g/kg, country: Germany \rightarrow aflatoxins (no specification) incidence: 1/19, conc.: 10.3 μ g/kg, country: Germany \rightarrow meat

55

incidence: 16/56, conc. range: < 2.5-8µg/kg, country: Canada incidence: 10/33, conc. range: tr-8 µg/kg, country: Canada incidence: 3/36, conc. range: 5.0-9.8 µg/kg, Ø conc.: 7.16 µg/kg, country: Germany incidence: 11/22, conc. range: tr-24 µg/kg, country: Germany \rightarrow aflatoxin B₂ incidence: 5/33, conc. range: traces, country: Canada \rightarrow spices

Celery seeds may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin G₁ incidence: 1/9, conc.: 3.7 µg/kg, country: unknown

Cereal flakes may contain the following \rightarrow mycotoxins:

 \rightarrow ochratoxin A

incidence: 13/51, conc. range: 0.1-0.49 $\mu g / kg$ (6 samples), 0.5-1.49 $\mu g / kg$ (4 sa), 1.5-9.99 $\mu g / kg$ (3 sa), country: Germany incidence: 1/5, conc.: 1 $\mu g / kg$, country: Germany \rightarrow corn flakes, \rightarrow maize flakes, \rightarrow oat flakes

Cereal food (mixed) Bsissa is a Tunesian-type of food composed of ground \rightarrow barley, chick pea, and \rightarrow wheat. Bsissa may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A incidence: 11/11, conc. range: 0.4-12,770 µg/kg, country: Tunesia

Cereal products may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A incidence: 63/158, Ø conc.: 0.9 µg/kg, country: Germany incidence: 5/25, conc. range: 0.1-0.49 μ g/kg (2 samples), 1.5-9.99 μ g/kg (1 sa), country: Germany incidence: 10/32, conc range: 0.1-0.49 μ g/kg (6 samples), 0.5-1.49 μ g/kg (4 sa), country: Germany incidence: 2/7, conc. range: 0.1-0.49 μ g/kg, country: Germany incidence: 8/54, conc. range: 0.3-5.3 μ g/kg, country: Germany incidence: 1/40*, conc.: 2 μ g/kg, country: Germany, *whole meal incidence: 1/30, conc.: 6.2 μ g/kg, country: Japan

Cereal products (whole meal) Contamination of cereal products with \rightarrow mycotoxins mainly result from infection of the \rightarrow grains in the field with mycotoxin producing fungi, especially \rightarrow Fusarium spp. \rightarrow Maize may be contaminated with aflatoxin producers (\rightarrow aflatoxins). Under moderate conditions the \rightarrow trichothecenes, mainly found in cereal grains, are relatively stable and very hard to remove from contaminated \rightarrow cereals. During \rightarrow milling processes they are distributed in food and feed. Food processing such as \rightarrow baking and boiling in water and \rightarrow oil does not cause their complete destruction. It is estimated that ca. 50% of trichothecenes remained in the final food products (e.g. \rightarrow bread, \rightarrow noodles).

Cereal grains may contain the following \rightarrow mycotoxins:

→ deoxynivalenol

incidence: 4/4, conc. range: 255-490 μ g/kg, Ø conc.: 386 μ g/kg, country: Austria

Cereals (no specification) Cereals and \rightarrow oil seeds belong to the most suitable substrates for \rightarrow mycotoxins. Although mycotoxin contamination has been reported in \rightarrow grains like \rightarrow oats, \rightarrow rice, \rightarrow rye, \rightarrow sorghum cer-

Cereals

eals such as \rightarrow barley, \rightarrow maize, and \rightarrow wheat seem to be more susceptible to mycotoxin formation. In general, lowgrade cereals show a higher degree of mycotoxin contamination. Such grains in the developed countries normally do not enter the human food chain. However, in many developing countries the high quality cereals are often shipped abroad whereas the low-quality grains serve for human consumption.

Mycotoxin contamination (\rightarrow Fusarium mycotoxins, \rightarrow trichothecenes, \rightarrow zearalenone) of cereal grains usually occurs during growth and maturing of the plants while it is less common after harvest and during storage. Here, an a_w of > 0,83 (\approx > 17% moisture content) is necessary. Even aflatoxin (\rightarrow aflatoxins) contamination of cottonseed, maize, and \rightarrow peanuts may occur before and during harvest although \rightarrow Aspergillus flavus Link belongs to the group of \rightarrow storage fungi. Wheat, barley, and maize constitute twothirds of the world production of cereals but similarly appear to be most affected by Fusarium mycotoxins. In detail, the relative vulnerability to mycotoxin formation is high for amber durum wheat, moderate for maize and six-row barley, but low for two-row barley and hard red spring wheat. Although contamination of oats, rye, and \rightarrow triticale with trichothecenes and zearalenone is possible, these crops, except for some triticale varieties, seem to be more resistant or undergo no significant mycotoxin contamination.

Four trichothecenes, viz. \rightarrow deoxynivalenol, \rightarrow diacetoxyscirpenol, \rightarrow nivalenol, and \rightarrow T-2 toxin in general predominate in cereals grown in wet temperate regions like Northern Europe, parts of Northern America (e.g. Canada) and Japan. If cereals show a \rightarrow zearalenone contamination there is a high probability that other *Fusarium* mycotoxins are also present. The application of fungicides may cause the accumulation of more mycotoxin in the grains by affecting the fungal metabolism.

Formation of Aspergillus and Penicillium mycotoxins is common in stored, inadequately dried agricultural products and / or rewetting of dried products, mainly by condensation, but also by flooding or when water leaks into storage bins. In cereals \rightarrow aflatoxin B₁ and \rightarrow aflatoxin B₂ are more often found than AFB₁, AFB₂, AFG₁, and AFG₂. However, aflatoxin contamination is a primarily a problem in maize. Rice is affected only in circumstances of poor storage in tropical and subtropical countries.

Stored ground (feed) seeds, lacking the outer protective testa, especially promote fungal growth since the rich nutrients inside are easily colonized by these storage fungi.

Cereals and \rightarrow cereal products are mainly responsible for the \rightarrow ochratoxin A intake at least in Europe since there is always the chance of contamination and the consumption of cereals is generally not low. In general, wheat and maize show a lower OTA contamination than rye. Mean levels of 0-2 µg OTA / kg on the EU market seem to be realistic. Temperate climatic conditions and drying with forced ambient air especially in Scandinavia favor OTA production in cereals. Other, also important, factors are mechanical injury and fungal infection, drying practice (e.g. promptness and rapidity of drying, rewetting) as well as improper storage techniques ([†] moisture contents, [†] temperatures, \uparrow oxygen, \uparrow time). It is suggested that OTA contamination mainly occurs during the first period just after harvest before the aw has decreased to a level which slows down or inhibits OTA formation. During longer periods of storage OTA production may occur if the storage conditions are unfavorable. In cereal fractions of wheat and barley (>2,5 mm), OTA concentrations reach

80-100% of the initial concentration in the corresponding grains. This mycotoxin is mainly present in the inner, deeper parts of the kernels but not on the surface of the grains. Chloroform extraction only removed 10-50% of the toxin from this part of the grain. \rightarrow Milling results of these \rightarrow grains show that the level of OTA in \rightarrow flour is similar to that in \rightarrow bran.

Chaff and kernels of small grain cereals (e.g. rye, wheat) may contain \rightarrow Alternaria mycotoxins. The amount of such mycotoxins depends on the percentage of "black heads" due to \rightarrow Alternaria alternata (Fr.) Keissler - not A. infectoria which is similar to A. alternata but a weak mycotoxin producer - at harvest time. However, cereals, e.g. wheat, without black heads or weather damage may also contain low levels of \rightarrow tenuazonic acid. Alternaria mycotoxin formation is favored by high humidity and rainy weather before harvest. The production of Alternaria mycotoxins during storage is unlikely due to the low $\rightarrow a_w$ of the stored grains.

According to Frisvad (1988) the following mycotoxins may be found in cereals, maize, \rightarrow peas and \rightarrow beans under field conditions: \rightarrow aflatoxins, \rightarrow alternariol, \rightarrow alternariol methyl ether, \rightarrow altertoxins I-III, \rightarrow butenolide, \rightarrow cyclopiazonic acid, \rightarrow fusarin C, \rightarrow moniliformin, \rightarrow tenuazonic acid, \rightarrow trichothecenes, and \rightarrow zearalenone. Stored cereals may be contaminated with aflatoxins, \rightarrow citrinin, cyclopiazonic acid, ochratoxin A, \rightarrow penicillic acid, \rightarrow sterigmatocystin, \rightarrow viomellein, and \rightarrow xanthomegnin. The most probable mycotoxin in airtight stored cereals is \rightarrow patulin.

Cereals may contain the following \rightarrow mycotoxins:

 \rightarrow aflatoxin B₁

incidence: 10/71*, conc. range: $< 5-300 \mu g / kg$, country: South Africa, *includes oats, wheat, barley

 \rightarrow aflatoxins (no specification) incidence: 1/52, conc.: 20 µg/kg, country: Japan \rightarrow citrinin incidence: 1*/52, conc.: 27 µg/kg, country: Japan, *maize flour incidence: 4/735, conc. range: tr-6000 μ g / kg, country: Poland ergometrine (\rightarrow ergot alkaloids) incidence: 2/2, conc. range: 0.8-6.4 μ g/kg, Ø conc.: 3.6 μ g/kg, country: Canada, *wheat, rye, flax (mixture) ergosine incidence: 2/2, conc. range: 12-14 µg/kg, \emptyset conc.. 13 µg/kg, country: Canada, *wheat, rye, flax (mixture) ergotamine incidence: 2/2, conc. range: 14-20 µg/kg, Ø conc.: 17 μ g / kg, country: Canada, *wheat, rye, flax (mixture) ergocornine incidence: 2/2, conc. range: 1.7-6.7 μ g/kg, Ø conc.: 4.2 μ g/kg, country: Canada, *wheat, rye, flax (mixture) α-ergokryptine incidence: 2/2, conc. range: 1.1-6.6 μ g/kg, Ø conc.: 3.85 μ g/kg, country: Canada, *wheat, rye, flax (mixture) ergocristine incidence: 2/2, conc. range: 36-40 µg/kg, \emptyset conc.: 38 µg/kg, country: Canada, *wheat, rye, flax (mixture) \rightarrow ochratoxin A incidence: 6/315, conc. range: 3-8 µg/kg, country: Canada incidence: 5/440, conc. range: 10-50 μ g / kg, country: Canada incidence: 19/33*, conc. range: 28-27,500 μg/kg, country: Denmark, *barley, oats incidence: 2/151*, conc. range: 15-50 μg/kg, country: Denmark, *rye, wheat incidence: 8/11*, conc. range: < 4.9-12.8 µg/kg, country: France, *maize, barley, oats incidence: $1/13^*$, conc.: 2 µg/kg, country: France, *wheat, barley

Cereals

incidence: 4/40, conc. range: < 4.9-22 μg/kg, country: France incidence: 24/765, Ø conc.: 11.8 μg/kg, country: Germany incidence: 2/49, conc. range: 18-22 μ g / kg, Ø conc.: 20 μ g / kg, country: Germany incidence: 12/39*, conc. range: 0.1-2.7 μg/kg, country: Germany, *partly imported from different countries incidence: 30/232*, conc. range: 0.1-206 μg/kg, country. Germany, *wheat, rye, oats incidence: 18/43, conc. range: 2-304 μg/kg, country: Germany incidence: 11/538, conc. range: 2-180 μg/kg, country: Norway incidence: 63/784, conc. range: tr-1100 μg/kg, country: Poland incidence: 6/100, conc. range: tr-1200 μg/kg, country: Poland incidence: 20/296*, conc. range: 20-470 μg/kg, country: Poland, *barley, rye, wheat incidence: 8/150, conc. range: 50-200 μ g / kg, country: Poland incidence: 158/1.353, conc. range: 5-2400 μg/kg, country: Poland incidence: 7/84, conc. range: 16-410 µg/kg, country: Sweden incidence: 6/47*, conc. range: 5-90 µg/kg, country: Yugoslavia, *barley, maize, wheat \rightarrow patulin incidence: 8/71*, conc.: nc, country: South Africa, *includes oats, wheat, barley \rightarrow penicillic acid incidence: 4/736, conc. range: tr-1300 μ g / kg, country: Poland \rightarrow zearalenone incidence: 2/377, conc. range: tr-700 μ g / kg, country: Poland \rightarrow barley, \rightarrow buckwheat, \rightarrow grains, \rightarrow maize, \rightarrow millet, \rightarrow oats, \rightarrow rice, \rightarrow rye, \rightarrow sorghum, \rightarrow triticale, \rightarrow wheat

Chapatti may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: nc/4, conc. range: 6-10 µg/kg, country: UK \rightarrow nivalenol incidence: 1/4, conc.: 16 µg/kg, country: UK \rightarrow ochratoxin A incidence: 2/4, conc.: 0.5-0.9 µg/kg, Ø conc.: 0.7 µg/kg, country: UK

Cheddar cheese → cheese, Cheddar

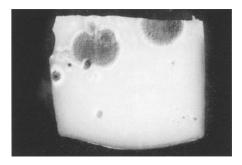
Cheese (no specification) Mycotoxin producers of the genus \rightarrow Penicillium, probably because of tolerance of low temperatures, are the most important contaminants on cheese during ripening and storage at low temperatures (ca. 80% of the total isolates) followed by \rightarrow Aspergillus spp. (ca. 10%) and molds of other genera (ca. 10%). A hazard to human health is not necessarily if cheese exhibits some moldy spots. However, during prolonged storage periods mold growth and subsequent mycotoxin production becomes more probable. Moldspoiled cheeses should therefore be excluded from human consumption. To inhibit mold growth and subsequent mycotoxin contamination, the relative humidity in the curing room must be precisely and regularly checked. Some shrinkage of the cheeses is better than the development of mycotoxin producing molds. Plastic emulsions are treated with i.e. sorbate or pimaricin, which coat the cheeses so as to give them further protection against fungal infection. Hygienic measure, like cleaning and disinfecting of shelves in the curing rooms, also contribute to optimal cheese manufacture. Although Penicillium spp. are well known mycotoxin producers, the most frequent \rightarrow mycotoxins in cheese are the \rightarrow aflatoxins, especially \rightarrow aflatoxin M₁. During

production processes no destruction of AFM₁ has been reported. There are three possible contamination routes: (i) \rightarrow Carry over of \rightarrow aflatoxin B₁ from cow feed into the raw \rightarrow milk leads to aflatoxin M₁ accumulation. (ii) Although cheese might be a less favorable substrate for mycotoxin production, it may still support surface growth of molds and subsequent mycotoxin formation (e.g. \rightarrow sterigmatocystin); molding might occur during ripening in warehouses and after cutting and slicing during storage in shops or at home. (iii) Contamination of \rightarrow milk powder used to enrich the milk used to make cheese.

Cheese processing may result in the accumulation of aflatoxin M_1 in the curd. The affinity of AFM₁ for casein due to (possible) hydrophobic interactions with hydrophobic areas of the milk protein may be the reason. However, AFM₁ contamination of the whey in the range of 50-100% has also been reported. Decisive factors for the pattern of distribution are type and degree of milk contamination as well as milk quality, cheese processing, "contamination" of whey with AFM₁ contaminated curd, extraction technique, methodology, and expression of the results. There is an almost homogenous distribution in the concentration of AFM₁ concentration from the rind to the center (related to dry weight). The stability of AFM₁ during ripening and storage was shown in different kinds of cheese, e.g. Camembert, Cheddar, Parmesan, Swiss.

Since the mid-1980s, AFM_1 contamination in cheeses has declined, most probably as a result of strict regulations regarding aflatoxin levels in feeds in different countries.

Production of AFB_1 and AFG_1 apparently does not occur in Romadur or Camembert cheeses. However, such aflatoxins have been detected in Tilsit and Brick



Cheese: Penicillium sp. on Edam cheese

Cheese and in Emmentaler after they had been inoculated with aflatoxigenic molds. In addition to the aflatoxins, cheeses may be contaminated with further mycotoxins such as ochratoxin A and citrinin (see Figure Cheese). They may migrate into the cheese to a depth of at least 20 mm. In the case of \rightarrow citrinin, diffusion lead to higher concentrations inside the cheese compared to its crust. Sterigmatocystin is stable in hard cheese for more than 3 months.

Cheese may contain the following \rightarrow mycotoxins: aflatoxin B₁ incidence: 6/26, conc. range: 5-15 µg/kg, country: India incidence: 1/248, conc.: 1 μ g/kg, country: Tunesia aflatoxin B_1 and/or \rightarrow aflatoxin G_1 incidence: 79/133, conc. range: 10-50 μg/kg, country: Egypt incidence: 16/222, conc. range: ≤ 10 μg/kg, country: Germany aflatoxin M₁ incidence: 1/1*, conc.: 0.1 µg/kg, country: Canada incidence: 60/60*, conc range: \leq 0.26-0.89 µg/kg, country: Canada, *imported incidence: 6/29, conc. range: 0.005-0.066 μg/kg, country: Czechoslovakia incidence: 19/19*, conc. range: 0.1-0.4 μ g/kg, Ø conc.: 0.18 μ g/kg, country: Denmark

Cheese

incidence: 11/45, conc. range: 0.1-0.4 μg/kg, country: Denmark incidence: 16/82, conc. range: < 0.005- $> 0.25 \ \mu g / kg$, country: France incidence: 102/343, conc. range: ≤ 5.2 μ g/l, country: France incidence: 9/14*, conc. range: 0.1-0.3 μ g / kg, Ø conc.: 0.17 μ g / kg, country: France incidence: 9/34, conc. range: < 0.005- $> 0.25 \ \mu g / kg$, country: Germany incidence: 2/5* conc. range: 0.15 µg/kg, \emptyset conc.: 0.15 µg/kg, country: Germany incidence: 5/22, conc. range: 0.1-0.4 μg/kg, country: Germany incidence: 19/51*, conc. range: 0.1-0.2 μ g / kg, Ø conc.: 0.13 μ g / kg, country: Ireland incidence: 4/50, conc. range: 0.05-0.1 μg/kg, country: Italy incidence: 130/416, conc. range: < 0.005- $> 0.4 \ \mu g / kg$, country: Italy incidence: 71/83, conc. range: < 0.005- $> 0.25 \ \mu g / kg$, country: Italy incidence: 5/6, conc. range: 0.05-0.4 μ g / kg, country: Italy incidence: 4/50, conc. range: 0.05-0.1 μ g / kg, country: Italy incidence: 2/4, conc. range: 0.02-0.04 μg/kg, country: Italy incidence: 7/10, conc. range: 0.2-1.14 µg/ kg, country: Italy incidence: 12/66, conc. range: 0.28-1.3 μg/kg, country: Italy incidence: 1/1*, conc.: 0.1 µg/kg, country: Italy incidence: 56/126, conc. range: 0.11-0.3 μg/kg, country: Japan incidence: 111/128, conc. range: 0.025-1.06 µg/kg, country: Japan incidence: 13/32, conc. range: 0.012-2.52 μg/kg, country: Japan incidence: 120/132, conc. range: 0.01-0.5 μg/kg, country: Japan incidence: 19/80*, conc. range: 0.1-1.2 µg/kg, country: Japan, *imported incidence: 23/43, conc. range: < 0.005- $> 0.25 \ \mu g / kg$, country: The Netherlands

incidence: 5/22*, conc. range: 0.15-0.5 µg/kg, country: The Netherlands incidence: 8/40, conc. range: 0.1-0.2 μ g / kg, country: The Netherlands incidence: $30/30^*$, conc. range: < 0.1µg/kg, country: New Zealand incidence: 2/248, conc. range: 6.2-10.6 µg/kg, country: Tunisia incidence: 86/143*, conc. range: < 0.10-0.50 µg/kg, country: UK, *imported incidence: 1/4, conc.: 0.2 µg/kg, country: UK incidence: 8/118*, conc. range: 0.1-1.0 μ g/g, country: USA, *imported \rightarrow aflatoxin M₄ incidence: 6/66, conc. range: 0.34-0.87 μg/kg, country: Italy \rightarrow aflatoxins incidence: 235/558, conc. range: < 0.25 μ g / kg (143 samples), > 25 μ g / kg (92 sa), country: Germany \rightarrow citrinin incidence: $17/44^*$, conc. range: < 50μg/kg, country: UK, *retail, domestic incidence: 3/nc, conc. range: nc, country: UK \rightarrow mycophenolic acid incidence: 38/100, conc. range: 20-15,000 μ g / kg, country: France $\rightarrow \beta$ -nitropropionic acid incidence: 5/18, conc. range: traces, country: USA \rightarrow ochratoxin A incidence: $18/44^*$, conc. range: ≤ 260 μg / kg, country: UK, *retail, domestic, wholesale \rightarrow dairy products, \rightarrow milk Cheese (hard) may contain the following → mycotoxins: \rightarrow aflatoxin M₁

incidence: 58/77, conc. range: 0.1-1.3

μg/kg, Ø conc.: 0.43 μg/kg, country: Germany

 \rightarrow mycophenolic acid

incidence: 4/48, conc. range: 10-1000 μg/kg*, country: France, *outer layer → patulin incidence: 1/48, conc.: 90 µg/kg*, country: France, *outer layer → penicillic acid incidence: 5/39, conc. range: ≤ 340 µg/kg*, country: France, *outer layer → sterigmatocystin incidence: 3/66, conc. range: 7.5-17.5 µg/kg, country: Czechoslovakia incidence: 9/39, conc. range: 5-600 µg/kg*, country: The Netherlands, *surface layer incidence: 3/48, conc. range: ≤ 330 µg/kg*, country: France, *outer layer

Cheese (processed) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ and/or \rightarrow aflatoxin G₁ incidence: 2/115, conc. range: nc, country: Germany \rightarrow aflatoxin M₁ incidence: 54/134, conc. range: 0.1-0.55 $\mu g/kg$, \emptyset conc.: 0.26 $\mu g/kg$, country: Germany incidence: 14/14, conc. range: 0.1-0.3 $\mu g/kg$, \emptyset conc.: 0.16 $\mu g/kg$, country: UK \rightarrow ochratoxin A incidence: 3/4, conc. range: 50-75 $\mu g/kg$, country: UK

Cheese (semi-hard) with a moldy crust may contain the following \rightarrow mycotoxins: \rightarrow mycophenolic acid incidence: 7/39, conc. range: 10-5000 µg/kg, country: France \rightarrow patulin incidence: 4/39, conc. range: 45-355 µg/kg, country: France \rightarrow penicillic acid incidence: 5/39, conc. range: \leq 710 µg/kg, country: France

Cheese (white, no further specification)

may contain the following \rightarrow mycotoxins: \rightarrow cyclopiazonic acid incidence: 2/6, conc. range: 250-370 μ g/kg, Ø conc.: 310 μ g/kg, country: France

Cheese, Bhutanese may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A incidence: 5/19, conc. range: 42-116 μ g/kg, country: India

Cheese, Bleu des Causses may contain the following \rightarrow mycotoxins: \rightarrow mycophenolic acid incidence: 3/6, conc. range: 10-1000 μ g/kg, country: France

Cheese, Blue may be contaminated by different metabolic products of \rightarrow Penicillium roquefortii Thom. \rightarrow PR toxin is the most acutely toxic but it is produced by only a limited number of industrial strains. Formation of PR toxin depends on specific cultural conditions (\downarrow pH, ↓ NaCl, presence of sucrose, sufficient oxygen) which significantly differ from industrial processing methods. These are quite the opposite in Blue Cheese ripening. In addition, because of reaction with neutral and basic amino acids PR toxin is not stable in Blue Cheese. Concentrations of the formed PR-imine, a probable degradation product of PR toxin, may be rather high (\leq 42,000 µg/kg). \rightarrow Roquefortine C as a frequent mycotoxin in Blue Cheese is concentrated in the moldy areas and often accompanied by roquefortine A, while roquefortine B $(\rightarrow$ roquefortine A & B) occurs to a minor degree. Blue cheese may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ and / or \rightarrow aflatoxin G₁ incidence: 2/62, conc. range: nc, country: Egypt

 \rightarrow aflatoxin M₁

Cheese, Blue

incidence: 5/5, conc. range: traces (4 samples), $< 0.1 \ \mu g / kg$ (1 sa), country: Germany \rightarrow mycophenolic acid incidence: 4/32, conc. range: 250-500 μg/kg, country: Germany incidence: 3/12, conc. range: $10- \le 1000$ μ g / kg, country: Germany (export to France) → penicillic acid incidence: 1/110, conc.: 820 µg/kg, country: France roquefortine A incidence: 1/1, conc.: 785 μ g/kg, country: Canada incidence: 7/7, conc. range: 135-4700 μ g / kg, Ø conc.: 1921 μ g / kg, country: Denmark incidence: 1/1, conc.: 1833 µg/kg, country: Finland incidence: 2/3, conc. range: 100-130 μ g / kg, Ø conc.: 115 μ g / kg, country: France incidence: 4/6, conc. range: tr-170 µg/kg, country: Germany incidence: 5/5, conc. range: 200-360 μg/kg, country: Japan incidence: 2/2, conc. range: tr(?)-80 μg/kg, country: UK roquefortine B incidence: 1/1, conc.: traces, country: Canada incidence: 4/7, conc. range: traces, country: Denmark incidence: 1/1, conc.: traces, country: Denmark \rightarrow roquefortine C incidence: 1/1, conc.: 1085 µg/kg, country: Canada incidence: 7/7, conc. range: 60-2300 μ g / kg, Ø conc.: 982 μ g / kg, country: Denmark incidence: 1/1, conc.: 66 µg, country: Finland incidence: 3/3, conc. range: 60-400 μ g/kg, Ø conc.: 230 μ g/kg, country: France

incidence: 4/6, conc. range: 370-6800 μ g/kg, Ø conc.: 2500 μ g/kg, country: Germany incidence: 3/3, conc. range: 490-1100 μ g/kg, Ø conc.: 737 μ g/kg, country: Switzerland incidence: 12/12, conc. range: 162-651 μ g/kg, Ø conc.: 424 μ g/kg, country: USA

Cheese, Blue Castello may contain the following \rightarrow mycotoxins: \rightarrow roquefortine C incidence: 1/1, conc.: 2290 µg/kg, country: France

Cheese, Blue Haverti may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 10/10, conc. range: 0.084-0.556 µg/kg, country: Denmark

Cheese, Bresse Bleu may contain the following \rightarrow mycotoxins: \rightarrow roquefortine C incidence: 1/1, conc.: 560 µg/kg, country: Denmark

Cheese, Brick \rightarrow Aspergillus flavus Link and \rightarrow Aspergillus parasiticus Speare produced \rightarrow aflatoxins on Brick Cheese at 23.9 °C and 12.8 °C, respectively.

Cheese, Brie may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 6/6, conc. range: 0.058-0.414 µg / kg, Ø conc.: 0.195 µg / kg, country: Denmark incidence: 6/14, conc. range: 0.055-0.714 µg / kg, country: France incidence: 2/2, conc. range: 0.024-0.029 µg / kg, Ø conc.: 0.0265 µg / kg, country: Germany

Cheese, butter may contain the following \rightarrow mycotoxins:

 \rightarrow aflatoxin M₁

incidence: 5/5, conc. range: $0.025-0.041 \mu g/kg$, Ø conc.: $0.037 \mu g/kg$, country: Germany incidence: 6/7, conc. range: traces (4 samples), < $0.1 \mu g/kg$ (2 sa), country:

Germany

Cheese, Camembert \rightarrow Cyclopiazonic acid represents an important mycotoxin in this kind of cheese. It occurs mainly in the crust rather than in the inner part. Not yet fully ripened cheeses stored in the cold do not contain more than 500 µg cyclopiazonic acid/kg (calculation on whole cheese). A significant increase up to 5000 µg cyclopiazonic acid/kg may result from temperatures during storage that are too high. Therefore, refrigerated storage and display, together with limited shelf life are recommended to prevent the accumulation of cyclopiazonic acid. However, the actual toxicological data, in combination with consumption habits, indicate that no risk to human health in reality exists.

Camembert may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 7/7, conc. range: 0.055-0.479 µg/kg, Ø conc.: 0.207 µg/kg, country: Denmark incidence: 18/25, conc. range: 0.013-0.565 µg/kg, country: France incidence: 1/100 conc.: traces, country: France incidence: 1/1, conc.: 0.018 µg/kg, country: Germany

incidence: 19/38, conc. range: traces (14 samples), < 0.1 μ g/kg (2 sa), > 0.1 μ g/

kg (3 sa), country: Germany

 \rightarrow cyclopiazonic acid

incidence: 11/20, conc. range: 0.05-0.1 μ g/kg (3 samples), 0.1-0.2 μ g/kg (5 sa), 0.4-1.5 μ g/kg (3 sa), country: France incidence: 1/3, conc.: 80 μ g/kg, country: Switzerland

Cheese, Camembert & Brie Camembert and Brie may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 33/65, conc. range: 0.1-0.73 $\mu g / kg$, \emptyset conc.: 0.31 $\mu g / kg$, country: Germany \rightarrow cyclopiazonic acid incidence: 11/11, conc. range: 60-290 $\mu g / kg$, country: Germany incidence: 11/20, conc. range: 50-1500

μg/kg, country: USA

Cheese, Cheddar \rightarrow Aspergillus flavus Link and \rightarrow Aspergillus parasiticus Speare produced substantial quantities of \rightarrow aflatoxins on Cheddar cheese at room temperature but no natural occurrence of these \rightarrow mycotoxins has been reported up to now.

Cheddar cheese may contain the following mycotoxins:

 \rightarrow aflatoxin M₁

incidence: 4/4, conc. range: 0.015-0.030 $\mu g\,/\,kg,\, \ensuremath{\varnothing}$ conc.: 0.020 $\mu g\,/\,kg,\, country:$ UK

incidence: 147/147*, conc. range: < 0.1-0.4 μ g / kg, country: UK, *home made \rightarrow citrinin

incidence: 2/2*, conc. $<100~\mu g\,/\,kg,$ country: UK, *1 mature English and 1 colored Scotch cheddar

 \rightarrow ochratoxin A

incidence: $2/2^*$, conc. range: $260-500 \mu g / kg$, country: UK, *1 mature English and 1 colored Scotch cheddar

Cheese, Chesire may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 36/36*, conc. range: < 0.1-0.4 μ g / kg, country: UK, *home made \rightarrow ochratoxin A incidence: 3/5*, conc. range: \leq 50 μ g / kg, country: UK, *colored, white and red

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Cheese, Chester

Cheese, Chester may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 4/4, conc. range: traces (1 sample), < 0.1 µg/kg (2 sa), > 0.1 µg/kg (1 sa), country: Germany

Cheese, Comte may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 1/279*, conc. range: 1.1 µg/kg, country: Japan, *imported

Cheese, Cottage may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 1/209, conc.: 0.08 µg/l*, country: USA, *1 = level reported on fluid milk basis incidence: 15/209, conc. range: 0.05-0.4 µg/kg, country: USA

Cheese, Cream may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 9/9, conc. range: 0.037-0.134 μ g/kg, Ø conc.: 0.79 μ g/kg, country: Denmark \rightarrow penitrem A incidence: 1/1*, conc. range: nc, country: USA, *visible moldy

Cheese, Danish Blue may contain the following \rightarrow mycotoxins: \rightarrow roquefortine C incidence: 3/3, conc. range: 950-1700 μ g/kg, Ø conc.: 1203 μ g/kg, country: Denmark

Cheese, Double Gloucester may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 10/10*, conc. range: < 0.1-0.15 µg/kg, country: UK, *home made \rightarrow ochratoxin A incidence: 2/2, conc. range: \leq 50 µg/kg, country: UK **Cheese, Edam** may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 31/32, conc. range: traces (11 samples), < 0.1 µg/kg (16 sa), > 0.1 µg/kg (4 sa), country: Germany incidence: 4/4, conc. range: 0.073-0.117 µg/kg, Ø conc.: 0.099 µg/kg, country: The Netherlands \rightarrow ochratoxin A incidence: 2/25, conc. range: 820-1100 µg/kg, Ø conc.: 960 µg/kg, country: Yugoslavia

Cheese, Edam Cake may contain the following \rightarrow mycotoxins: \rightarrow sterigmatocystin incidence: 2/66*, conc. range: 7.5-17.5 µg/kg, Ø conc.: 12.5 µg/kg, country: Czechoslovakia, *and different other kinds of cheese

Cheese, Emmental may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 15/358*, conc. range: 0.1-1.1 µg/kg, Ø conc.: 0.53 µg/kg, country: Japan *imported natural cheese \rightarrow ochratoxin A incidence: 3/3, conc. range: \leq 50 µg/kg, country: UK

Cheese, Fresh may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 27/80, conc. range: 0.1-0.51 μ g/kg, Ø conc.: 0.23 μ g/kg, country: Germany

Cheese, Goat may contain the following \rightarrow mycotoxins: \rightarrow patulin incidence: 1/18, conc.: 30 µg/kg, country: France. \rightarrow penicillic acid incidence: 2/18, conc. range: \leq 45,210 µg/kg, country: France **Cheese, Gorgonzola** may contain the following \rightarrow mycotoxins: \rightarrow mycophenolic acid incidence: 3/12, conc. range: 10-100 μ g/kg, country: France \rightarrow roquefortine C incidence: 2/2, conc. range: 490-940 μ g/kg, Ø conc.: 715 μ g/kg, country: Italy incidence: 2/2, conc. range: 150-190 μ g/kg, Ø conc.: 170 μ g/kg, country: Italy

Cheese, Gouda may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 9/9, conc. range: 0.039-0.087 µg / kg, Ø conc.: 0.063 µg / kg, country: The Netherlands \rightarrow sterigmatocystin incidence: 6*/6, conc. range: nc, country: The Netherlands, *surface layer

Cheese, Gouda & Cheddar may contain the following \rightarrow mycotoxins: \rightarrow cyclopiazonic acid incidence: nc, conc. range: 35,000-70,000 µg / kg, country: South Africa

Cheese, Grana Padano is a Parmesan-like cheese. Grana Padano may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 219/223, conc. range: 0.005-0.1 µg/kg (203 samples), 0.101-0.25 µg/kg (15 sa), > 0.25 µg/kg (1 sa), country: Italy

 $\begin{array}{ll} \mbox{Cheese, Lancashire} & may contain the following \rightarrow mycotoxins: \\ \rightarrow aflatoxin M_1 \\ \mbox{incidence: } 5/5^*, \mbox{ conc. range: } < 0.1\text{-}0.15 \\ \mbox{$\mu g / kg, country: UK, *home made} \end{array}$

Cheese, Leicester may contain the following \rightarrow mycotoxins:

→ aflatoxin M_1 incidence: 6/6*, conc. range: < 0.1-0.15 μ g / kg, country: UK, *home made → ochratoxin A incidence: 1/2, conc.: \leq 50 μ g / kg, country: UK

Cheese, Maribo may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 3/3, conc. range: 0.087-0.412 μ g/kg, Ø conc.: 0.264 μ g/kg, country: Denmark

Cheese, Mold-cured is a food product that has been consumed for centuries without causing any detrimental effects on human health. This is confirmed by long-term trials with \rightarrow Penicillium roquefortii Thom and → Penicillium camembertii Thom as well as Camembert (\rightarrow cheese, Camembert) and Blue cheese (\rightarrow cheese, Blue). No harmful effects could be demonstrated in experimental animals. Because of the mycotoxicological potential of the starter cultures the following points should be considered: i) the use of non-toxic starter cultures, ii) provision of optimal conditions during manufacture and ripening, iii) sanitary precautions to prevent unwarranted mold growth.

Cheese, Moravian Block may contain the following \rightarrow mycotoxins: \rightarrow sterigmatocystin incidence: 1/66, conc.: 7.5 µg/kg, country: Czechoslovakia

Cheese, Mozarrella may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 4/4, conc. range: 0.181-0.433 µg/kg, Ø conc.: 0.334 µg/kg, country: Denmark incidence: 5/5, conc. range: 0.028-0.252 µg/kg, Ø conc.: 0.091 µg/kg, country: Germany

Cheese, Parmesan

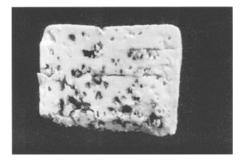
Cheese, Parmesan may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 18/200, conc. range: 0.035-0.190 µg/kg, country: Italy

Cheese, pepper may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ and / or \rightarrow aflatoxin G₁ incidence: 1/1, conc.: "high", country: France

Cheese, Romadur may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 35/50, conc. range: traces (19 samples), < 0.1 µg/kg (8 sa), > 0.1 µg/ kg (8 sa), country: Germany

Cheese, Roquefort may contain the following \rightarrow mycotoxins (see Figure Roquefort): \rightarrow mycophenolic acid incidence: 4/5, conc. range: 250-5000 $\mu g / kg$, \emptyset conc.: 3375 $\mu g / kg$, country: France \rightarrow roquefortine C incidence: 3/3, conc. range: 200-1330 $\mu g / kg$, \emptyset conc.: 670 $\mu g / kg$, country: France incidence: 21/25, conc. range: 10- \leq 15,000 $\mu g / kg$, country: France

Cheese, Samsoe may contain the following \rightarrow mycotoxins:



Roquefort. *Penicillium roquefortii* in Roquefort cheese

→ aflatoxin M_1 incidence: 5/5, conc. range: 0.07-0.504 µg/kg, Ø conc.: 0.214 µg/kg, country: Denmark

Cheese, Stilton may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 8/8*, conc. range: 0.1-0.3 μ g/kg, country: UK, *home made \rightarrow roquefortine incidence: 2/2, conc. range: 970-3400 μ g/kg, Ø conc.: 2185 μ g/kg, country: UK

Cheese, Swiss may contain the following \rightarrow mycotoxins: \rightarrow penicillic acid incidence: 4/33, conc. range: \leq 500 µg/kg, country: USA

Cheese, Tilsit Washing of a Tilsit cheese previously inoculated with \rightarrow Aspergillus flavus Link and \rightarrow Aspergillus parasiticus Speare, seems to cause \rightarrow aflatoxins to diffuse from the surface layer into the body of the cheese. Tilsit cheese may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1*/1, conc.: 7 µg/kg, country: Germany, *total: 9 µg aflatoxin / kg, incidence: 18/24, conc. range: traces (7 samples), < 0.1 µg/kg (10 sa), > 0.1 µg/kg (1 sa), country: Germany

Cheese, Wensleydale may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 5/5*, conc. range: < 0.1-0.2 µg/kg, country: UK, *home made \rightarrow ochratoxin A incidence: 1/1, conc.: \leq 50 µg/kg, country: UK

Cheese, Wine may contain the following \rightarrow mycotoxins:

 \rightarrow aflatoxin M₁ incidence: 3/7, conc. range: traces (2 samples), > 0.1 $\mu g \, / \, kg$ (1 sa), country: Germany

Cheese cake may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A incidence: 1*/3, conc.: 1075 µg/kg, country: Poland, *moldy

Cheese dressing, blue may contain the following \rightarrow mycotoxins: \rightarrow roquefortine C incidence: 2/2, conc. range: 18-72 µg/kg, \emptyset conc.: 45 µg/kg, country: USA

Cheese rind may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ and / or \rightarrow aflatoxin G₁ incidence: 6/34, conc. range: nc, country: Romania

Cheese trimmings (no specification) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ and / or \rightarrow aflatoxin G₁ incidence: 1/1, conc.: nc, country: USA \rightarrow ochratoxin A incidence: 1/1, conc.: nc, country: USA

Cherries (sweet) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/8*, conc.: 5 µg/kg, country: Germany, *moldy \rightarrow fruits

Chicken No natural contamination of Broiler-type chickens with \rightarrow aflatoxins has yet been reported. Feeding results indicate a rapid tissue clearance (4 days) after the removal of the aflatoxins from the diet although the \rightarrow mycotoxins were deposited in all tissues, especially gizzards, liver, and kidneys. However, contamination with \rightarrow ochratoxin A is evident. Chicken may contain the following mycotoxins: ochratoxin A incidence: 36/65, conc. range: ≤ 0.18 µg/kg, Ø conc.: 0.03 µg/kg, country: Denmark \rightarrow meat

Chicken, yolk may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A incidence: nc/nc, conc. range: 1.6-4 µg/kg, country: Germany

Chicken liver Feed tissue ratios of \rightarrow aflatoxin B₁ to AFB₁ and \rightarrow aflatoxin M₁ are much higher for kidney and liver than for muscle. The liver may contain the following \rightarrow mycotoxins: aflatoxin B₁ incidence: 1/5, conc.: < 5 µg/kg, country: Germany \rightarrow meat

Chilli \rightarrow Pepper (red) , \rightarrow spices

Chilli pickles may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: nc/4, conc. range: 1-58.5 μ g/kg, country: UK \rightarrow fumonisins (FB₁, FB₂) incidence: 1/4, conc.: 121 μ g/kg, country: UK \rightarrow ochratoxin A incidence: 3/4 conc. range: 0.5-1.2 μ g/kg, country: UK \rightarrow spices

Chilli powder may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: nc/4, conc. range: 1.1-5.4 µg / kg, country: UK \rightarrow diacetoxyscirpenol

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Chilli powder

incidence: nc/4, conc. range: 47-81 $\mu g / kg$, country: UK \rightarrow deoxynivalenol incidence: 1/4, conc.: 8 $\mu g / kg$, country: UK \rightarrow HT-2 toxin incidence: 1/4, conc.: 24 $\mu g / kg$, country: UK \rightarrow ochratoxin A incidence: nc/4, conc. range: 1.6-50.4 $\mu g / kg$, country: UK \rightarrow zearalenone incidence: nc/4, conc. range: 4.5-15.4 $\mu g / kg$, country: UK \rightarrow spices

Chilli sauce may contain the following \rightarrow mycotoxins: \rightarrow nivalenol incidence: 1/4, conc.: 15 µg/kg, country: UK \rightarrow ochratoxin A incidence: 1/4, conc.: 3.3 µg/kg, country: UK \rightarrow zearalenone incidence: 1/4, conc.: 7.1 µg/kg, country: UK \rightarrow spices

Chips → maize chips

Chocolate may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins incidence: 1*/36, conc.: 5 µg AFB₁ resp. 10 µg aflatoxins / kg, country: Germany, *containing \rightarrow Brazil nuts

Cider Due to alcoholic fermentation (*Saccharomyces cerevisiae*) cider is usually free of \rightarrow patulin. In Canada and the USA this term is also used for not fermented \rightarrow apple juice which can be misleading.

Cider may contain the following \rightarrow mycotoxins:

patulin incidence: 9/13, conc. range: 100-300 μ g/l, country: France \rightarrow apple juice

Cirrhosis Disease of the liver characterized by excessive \rightarrow fibrosis.

Citreoviridin is an unsaturated lactone (2,5-anhydro-1,6-dideoxy-2-c-[(1E,3E,5E,7E)-8-(4-methoxy-5-methyl-20x0-2H-pyran-6-yl)-2-methyl-1,3,5,7octatetraenyl]-4-c-methyl, \rightarrow mycotoxins) which was isolated in 1947 from \rightarrow Penicillium citreonigrum Dierckx (formerly *P. toxicarium*), a contaminant of yellow rice (see Figure Citreoviridin).

CHEMICAL DATA Empirical formula: $C_{23}H_{30}O_6$, molecular weight: 402

FUNGAL SOURCES

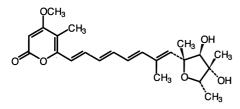
→ Penicillium spp. (e.g. *P. citreonigrum*, *P. miczynskii*, *P. manginii*, *P. smithii* (syn. *P. corynephorum*), *Eupenicillium ochrosal-moneum*, → Aspergillus terreus Thom

NATURAL OCCURRENCE

 \rightarrow pecans, probably in "yellow rice" Toxin formation on \rightarrow rice is favored by low temperatures and high humidity. These climatic conditions predominate in the northern part of Japan. Rice ("soft \rightarrow grains") grown in this area often shows a contamination with P. citreonigrum, a fungus that is a major source of this mycotoxin. Citreoviridin has also been isolated from naturally contaminated moldy pecan fragments (\rightarrow pecans) and from standing \rightarrow maize in the field (USA). The natural occurrence of this toxin in \rightarrow food has rarely been reported because adequate analytical methods and sources of standard for this mycotoxin are not available, generally.

Тохісіту

acute toxic, neurotoxic, paralytic, potent inhibitor of ATPase



Citreoviridin

The symptoms include early onset of a progressive \rightarrow paralysis in the extremities of laboratory animals. Similarly \rightarrow convulsions, vomiting and impairment of the respiratory center occurred. In a later stage, the disease is characterized by \rightarrow hypothermia, flaccid paralysis and cardiovascular disturbances. Along with \rightarrow dyspnea, gasping and coma respiratory arrest leads to death. These symptoms are very similar to those in human patients who consume rice as a staple food and suffer from \rightarrow acute cardiac beriberi.

LD₅₀ (po): 3.6 mg/kg bw rat Detection

TLC

Possible Mycotoxicosis This highly toxic fungal metabolite is associated in the complex of \rightarrow yellow rice disease in Japan and represents a (possible) causative agent in acute cardiac beriberi in humans.

Citrinin (Syn.: antimycin, monascidin A) is a (3R-*trans*)-4,6-dihydro-8-hydroxy-3,4,5-trimethyl-6-oxo-3H-2-benzopyran-7-carboxylic acid which is derived from the condensation of five acetate and the introduction of three one-carbon units (see Figure Citrinin). This major "yellow rice" toxin (\rightarrow yellow rice disease) was first isolated from \rightarrow Penicillium citrinum Thom in 1931. The mold occurs most frequently in "yellow rice" and produces copious quantities of this yellow toxic metabolite. Citrinin, therefore, was first implicated in the "yellow rice" syndrome in Japan.

CHEMICAL DATA

Empirical formula: $C_{13}H_{14}O_5$, molecular weight: 250

FUNGAL SOURCES

e.g. \rightarrow Aspergillus spp. (e.g. \rightarrow Aspergillus candidus Link, *A. carneus*, \rightarrow Aspergillus terreus Thom), *Monascus purpureus*, *M. ruber*, \rightarrow Penicillium spp. (e.g. \rightarrow Penicillium citreonigrum Dierckx, \rightarrow Penicillium citrinum Thom, \rightarrow Penicillium expansum Link, \rightarrow Penicillium verrucosum Dierckx chemotype II).

NATURAL OCCURRENCE

 \rightarrow Acacia concinna, \rightarrow bakery products, \rightarrow barley, \rightarrow bread, \rightarrow cardamom, \rightarrow Cassia fistula, \rightarrow cereals, \rightarrow cheese, \rightarrow cheese, Cheddar, \rightarrow confectionery, \rightarrow coriander, \rightarrow cumin, \rightarrow fennel, \rightarrow flour, \rightarrow Hydnocarpus laurifolia, \rightarrow maize, \rightarrow maize flour, \rightarrow meat, \rightarrow oil seed rape, \rightarrow pastries, \rightarrow peanuts, \rightarrow pepper, \rightarrow pig kidneys, \rightarrow Piper betle, \rightarrow rice, \rightarrow rye, \rightarrow shoyu, \rightarrow triticale, \rightarrow tumeric, \rightarrow wheat, \rightarrow wheat grits Citrinin mainly occurs in rice and other cereals. Different kinds of foodstuff, especially \rightarrow grains, often are contaminated with both citrinin and \rightarrow ochratoxin A. Since citrinin is more readily lost in analytical procedures, it seems to occur much less frequently than ochratoxin A. In general, significantly higher citrinin concentrations, compared to OTA levels, occur. Although citrinin represents a contaminant of different kinds of food products, it seems unlikely that it does constitute a human health problem.

Тохісіту

Fetotoxic, embryocidal, \rightarrow mutagenic (?) and mildly \rightarrow teratogenic, nephrotoxic, hepatotoxic antibacterial, antifungal, antiprotozal,

phytotoxic

Citrinin

In the view of kidney damage and the development of \rightarrow renal tumors, a probable synergistic effect with ochratoxin A is important.

 LD_{50} (po): 50 mg/kg bw rats

DETECTION

HPLC, NMR, spectrofluorometric determination, TLC

Possible Mycotoxicosis

 \rightarrow Mycotoxic porcine nephropathy, \rightarrow Balkan endemic nephropathy (citrinin and ochratoxin A); \rightarrow Yellow rice disease (citrinin, \rightarrow citreoviridin, other \rightarrow Penicillium toxins)

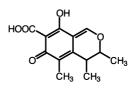
FURTHER COMMENTS

Citrinin was mainly located in the spore wall and may be a major component of the spores of *P. verrucosum*. It was suggested that this mycotoxin, which is released in an aqueous environment, may have important function(s) in spore survival.

Stability: Citrinin was fairly stable in air or oven dried whole \rightarrow maize kernels inoculated with \rightarrow Penicillium spp. over a period of a year. However, during mashing this mycotoxin is degraded and therefore, \rightarrow beer is citrinin-free. Because citrinin is more heat sensitive than OTA, heat treament of contaminated food will significantly reduce the citrinin level. The instability of citrinin may explain its absence from \rightarrow apple juice and other \rightarrow apple products. However, heating with water (ca. 140 °C) yielded a decomposition product as toxic as or even more toxic than citrinin.

Citrinin ist unstable during prolonged exposure to light or heat.

Milling: Compared to the milled product, citrinin is accumulated in the bran and polish fraction of rice. A positive correlation between highly contaminated samples and levels found in the aforementioned fractions could be established. Citrinin probably survives milling at least to some extent because maize flour (e.g.



Citrinin

Thailand) was contaminated in the range of 10-98 μ g/kg.

Production: Citrinin production (\rightarrow Pencillium viridicatum Westling) occurred on bread at a minimum \rightarrow a_w of 0.80, optimum a_w 0.92. Durum wheat (15% and 19% moisture) enabled citrinin production after 24 weeks with maximal production after 48 weeks. Citrinin formation may occur on every kind of meat and therefore should be regarded as a serious toxin.

Cladosporium anamorphic \rightarrow Mycosphaerellaceae, teleomorphs *Mycospherella*, *Venturia*

Cladosporium spp. may grow on chilled and overwintered grain. C. herbarum together with C. fagi may be associated with some forms of \rightarrow alimentary toxic aleukia. The \rightarrow mycotoxins epicladosporic and fagicladosporic acid (see Figure Cladosporium) may be responsible for the toxicity of \rightarrow grains which have been exposed to cold winter climatic conditions since they are frequently infected by these two fungi.

Clavacin (Syn.: \rightarrow Patulin)

Clavatin (Syn.: \rightarrow Patulin)

Cladosporium. Epicladosporic acid and fagicladosporic acid **Claviceps** \rightarrow Clavicipitaceae Fungi of this genus grow parasitically in the spikes of \rightarrow cereals, especially \rightarrow rye, and grasses. During overwintering 2 - 4 cm long granules are formed, called \rightarrow ergots. The most important species is *C. purpurea* which mainly infects \rightarrow rye. Minor infections also occurred on \rightarrow barley, \rightarrow maize, \rightarrow oats, and \rightarrow wheat. \rightarrow Clavine alkaloids, \rightarrow ergot alkaloids, \rightarrow ergotism, ergots

Clavicipitaceae → Hypocreales

Claviformin (Syn.: \rightarrow patulin)

Clavine alkaloids In contrast to the wellknown lysergic acid derivatives (\rightarrow ergot alkaloids), the carboxyl group has been reduced to a hydroxymethyl or a methyl group. \rightarrow Sclerotia of \rightarrow Claviceps species which occur on wild grasses in Africa and in the Far East contain substantial amounts of these alkaloids. Only trace amounts are found in the sclerotia and saprophytic cultures of *C. purpurea* and *C. paspali*. Important clavine alkaloids are e.g. fumigaclavine A & B.

Cocoa beans In the Central American countries like Costa Rica, outdoor drying of cocoa beans on movable rail and wheel beds (ca. 10 m^2) is the usual practice. Pushing the cocoa beans under a crude roof and storage in a layer-type fashion is a good protection against the rain. Too-wet stored or rewetted cocoa beans are prone to mold growth and subsequent mycotoxin contamination. A significant destruction of ochratoxin A occurred during the processing of cocoa beans to dark \rightarrow chocolate. Cocoa beans may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁

incidence: 1/40*, conc.: 5 μ g / kg, country: Norway, *imported

 \rightarrow aflatoxin B₂ incidence: 1/40*, conc.: traces, country: Norway, *imported \rightarrow aflatoxin G₁ incidence: $1/40^*$, conc.: $4 \mu g / kg$, country: Norway, *imported \rightarrow aflatoxin G₂ incidence: 1/40*, conc.: traces, country: Norway, *imported \rightarrow aflatoxin incidence: 3/91, conc. range: 2-20 µg/kg (1 sample), > 20 μ g/kg (2 sa), country: Uruguay \rightarrow aflatoxins (no specification) incidence: 2*/47, conc. range: 5-9.9 μ g / kg, country: Canada, *AFB₁, AFB₂, AFG₁, AFG₂ incidence: 1/14, conc.: > 4 μ g/kg, country: Ghana incidence: 1/6, conc.: > 4 μ g/kg, country: Malaysia incidence: 5/14, conc. range: > 4 μ g/kg, country: Nigeria incidence: 4/6, conc. range: > 4 μ g / kg, country: Papua New Guinea incidence: 2/9, conc. range: $\leq 17 \,\mu\text{g}/\text{kg}$, country: Philippines incidence: 1/4, conc.: > 4 μ g/kg, country: Trinidad \rightarrow ochratoxin A incidence: 2/3, conc. range: > 60 μ g/kg, country: Ecuador incidence: 4/14, conc. range: > 60 μ g/kg, country: Ghana incidence: 2/5, conc. range: > 60 μ g/kg, country: Grenada incidence: 1/2, conc.: > 60 µg/kg, country: Ivory Coast incidence: 1/6, conc.: > 60 μ g/kg, country: Malaysia incidence: 1/14, conc.: > 60 μ g/kg, country: Nigeria incidence: 1/1, conc.: > 60 μ g/kg, country: Venezuela \rightarrow coffee beans

Cocoa beans (raw)

Cocoa beans (raw) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins (no specification) incidence: 7/56, conc.: < 5 µg/kg (6 samples), 5-10 µg/kg (1 sa), country: UK \rightarrow ochratoxin A incidence: 10/56, conc. range: < 100 µg/kg (5 samples), 101-200 µg/kg (4 sa), 201-500 µg/kg (1 sa), country: UK

Cocoa beans (roasted) may contain the following \rightarrow mycotoxins: aflatoxin (no specification) incidence: 6/8, conc. range: $\leq 28 \ \mu g / kg$, \emptyset conc.: 18 $\mu g / kg$, country: Philippines \rightarrow aflatoxins (no specification) incidence: 7/19, conc.: $< 5 \ \mu g / kg$ (6 samples), 5-10 $\mu g / kg$ (1 sa), country: UK \rightarrow ochratoxin A incidence: 3/19, conc. range: 100 $\mu g / kg$, country: UK

Cocoa nibs may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins (no specification) incidence: 1/2, conc.: 11-20 µg/kg, country: UK \rightarrow ochratoxin A incidence: 1/2, conc.: 101-200 µg/kg, country: UK

Cocoa presscake may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins (no specification) incidence: 1/4, conc.: < 5 µg/kg, country: UK \rightarrow ochratoxin A incidence: 1/4, conc.: 101-200 µg/kg, country: UK

Cocoa products may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A incidence: 1/20, conc.: $\leq 0.6 \ \mu$ g/kg, country: Germany

Coconut (processed)

Coconut is an excellent medium for the growth of \rightarrow Aspergillus spp. and subsequent aflatoxin accumulation. Coconut may contain the following \rightarrow mycotoxins: aflatoxin (no specification) (\rightarrow aflatoxins) incidence: 20/29, conc. range: ≤ 26 µg/kg, Ø conc.: 11 µg/kg, country: Philippines \rightarrow nuts

Coconut ice may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins (no specification) incidence: 1/4, conc. range: nc, country: UK

Coconut oil (crude)

derived from moldy coconut may contain high levels of \rightarrow aflatoxins and even commercially avaiable coconut oil from processed \rightarrow copra may be contaminated by low to medium aflatoxin levels. Only by refining can the aflatoxin and the pigments be removed from the \rightarrow oil but the expense of this method limits its use in poorer countries. Contaminated oil may effectively be decontaminated by exposure to sunlight.

Coconut oil may contain the following \rightarrow mycotoxins:

aflatoxin (no specification)

incidence: 3/3, conc. range: $\leq 9 \,\mu g / kg$, Ø conc.: 3 $\mu g / kg$, country: Philippines \rightarrow oil, \rightarrow olive oil, \rightarrow peanut oil, \rightarrow sunflower seed oil

Coffee The manufacture of coffee cherry includes several steps: harvesting, direct drying or pulping/fermentation and drying, hulling, cleaning, sorting = producer country; decaffeination (alternatively), blending, roasting, industrial extraction (alternatively), packaging = producer or consumer country

Coffee may be an important contributor to \rightarrow ochratoxin A (\rightarrow Aspergillus ochra-

ceus group) intake ($\approx 20\%$) in humans. A mean level of 0.5-1.5 µg OTA / kg has been detected in the roasted coffee sold on the EU market. Transmission of OTA to the final brew is possible.

Mycotoxin contamination of the beans mainly occurs during green coffee processing, and/or transportation. It seems that superficial OTA contamination is higher than deep bean contamination. Together with the chaff this portion is eliminated during roasting. Industrial decaffeination may cause a 60

% reduction of ochratoxin A in a naturally-contaminated sample. During roasting, as well as during brewing, partial to almost complete OTA destruction has been observed.

Although OTA levels as low as $0.1 \ \mu g / kg$ coffee can now be easily detected, detection of single contaminated beans is difficult because of the extremely inhomogenous distribution of the mycotoxin in the batch. A suitable sampling procedure for OTA detection in green coffee is lacking. Highly contaminated batches of green coffee possess musty / moldy off-flavors which are carried through to the finished product and beverage. Because such batches are rejected by the coffe trade,the amount of OTA contamination in commercial roast, ground and instant coffee products is usually low.

The daily intake of four cups of coffee (24 g roasted & ground) contributes on average 19 ng OTA / day, 8 g instant coffee = 10 ng OTA / day. The resulting weekly OTA consumption constitutes not more than 2% of the PTWI of 100 ng / kg set by the Joint FAO / WHO Expert Committee on Food Additives.

Coffee may contain the following

 \rightarrow mycotoxins:

 \rightarrow ochratoxin A

incidence: 7/22*, conc. range: 0.2-4 μ g/kg, country: Australia, *pure soluble

incidence: 2/2*, conc. range: 1.9-4.8 µg/kg, country: Czech Republic, *pure soluble incidence: $2/2^*$, Ø conc.: 1.6 µg/kg, country: Czech Republic, *adulterated soluble incidence: 11/11*, conc. range: ≤ 3.2 μ g / kg, Ø conc.: 0.51 μ g / kg, country: Denmark, *roasted incidence: 20*, conc. range: 0-5.5 µg/kg, \emptyset conc.: 1.1 µg/kg, country: Europe**, *instant, decaffeinated incidence: 10*, conc. range: 0-1 µg/kg, \emptyset conc.: 0.5 μ g/kg, country: Europe**, *instant, mixed incidence: 119*, conc. range: 0-27.2 μ g/kg, Ø conc.: 1.4 μ g/kg, country: Europe**, *instant, regular incidence: 39*, conc. range: 0-2.8 µg/kg, Ø conc.: 0.7 μ g/kg, country: Europe**, *roasted and ground, decaffeinated incidence: 445*, conc. range: 0-8.2 µg/kg, \emptyset conc.: 0.8 μ g/kg, country: Europe**, *roasted and ground, regular ** collaborative study of different European countries incidence: 2/4* **, conc. range: 10-90 μ g/kg, Ø conc.: 50 μ g/kg, country: Germany, *moldy, **raw incidence: $4/14^*$, conc.: $\leq 4.9 \ \mu g / kg$, country: Germany, *roasted incidence: 25/30*, conc. range: \leq 4.9-7.54 μ g/kg, Ø conc.: 1.43 μ g/kg, country: Germany, *roasted incidence: $1/29^*$, conc.: $3 \mu g/kg$, country: Germany, *roasted and raw incidence: 5/9*, conc. range: 0.3-2.2 μg/kg, country: Germany, *pure soluble incidence: 6/6*, conc. range: 0.5-1.6 μ g / kg, country: Greece, *pure soluble incidence: 14/14, conc. range: 0.5-6.5 µg/kg, country: Hungary, *pure soluble incidence: 1/1, Ø conc.: 1.2 µg/kg, country: Hungary, *adultered soluble incidence: 2/7* **, conc. range: 3.2-4.4 μ g/kg, Ø conc.: 3.8 μ g/kg, country: Indonesia, *commercial, **roasted (a total of 68 samples has been investigated

Coffee

in Japan, 5 samples (2 from Indonesia, 3 from Yemen) contained OTA incidence: 5/68, conc. range: 3.2-17 μg/kg, country: Japan incidence: 21/22, conc. range: 0.2-3.5 μ g / kg, country: Russia, *pure soluble incidence: $12/12^*$, Ø conc.: 6.93 µg/kg, country: Russia, *adulterated soluble incidence: 6/6, conc. range: 0.3-3.6 μg/kg, country: Salvador, *pure soluble incidence: 4/4, conc. range: 1.5-5.3 μg/kg, country: Slovakia, *pure soluble incidence: 16/40*, conc. range: 1-7.8 μg/kg, country: Switzerland, *brew incidence: 2/3, conc. range: 0.2-0.3 μg/kg, country: Switzerland, *pure soluble incidence: 3/3, conc. range: 1.3-1.9 μ g / kg, country: Thailand, *pure soluble incidence: 64/80*, conc.: 0.1-8.0 µg/kg, country: UK, *soluble incidence: 17/20*, conc. range: 0.2-2.1 μg/kg, country: UK, *roasted and ground, regular incidence: 2/4, conc. range: 0.3-0.4 μ g / kg, country: unknown incidence: 9/13*, conc. range: 0.1-1.2 μ g/kg, Ø conc.: 0.41 μ g/kg, country: USA, *import from South America incidence: 3/6, conc. range: 1.5-2.1 μ g/kg, country: USA, *pure soluble incidence: 3/10* **, conc. range: 6.5-17 μ g/kg, Ø conc.: 10.1 μ g/kg, country: Yemen, *commercial, **roasted (a total of 68 samples has been investigated in Japan, 5 samples (2 from Indonesia, 3 from Yemen) contained OTA

Coffee beans (green)

may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin (no specification) incidence: 2/201, conc. range: 3-12 µg/kg Ø conc.: 7.5 µg/kg, country: USA \rightarrow ochratoxin A incidence: 1/1, conc.: 8 µg/kg, country: Austria

incidence: 3/7, conc. range: ca. 20-360 μg/kg, country: Brazil incidence: 17/139, conc. range: ca. 20 μ g / kg (13 samples), 35 μ g / kg (2 sa), 50 μ g/kg (2 sa), country: Colombia incidence: 1/1*, conc.: 0.5 µg/kg, country: India, *commercial incidence: 2/2*, conc. range: 0.5-1 µg/kg, \emptyset conc.: 0.75 µg/kg, country: Indonesia, *commercial incidence: 2/2, conc. range: $\leq 2.2 \ \mu g / kg$, country: Ireland incidence: 19/29*, conc. range: 0.2-15 μg/kg, country: Italy, *commercial incidence: 3/68, conc. range: 20-80 μ g/kg, Ø conc.: 40 μ g/kg, country: Italy incidence: 4/5*, conc. range. < 20-400 μg/kg, country: Italy, *molded incidence: 9/40*, conc. range: 0.5-23 μg/kg, country: Italy, *commercial incidence: 1/12, conc.: ca. 20 μg/kg, country: Ivory Coast incidence: 1/3*, conc.: 3.8 µg/kg, country: Ivory Coast, *commercial incidence: 4/22, conc. range: 9.8-46 μg/kg, country: Japan incidence: 1/1*, conc.: 1.8 µg/kg, country: Kenia, *commercial incidence: 13/25*, conc. range: 1.2-56 μg/kg, country: Switzerland, *commercial incidence: 7/7, \emptyset conc.: 3.9 µg/kg, country: Thailand incidence; 2/14, conc. range: \leq 7 µg/kg, country: The Netherlands incidence: 1/1*, conc.: 5.2 µg/kg, country: Togo, *commercial incidence: 1/2, conc.: ca. 20 µg/kg, country: Uganda incidence: 2/2*, conc. range: 1.5-23 μg/kg, country: Uganda, *commercial incidence: 9/31*, conc. range: < 10-200 μg/kg, country: UK, *commercial incidence: 2/201, conc. range: 24-96 μ g/kg, Ø conc.: 60 μ g/kg, country: USA incidence: 19/267*, conc. range: 20-360 μg/kg, country: USA, *imported, handcleaned coffee beans

incidence: $3/68^*$, conc. range: tr-80 µg/kg, country: USA, *imported, commercial incidence: $9/19^*$, conc. range: 0.1-4.6 µg/kg, Ø conc.: 1.41 µg/kg, country: USA, *import form South America incidence: 1/2, conc.: 1.3 µg/kg, country: Zaire \rightarrow sterigmatocystin incidence: 1*/502, conc.: 1200 µg/kg, country: Italy, *very moldy incidende: 1*/2, conc.: 1143 µg/kg, country: South Africa, *condemned as unfit for human consumption \rightarrow cocoa beans

Comte cheese \rightarrow cheese, Comte

Confectionery may contain the following \rightarrow mycotoxins: \rightarrow citrinin incidence: 1/1, conc.: < 100 µg/kg, country: UK \rightarrow ochratoxin A incidence: 1/1, conc.: traces, country: UK \rightarrow marzipan, \rightarrow nuts, \rightarrow persipan

Congestion having an abnormal accumulation of blood.

Congressbele is an Indian peanut $(\rightarrow \text{ peanuts})$ based spiced snack which consists of the kotyledons of the groundnuts. After light frying in small quantity of oil the kotyledons are spiced with \rightarrow turmeric powder, \rightarrow pepper and salted. A lower aflatoxin contamination, compared to \rightarrow bondakaledkai, may result from a certain degree of cleaning from infested seeds. Congressbele may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 9/41, conc. range: 6-1100 μg/kg, country: India \rightarrow aflatoxin B₂

incidence: 5/41, conc. range: 4-700 μ g / kg, country: India \rightarrow groundnut toffee, \rightarrow bondakaledkai

Convulsions Violent irregular movement of a limb or limbs, or of the body, caused by contraction of muscles.

Cookies may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 35 products analysed, Ø conc.: 120 µg/kg, country: Canada \rightarrow biscuits, \rightarrow cereals **Copra** (and copra meal) contained the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 10/16, conc. range: 10-100 μ g / kg, Ø conc.: 39 μ g / kg, country: Germany \rightarrow aflatoxin B₂ incidence: 3/16, conc. range: 5-10 µg/kg, Ø conc.: 8.3 µg/kg, country: Germany aflatoxin (no specification) incidence: 129/182, conc. range: \leq 513 μ g / kg, Ø conc.: 39 μ g / kg, country: Philippines \rightarrow aflatoxins (no specification) incidence: 7/105, conc. range: 30-120 μ g / kg, Ø conc.: 42.8 μ g / kg, country: India incidence: 63/72*, conc. range: tr-200 μ g/kg, Ø conc.: 46 μ g/kg, country: USA, incidence: 10/16*, conc. range: 10-100 μ g / kg, Ø conc.: 37 μ g / kg, country: USA, *imported ochratoxin A incidence: 1/384, conc.: 50 µg/kg, country: India \rightarrow coconut, \rightarrow nuts

Coriander may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/15, Ø conc.: 8 µg/kg, country: Egypt

Coriander

incidence: 2/10, conc. range: tr-5.2 μg/kg, country: Germany incidence: 6/9, conc. range: 25-230 μg/kg, country: India incidence: 3/10, conc. range: 19-37 μ g/kg, Ø conc.: 25.7 μ g/kg, country: India incidence: 1/9, conc.: 45.5 µg/kg, country: Morocco \rightarrow aflatoxin B₂ incidence: 6/9, conc. range: 20-72 µg/kg, country: India incidence: 1/10, conc.: 5 µg/kg, country: India \rightarrow aflatoxin G₁ incidence: 1/15, Ø conc.: 2 µg/kg, country: Egypt incidence: 6/9, conc. range: 13-40 µg/kg, country: India incidence: 3/10, conc. range: 3-4 µg/kg, Ø conc.: 3.7 μ g/kg, country: India \rightarrow aflatoxin G₂ incidence: 6/9, conc. range: 14-35 µg/kg, country: India \rightarrow aflatoxins (no specification) incidence: 4/10, conc. range: 10-75 μg/kg, country: India incidence: 1/3*, conc.: 0.7 µg/kg, country: UK, *AFB₁, AFB₂, AFG₁, AFG₂ \rightarrow citrinin incidence: 1/9, conc.: 34 µg/kg, country: India \rightarrow deoxynivalenol incidence: 1/4, conc.: 21 µg/kg, country: UK \rightarrow ochratoxin A incidence: 1/9, conc.: nc, country: India incidence: 1/3, conc.: 4 µg/kg, country: UK \rightarrow zearalenone incidence: 1/9, conc.: nc, country: India incidence: nc/4, conc. range: 3.6-6.7 μ g / kg, country: UK \rightarrow spices

Corn \rightarrow Maize

Corn flakes may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/2, conc.: < 5 μ g/kg, country: UK \rightarrow fumonisin B₁ incidence: 1/2, conc.: 10 μ g / kg, country: Italy incidence: 2/12, conc. range: 50-100 $\mu g/kg$, Ø conc.: 60 $\mu g/kg$, country: Spain incidence: 1/12, con.: 55 µg/kg, country: Switzerland incidence: 4/17, conc. range: 140-1281 μ g/kg, Ø conc.: 497 μ g/kg, country: Thailand incidence: 1/5, conc.: 1430 µg/kg, country: The Netherlands \rightarrow fumonisin B₂ incidence: 3/17, conc. range: 120-466 μ g/kg, Ø conc.: 166 μ g/kg, country: Thailand \rightarrow fumonisins incidence: 8/8, conc. range: < 20-760 μ g / kg, country: Germany and unknown origin incidence: 4/6, conc. range: \leq 400 µg/kg, country: USA → ochratoxin A incidence: 2/13, conc. range: 0.1-0.19 μg/kg, country: Germany incidence: 1/34, conc.: 0.4 µg/kg, country: Germany \rightarrow sterigmatocystin incidence: 1/2, conc.: nc, country: UK \rightarrow zearalenone incidence: 1/1, conc. range: 13-20 µg/kg, country: Canada \rightarrow cereal flakes, \rightarrow maize flakes, \rightarrow oat flakes

Cortex Outer layer of an organ.

Cow After oral dosing, the residues of \rightarrow aflatoxin B₁ and \rightarrow aflatoxin M₁ can be found in the liver and kidneys for up to 7 days. After withdrawal from the contami-

nated diet, the cattle tissue was completely free of \rightarrow aflatoxins within 18 days. Probably, a longer withdrawal period is necessary for the cow's meat than for \rightarrow pork.

In vitro and in vivo studies show a rapid detoxification of \rightarrow ochratoxin A in ruminants by the action of rumen inherent proteolytic enzymes that cleave phenylalanine from the isocoumarin of the OTA molecule.

 \rightarrow meat

Cow kidney Feeding experiments with two milking cows (317-1125 $\mu g \rightarrow$ ochratoxin A/kg feed for 11 weeks) resulted in the contamination of the kidneys of one of the cows (5 μg OTA/kg). Neither ochratoxin α nor OTA was found in any tissue or in \rightarrow milk.

Cowpeas may contain the following \rightarrow mycotoxins: aflatoxin (no specification) (\rightarrow aflatoxins) incidence: 10/16, conc. range: \leq 86 μ g/kg, Ø conc.: 16 μ g/kg, country: Philippines \rightarrow ochratoxin A incidence: 5/31, Ø conc.: 34 μ g/kg, country: Senegal \rightarrow beans, \rightarrow cabbage, \rightarrow lentils, \rightarrow peas, \rightarrow pigeon peas, \rightarrow soybeans, \rightarrow vegetables **Crackers** may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol

incidence: 20 products analysed, \emptyset conc.: 270 µg/kg, country: Canada \rightarrow cereals

Cranberries may contain the following \rightarrow mycotoxins: \rightarrow patulin incidence: nc, conc. range: $\leq 265 \ \mu g / kg$, country: Sweden \rightarrow fruits Cream (full)

may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 19/28, conc. range: tr- > 2 μ g/kg, country: Germany \rightarrow milk

Cream cheese \rightarrow cheese, cream

Croissant butter may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 8/8, conc. range: 326-648 μ g/kg, Ø conc.: 453 μ g/kg, country: Argentina \rightarrow cereals, \rightarrow milk

Croissant fat may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 4/5, conc. range: 336-563 μ g/kg, Ø conc.: 377 μ g/kg, country: Argentina \rightarrow cereals, \rightarrow milk

Cumin may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 2/20*, conc. range: 0.29-0.96 μ g / kg, Ø conc.: 0.625 μ g / kg, country: Egypt, *different \rightarrow spices incidence: 2/8, conc. range: 24-104 μ g/kg, Ø conc.: 64 μ g/kg, country: India \rightarrow aflatoxin B₂ incidence: 2/8, conc. range: 12-78 µg/kg, \emptyset conc.: 45 µg/kg, country: India \rightarrow aflatoxin G₁ incidence: 2/8, conc. range: 8-45 µg/kg, Ø conc.: 26.5 μ g/kg, country: India \rightarrow aflatoxin G₂ incidence: 1/8, conc.: 30 µg/kg, country: India \rightarrow citrinin incidence: 1/8, conc.: 22 µg/kg, country: India \rightarrow spices

Curcuma

Curcuma may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 6/7, conc. range: < 2.5-3.8 μ g / kg, country: Canada \rightarrow spices

Curry may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 10/29*, conc. range: 1-3.9 μ g / kg (8 samples), 4-10 μ g / kg (2 sa), country: UK, *imported incidence: nc/3**, conc. range: 0.8-61.2 μg/kg, country: UK incidence: 1/3***, conc.: 0.4 µg/kg, country: UK → diacetoxyscirpenol incidence: 1/3****, conc.: 25 µg/kg, country: UK \rightarrow fumonisins (FB₁, FB₂) incidence: nc/3**, conc. range: 15-16 μg/kg, country: UK incidence: 1/3****, conc.: 230 µg/kg, country: UK \rightarrow fusarenon X incidence: 1/3****, conc.: 7 µg/kg, country: UK \rightarrow HT-2 toxin incidence: 1/3****, conc.: 17 µg/kg, country: UK → neosolaniol incidence: 1/3****, conc.: 9 µg/kg, country: UK \rightarrow nivalenol incidence: nc/3**, conc. range: 15-50 μ g / kg, country: UK incidence: nc/3***, conc. range: 9-67 μ g/kg, country: UK incidence: 1/3***, conc.: 14 µg/kg, country: UK \rightarrow ochratoxin A incidence: 3/3, conc. range: 5-33 µg/kg, country: Austria incidence: 2/3**, conc. range: 2.3-21.3 μ g/kg, Ø conc.: 11.8 μ g/kg, country: UK incidence: nc/3***, conc. range: 1.8-9.4 μg/kg, country: UK incidence: nc/3****, conc. range: 1.2-5.4 μg/kg, country: UK incidence: 4/4, conc. range: \leq 4.9-5.4 μg/kg, country: UK \rightarrow T-2 toxin incidence: 1/3****, conc.: 13 µg/kg, country: UK \rightarrow zearalenone incidence: nc/3**, conc. range: 1.2-10.8 μg/kg, country: UK incidence: 1/3****, conc.: 5.2 µg/kg, country: UK **curry powder hot, ***curry powder mild, ****mixes \rightarrow spices

Curry paste may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 1/4, conc.: 1.2 µg/kg, country: UK \rightarrow fumonisins (FB₁, FB₂) incidence: 1/4, conc.: 56 μ g / kg, country: UK \rightarrow nivalenol incidence: nc/4, conc. range: 5-16 µg/kg, country: UK \rightarrow ochratoxin A incidence: 4/4, conc. range: 0.6-15.5 μ g / kg, country: UK \rightarrow zearalenone incidence: 4/4, conc. range: 3.1-4.2 μ g/kg, country: UK \rightarrow spices

Cyclopiazonic acid (Abbr.: CPA) is an indole-tetramic acid (6a,7,11a,11b-tetrahydro-10-(1-hydroxyethylidene)-7,7-dimethyl-6H-pyrrolol[1',2':2,3]isoin-dolo[4,5,6-cd]indole-9,11-(2H,1OH)-dione) that was first isolated from \rightarrow Penicillium aurantiogriseum Dierckx in 1968 (see Figure Cyclopiazonic acid).

CHEMICAL DATA

molecular formula: $C_{20}H_{20}N_2O_3$, molecular weight: 336

FUNGAL SOURCES

e.g. A. flavus, \rightarrow Aspergillus oryzae (Ahlburg) Cohn, \rightarrow Aspergillus versicolor (Vuill.) Tiraboshi, \rightarrow Aspergillus spp., P. aurantiogriseum (also produces cyclopiazonic acid imine and bissecodehydrocyclopiazonic acid), \rightarrow Penicillium camembertii Thom (consistent producer), \rightarrow Penicillium commune Thom, \rightarrow Penicillium roquefortii Thom, \rightarrow Penicillium spp. P. aurantiogriseum (P. cyclopium) was previously known to be the most important CPA producer of the genus Penicillium. Because all CPA-producing strains of P. aurantiogriseum have now been assigned to P. commune this Penicillium species is currently regarded as being the most prominent CPA producer on natural substrates. Aflatoxin synthesis of \rightarrow Aspergillus flavus Link is often accompanied by similar production of CPA. The importance of A. flavus as CPA producer should therefore not be underestimated (\rightarrow turkey "X" disease).

NATURAL OCCURRENCE

 \rightarrow cheese, \rightarrow cheese, Camembert, \rightarrow cheese, Camembert & Brie, \rightarrow cheese, Gouda & Cheddar, \rightarrow kodo millet, \rightarrow maize, \rightarrow peanuts, \rightarrow sunflower seeds. Co-contamination of peanuts and maize with aflatoxin has been reported. CPA has been detected in the \rightarrow milk of lactating ewes within one day after experimental application. Presence of CPA in the milk was obvious even several days after withdrawal of the mycotoxin. With the exception of manufacturing unsweetened condensed milk (reduction ca. 40%) storing (4 °C) and processing caused only a minor decrease in CPA levels. In addition, CPA proved to be quite stable in \rightarrow fermented products.

TOXICITY

Necrotic (liver, gastrointestinal tissue, kidneys, skeletal muscles), carcinogenic, neurotoxic, \rightarrow mutagenic (Ames test). In humans clinical symptoms such as tremors (\rightarrow tremorgenic mycotoxins), sleepiness and giddiness have been observed. LD₅₀ (po): 36 and 63 mg/kg bw male and female rats, respectively.

DETECTION

capillary electrophoresis, colorimetic and spectrophotometry technique, ELISA, GC, HPLC (normal- and reversed-phase, ligand exchange), TLC

Possible Mycotoxicosis

Besides the \rightarrow aflatoxins, this mycotoxin is involved in the turkey "X" disease. An additive effect with aflatoxin has been demonstrated.

Implication of this mycotoxin in \rightarrow Kodua poisoning, a human malady in India, caused by the ingestion of kodo \rightarrow millet seeds invaded by *Aspergillus* has been suggested.

FURTHER COMMENTS

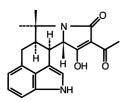
Production: CPA formation occurred in the range of 4 °C (refrigerator temperature), 13 °C (ripening temperature for cheeses), 25 °C (optimal).

The minimum $\rightarrow a_w$ for CPA production on maize was $a_w 0.90$ at 30 °C ($a_w 0.85$ yeast extrat agar*). Largest amounts were produced at $a_w 0.98$ at 20 °C (optimum $a_w 0.996$ yeast extract agar*) (\rightarrow Penicillium commune Thom*, \rightarrow Aspergillus flavus Link).

Stability: Simulation of the heat-treatments used by the dairy industry caused no considerable degradation of CPA in the milk. \rightarrow Yogurt processing of artificially contaminated milk resulted in a significant reduction (> 70%) of CPA concentration after the first day of storage.

Significant decomposition of CPA occured in acidic buffers. In basic environments it was less pronounced while a

Cyclopiazonic acid



Cyclopiazonic acid

neutral pH caused minor rates of decomposition.

Assessment of possible health effects is difficult at this stage because analytical methods for the detection of CPA in foods are still being developed.

D

Dairy products Dairy products may be contaminated by \rightarrow mycotoxins via two different routes. Indirect contamination occurs when contaminated feedstuffs are consumed by dairy \rightarrow cattle. The cause of the direct contamination is the colonization of dairy products by mycotoxigenic molds, which may result in the contamination of the substrate. With respect to indirect contamination \rightarrow aflatoxin M₁ the \rightarrow milk metabolite of \rightarrow aflatoxin B₁ is most important.

The insolubility of AFM₁ in the milk fat and absorption in the curd resulted in a specific pattern of distribution depending on the end-product, e.g. \rightarrow butter, \rightarrow cheese, \rightarrow cream or whey (\rightarrow whey powder). About 10% of the original AFM₁ concentration in the milk is found in cream the remaining in the skimmed milk. Approximately 10% of the AFM₁ in the cream goes into the butter while up to 90% is retained in the buttermilk $(\rightarrow \text{ milk-, butter})$. AFM₁ distribution in the single fractions is related to their content of non-fat milk solids, probably due to casein binding. The acidification during cottage cheese (\rightarrow cheese, cottage) production caused losses of AFM1 concentration in the range of 20%, 30% is accumulated in the curd, 50% in the whey (see Figure Losses of AFM₁ during processing of milk).

Although \rightarrow carry over of e.g. \rightarrow ochratoxin A, \rightarrow sterigmatocystin, \rightarrow deoxynivalenol, \rightarrow T-2 toxin and \rightarrow zearalenone in milk has been reported, the rate of transmission and/or toxicity of the metabolites is low. Therefore, these mycotoxins do not represent a reasonable cause of concern.

Direct mycotoxin contamination may be due to starter cultures (e.g. \rightarrow Penicillium roquefortii Thom and \rightarrow Penicillium camembertii Thom) during cheese fermentation or accidental growth of molds on dairy products. Direct aflatoxin contamination is unlikely because \rightarrow Aspergillus flavus Link and \rightarrow Aspergillus parasiticus Speare do not belong to the frequent colonizers of these substrates. Cheeses, with their lower aw-values promote fungal growth (and potential mycotoxin contamination), and therefore belong to the more susceptible dairy products. Dairy products may contain the following mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/22, conc.: 6.4 µg/kg, country: Germany

incidence: 2/23*, conc. range: 10-20 $\mu g / kg$, \emptyset conc.: 15 $\mu g / kg$, country: India, *indigenous

 \rightarrow cheese

Decontamination Decontamination should be cheap and simple, ideally using the existing technology. The procedure should be effective against a variety of \rightarrow mycotoxins and not lead to the toxic degradation metabolites. No reduction in the nutritional and palatable properties of \rightarrow grains or grain products should occur. Detoxification processes may be divided into three categories: physical, chemical, and biological. Physical methods include cleaning and washing, dehulling as well as \rightarrow milling. Their effectiveness greatly depends on the relative distribution of mycotoxins throughout the grains and the degree of contamination. Because of additional cost for drying, washing is only suitable as a cleaning step prior to wet milling. Separation of mycotoxin-contaminated grains due to differing physical properties is possible by fractionation (specific gravity table), density segregation (certain liquids) or fluorescence under ultraviolet light. The heat stability of most food-relevant mycotoxins reduces the

Decontamination

effectivity of cooking, baking, roasting and microwave heat.

Most of the chemicals used for decontamination have only a limited effect on the mycotoxins. Their effectivity is greatly influenced by the moisture content of the substrate and the processing temperature. Different chemicals like calcium hydroxide monomethylamine, hydrogen peroxide, sodium hypochlorite or sodium bisulfite have been used. Ascorbic acid essentially removed \rightarrow patulin from contaminated \rightarrow apple juice. For commercial decontamination of aflatoxin-containing cotton seeds, \rightarrow maize and peanut cakes / meal (\rightarrow peanuts) ammonia is used in the US, France, Nigeria, etc. At present, ammonia decontamination is the most effective and economically feasible method. Biological methods include e.g. the addition of mold inhibitors or potential mycotoxin-binding agents to the feed. In addition, various microorganisms have been tested for their detoxification potential. \rightarrow Flavobacterium aurantiacum essentially removed \rightarrow aflatoxin B₁ from different kinds of food while Saccharomyces *cerevisiae* detoxified \rightarrow patulin in \rightarrow apple juice during \rightarrow cider production.

Deer \rightarrow Roe deer

Deoxynivalenol (Syn.: DON, Rd-toxin, vomitoxin) belongs to the group of naturally-occurring \rightarrow trichothecenes (3α , 7α ,15-trihydroxy-12,13-epoxytrichothec-9-en-8-one) and is produced by different species of the genus \rightarrow Fusarium, with \rightarrow Fusarium graminearum Schwabe being the most important (see Figure Deoxynivalenol). The first isolation of Rd-toxin (previous name) was reported in 1972 for Japanese *Fusarium*-damaged \rightarrow barley, which showed a simultaneous contamination with \rightarrow nivalenol. Subse-

quent isolations from *F. graminearum*infected \rightarrow maize, which caused vomiting in swine in the United States, led to the trivial name vomitoxin (1973).

CHEMICAL DATA

Empirical formula: $C_{15}H_{20}O_6$, molecular weight: 296

FUNGAL SOURCES

Fusarium acuminatum (?), \rightarrow Fusarium culmorum (W. G. Smith) Sacc., \rightarrow Fusarium graminearum Schwabe, \rightarrow Fusarium nivale (Fr.) Ces., \rightarrow Fusarium sporotrichioides Sherb.

NATURAL OCCURRENCE

 \rightarrow baby cereals, \rightarrow baby food, \rightarrow barley, \rightarrow barley flour, \rightarrow barley grits, \rightarrow barley malt, \rightarrow beans, \rightarrow beer, \rightarrow beer, barley, \rightarrow beer, wheat, \rightarrow bran, \rightarrow bread, \rightarrow breakfast cereals, \rightarrow buns, \rightarrow cereal products, \rightarrow chapatti, \rightarrow chilli powder, \rightarrow cookies, \rightarrow coriander, \rightarrow crackers, \rightarrow croissant butter, \rightarrow croissant fat, \rightarrow figazzas, \rightarrow flour, \rightarrow foods, \rightarrow garlic, \rightarrow ginger, \rightarrow grains, \rightarrow job's-tears, \rightarrow libritos, maize, \rightarrow maize flour, \rightarrow maize grits, \rightarrow maize meal, \rightarrow maize, brewers, \rightarrow maize, brewers flaked, \rightarrow maize, brewers grits, \rightarrow maize, canned, \rightarrow maize, fiber cereal, \rightarrow maize, hominy, \rightarrow maize, infant cereal, \rightarrow maize, infant cream corn, \rightarrow maize, popped, \rightarrow maize, preharvest, \rightarrow maize, puffed, \rightarrow maize, quality-protein, \rightarrow maize, shelled, \rightarrow maize, sweet, \rightarrow masa, \rightarrow millet, \rightarrow millet meal, \rightarrow muesli ingredients, \rightarrow noodles, \rightarrow oats, \rightarrow pop corn, potatoes, \rightarrow rice, \rightarrow rye, \rightarrow rye bran, \rightarrow rye flour, \rightarrow snack food, \rightarrow sorghum, \rightarrow soybean, \rightarrow spaghetti, \rightarrow triticale, \rightarrow wheat, \rightarrow wheat grits, \rightarrow wheat products Cereals like wheat, barley and maize usually contain the highest DON-concentrations. DON is the most important mycotoxin in grains in several countries such as Austria, Canada, Italy, South Africa, Sweden, UK, USA. Because of its

stability, DON survives processing

 $(\rightarrow \text{ milling})$ resulting in the contamination of cereal products (e.g. corn steep liqour, corn starch). Fractions which are used as animal feed (e.g. gluten meal and wet fibre) may show high DON-levels. Simultaneous occurrence of DON, \rightarrow zearalenone and \rightarrow aflatoxin B₁ in scabby wheat is possible.

Rate of transmission (\rightarrow Carry over) into cow \rightarrow milk is extremely low (<4 µg/l). Because of rapid elimination low to medium DON-levels in the diet do not result in the accumulation of residues in swine. Transmission/residues of DON in(to) meat, milk or eggs is negligible.

Тохісіту

Acute toxicity is characterized by intestinal disorders and emesis, especially in swine. However, the presence of DON limits feed consumption at concentrations > 1 μ g/kg, so that acute toxicity seldom occurs. \rightarrow Poultry (egg quality, weight reduction) and \rightarrow cattle (reductions in feed intake, conception rate and milk production), possibly due to extensive degradation to secondary metabolites in the rumen, are more tolerant. Dermatological lesions, gastrointestinal disorders, hematological changes $(\rightarrow \text{hemorrhage}) \text{ and } \rightarrow \text{immunosuppres-}$ sive, \rightarrow teratogenic as well as nephrotoxic (?) effects in animals have also been reported.

Humans seem to be quite sensitive to DON.

 LD_{50} (po): 46 mg/kg bw mice A potently-synergistic toxic effect to laboratory animals occurred in combination with culmorin, dihydroxycalonectrin and sambucinol as metabolites of *F. graminearum*.

The co-contamination of grains with other mycotoxins may cause unanticipated interactions to the detriment of animals and humans. It seems possible that the carcinogenicity of AFB_1 is enhanced by the immunosuppressive acting \rightarrow trichothecenes.

DETECTION

ELISA, GC, HPLC, MS, TLC

Possible Mycotoxicosis Outbreaks of acute gastrointestinal illness in humans (China, India).

FURTHER COMMENTS

DON is probably the most common mycotoxin contaminating food and feed. DON is a contaminant virtually wherever cereals are grown. Co-occurrence with \rightarrow zearalenone is common in grain worldwide. The contamination of cereals with DON may be eliminated by plant breeding. DON is often co-occurring with \rightarrow nivalenol, \rightarrow diacetoxyscirpenol, and \rightarrow T-2 toxin.

Distribution in grains: DON is primarily located in the grain at the sites of fungal growth. Only little translocation occurred to other sites in the kernel. Low levels of fungal and mycotoxin contamination (50-1000 µg/kg) typically result in DONaccumulation near the exterior surface of the kernel. Here, most of the fungal mycelium is to be expected. The \rightarrow flour of such wheat will contain relatively low mycotoxin levels with respect to the whole kernel. Higher concentrations (>4000 µg DON / kg) may cause a more even distribution throughout the kernel due to a deeper penetration of the fungus. Mycotoxin levels of flours prepared from highly contaminated grains are comparable to those in the \rightarrow bran and other outer portions of the kernels. However, in some cases this pattern of distribution is not related to high DON-concentrations in individual kernels. It seems possible that a correlation exists between the distribution of DON and the degree of fungal (Fusarium) contamination of the kernels.

Stability: Processing (e.g. cleaning, milling, \rightarrow baking) of contaminated \rightarrow cereals usually does not result in significant

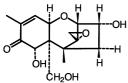
losses of DON in the finished product. During milling of wheat, DON was detected throughout all of the milling fractions: bran shorts, reduction flour, break flour (in decreasing order). Several studies confirmed an accumulation of DON in the bran fraction whereas the lowest concentrations were found in the flour (reduction \approx 50%).

About 50% of DON survived the baking process. An even dramatic increase in DON-concentration (180%) has been observed during doughnut preparation. This might be due to enzymatic conversions of DON-precursors already present in the used soft wheat.

DON is the mycotoxin which best survives the brewing process. An increase in amount during mashing may occur.

Deoxynivalenol monoacetate → 3-acetyldeoxynivalenol

Deoxynivalenol toxicosis Between July through September 1987, human food poisonings occurred in the Kashmir Valley in northwestern India. Approximately 50,000 people were affected by this noncommunicable disease irrespective of age or sex. The consumption of \rightarrow bread made from certain consignments of \rightarrow wheat led to different symptoms like abdominal pain, a feeling of fullness in the abdomen, throat irritation, diarrhoea, emesis, blood in the stool and allergic reactions 15 min to 1 h after ingestion. Unseasonal rains during the harvest season caused a considerable mold contamination of the wheat. Local millers bought





the moldy wheat for a much lower price and mixed it with good wheat (50: 50). The corresponding flour was sold to local bakers, who in turn sold it to consumers as flour or bread. The consistency of bread made from the moldy wheat resembled "chewing gum". Mycological examination of the grains and the flours (24 brands) always revealed a \rightarrow Fusarium contamination. \rightarrow Aspergillus spp. and \rightarrow Penicillium sp. occurred to a minor extent. In addition, several mycotoxins could be isolated: \rightarrow deoxynivalenol (conc. 346-8380 µg/kg, 11 samples), \rightarrow nivalenol (conc. 30-100 μ g / kg, 2 sa), acetyldeoxynivalenol (conc. 600-2400 µg/kg 4 sa), \rightarrow T-2 toxin (conc. 550-4000 μ g/kg, 5 sa). While identification of different \rightarrow trichothecenes failed, the detection of pesticide residues, \rightarrow aflatoxins and \rightarrow ergot alkaloids was negative.

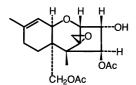
Diacetoxyscirpenol (Syn.: anguidine, DAS) belongs to the group of naturally occurring \rightarrow trichothecenes (3 α -hydroxy-4,15-diacetoxy-12,13-epoxytrichothec-9ene), which is produced by different \rightarrow Fusarium species, with *Fusarium sporotrichioides* Sherb. being the most important (see Figure Diacetoxyscirpenol). The first isolation was reported for \rightarrow Fusarium equiseti (Corda) Sacc. sensu Gordon in 1961. Structure elucidation followed in 1965/1966.

Chemical Data

Empirical formula: $C_{19}H_{26}O_7$, molecular weight: 366

FUNGAL SOURCES

Fusarium acuminatum, F. avenaceum (?), Fusarium equiseti (Corda) Sacc. sensu Gordon, \rightarrow Fusarium graminearum Schwabe, \rightarrow Fusarium moniliforme Sheldon, \rightarrow Fusarium oxysporum Schlecht. emend. Snyd. & Hansen, \rightarrow Fusarium poae (Peck) Wollenw., \rightarrow Fusarium sambucinum



Diacetoxyscirpenol

Fuckel (good producer), *F. semitectum*, \rightarrow Fusarium sporotrichioides Sherb.

NATURAL OCCURRENCE

→ barley, → beans, → beer, → chilli powder, → curry, → maize, → oats, → wheat

Тохісіту

cancerogenic, dermatoxic, hemorrhagic (\rightarrow hemorrhage) (enteritis), phytotoxic LD₅₀ (po): 7.3 mg/kg bw rats (21-dayold)

DETECTION GC, MS, spectroscopy, TLC

Possible Mycotoxicosis Besides T-2 toxin DAS should also be involved in \rightarrow alimentary toxic aleukia.

FURTHER COMMENTS

DAS often occurs naturally together with \rightarrow deoxynivalenol.

The rapid and extensive metabolization of DAS in pigs has been reported. Although accumulation of this mycotoxin is not be expected in naturally exposed animals the toxicity and tissue distribution of unknown metabolites needs further clarification.

Dihydroalterperylenol (Syn.: altertoxin I, \rightarrow altertoxin I-III)

DON → Deoxynivalenol

Dothideales → Ascomycota

Duck may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 33/41*, conc. range: 0.203-2.484 μg / kg, Ø conc.: 0.84 μg / kg, country: Czechoslovakia, *wild duck, liver incidence: 31/41*, conc. range: 0.3-3.605 μ g/kg, Ø conc.: 0.594 μ g/kg, country: Czechoslovakia, *wild duck, kidney → ochratoxin A incidence: 11/19, conc. range: $\leq 0.09 \ \mu g/$ kg, \emptyset conc.: 0.02 µg/kg, country: Denmark incidence: 4/7*, conc. range: $\leq 0.16 \ \mu g$ / kg, \emptyset conc.: 0.06 µg/kg, country: Denmark, *liver \rightarrow meat

Durum wheat \rightarrow wheat

Dyspnea shortness of breath, difficult or labored breathing

Е

Edema is characterized by the accumulation of an exessive amount of tissue fluid in intercellular spaces.

Egg products may contain the following \rightarrow mycotoxins:

aflatoxin (no specification) (\rightarrow aflatoxins) incidence: 1/112, conc.: 0.06 µg/kg, country: USA

ELEM \rightarrow Equine leukoencephalomalacia, \rightarrow fumonisins

Emericella \rightarrow Trichocomaceae; anamorph \rightarrow Aspergillus

Emu aran is a Nigerian indigenous beverage (palm juice) made from the sap of *Raphia vinifera* and *R. raphia*. Emu aran may contain the following \rightarrow mycotoxins: aflatoxin B (\rightarrow aflatoxins) incidence: 2/2, conc. range: 83-86 µg/kg, Ø conc.: 84.5 µg/kg, country: Nigeria

Encephalopathy and fatty degeneration of the viscera $(Syn: \rightarrow Reye's syndrome)$

Enchilada → Tortilla

Endemic Balkan nephropathy \rightarrow Balkan Endemic Nephropathy

Endemic familiar arthritis of malnad This non-congenital disease is characterized by abnormal bone growth and occurred in the Malnad district in southern India from 1965-1975. In this area heavy rainfalls are common.

Members of 140 families belonging to the most impoverished castes were affected. Their diet mainly comprised \rightarrow rice and various fauna like \rightarrow fish and crabs. The victims were of both sexes and all ages although children younger than five years

old did not show any symptoms. The bilateral, symmetrical lesions (osteoarthritis) primarily occur in the hip joint, pelvis, as well as vertebrae and may progress up to the knees. Other joints are rarely affected. Severe impairment of the patients mobility may result from the disease.

Although the consumed foodstuffs have not been investigated for mold and mycotoxin contamination, similarities (epidemiological, pathological) with other bone growth disorder diseases (\rightarrow Kashin-Beck disease = osteoarthritis, \rightarrow Mseleni joint disease = lesions of the hip joint) in which \rightarrow mycotoxins have been investigated as possible etiological agents are obvious. In addition, all three diseases occur in geographically isolated areas.

Endemic panmyelotoxicosis \rightarrow Alimentary toxic aleukie

Enteritis is characterized by an inflammation of the intestines.

Epicladosporic acid → Cladosporium

Equine leukoencephalomalacia (Syn.: blindstaggers, foraging disease, corn stalk disease, leucoencephalitis, \rightarrow moldy corn poisoning) (Abbr.: ELEM) is a fatal, disease which affects the co-ordination of horses and was first described in the late 1800s. This disease was associated with \rightarrow Fusarium as early as 1904. Sporadic, seasonal, epidemic-like outbreaks have been reported in e.g. Argentina, Brazil, China, Egypt, South Africa and the United States. At present, two different forms have been reported: hepatotoxic and neurotoxic ELEM. The latter is more common.

The feeding of corn and other feeds highly infected with \rightarrow Fusarium moniliforme Sheldon and contaminated with

 \rightarrow fumonisins results in extensive damage to brain tissue. Lesions occur in form of none or more focal areas of liquefactive necrosis (= encephalomalacia) in the white matter (= leukoencephalomalacia) of the brain. One or both hemispheres may be affected. The encephalomalacic areas consist of large, irregular empty spaces. Random liquefactive (or malacic) lesions are characteristic for the subcortical white matter of the brain and the blood vessels show perivascular hemorrhages (\rightarrow hemorrhage) and \rightarrow edema or a cuffing by infiltrating leukocytes. \rightarrow esophageal cancer, \rightarrow porcine pulmonary edema

Equsi meal is a Nigerian type of foodstuff from the plant *Cocumeropsis edulis*. Equsi meal may contain the following \rightarrow mycotoxins aflatoxin B (\rightarrow aflatoxins) incidence: 1/1, conc.: 186 µg/kg, country: Nigeria

Ergot alkaloids In the view of toxicology and medicine, the alkaloids are the most important substances isolated from ergot. They particularly act on the nervous system. Based on their chemical structure the alkaloids are divided into lysergic acid, isolysergic acid and the clavine alkaloids (see Figure Ergot alkaloids). Lysergic acid derivatives are of the acid amide type and subdivided into the simple amides (e.g. ergometrine and ergine) and the peptide type comprising the ergotamine (e.g. ergotamine, ergosine), the ergotoxine (e.g. ergocristine, ergocornine, α -ergocryptine) and the ergoxine group. In the case of the clavine alkaloids, the carboxyl group, which is characteristic for the lysergic acid derivatives is reduced to a hydroxymethyl or a methyl group.

Ergot alkaloids are found in the sclerotia $(\rightarrow \text{ ergots})$ of $\rightarrow \text{Claviceps purpurea.}$

Each sclerotium contains a total of over 100 compounds; ergocristine and ergotamine (lysergic acid dervatives) are generally the major components but alkaloid variation in individual sclerotia and throughout a contaminated field is high. The concentration and composition of alkaloids in ergot is influenced by different factors like strain and stage of maturity of the fungus, type of the host plant, climatic and geographic conditions. Ergots of pearl \rightarrow millet mainly contain alkaloids of the clavine type (Claviceps fusiformis), whereas ergot alkaloids of \rightarrow rye and \rightarrow wheat belong mainly to the ergotamine group (C. purpurea).

FUNGAL SOURCES

Claviceps spp., → Aspergillus spp. (e.g. → Aspergillus clavatus Desm., → Aspergillus fumigatus Fres.), → Emericella spp., → Penicillium spp. (e.g. *P. chermesinum*, *P. concavo-rugulosum*), → Rhizopus *nigricans*, and higher plants, e.g. *Ipomoea* spp.

NATURAL OCCURRENCE

→ baby cereals, → cereals, → rye flour, → triticale flour, → wheat Hostplants like wheat, → barley, rye, → oats, → millet and Indian corn are found in the family of Graminae comprising the most important plants for human nutrition.

Wheat and rye flours usually contain only low alkaloid levels (< $100 \mu g/kg$). Because of this situation there is almost no reason for concern.

No ergot alkaloids could be detected in \rightarrow meat and \rightarrow milk of livestock and \rightarrow poultry after ingestion of contaminated feed which caused typical ergotism. Transmission of ergotism to breast-fed infants is not possible.

TOXICITY

Some ergot alkaloids are destroyed by ultraviolet light and there is much evidence to show that ergot sclerotia were more toxic when fresh than after storage. Ingestion of higher alkaloid levels will result in neurological and / or gangrenous disorders. The nervous disorders include \rightarrow ataxia, tremors, staggers, and \rightarrow convulsions. The gangrenous form is characterized by vascoconstrictant effects (necrosis, sloughing of the extremities). Lower chronic levels are responsible for cardiac disorders.

Acute poisoning with gangrene occurred after the ingestion of between 5 and 72 mg ergotamine and 9 mg ergometrine. However, it was estimated that humans tolerate ca. $26 \ \mu g$ clavine alkaloids / kg bw without any toxic effects.

DETECTION

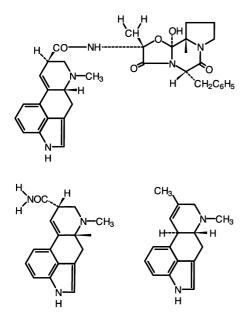
ELISA, densiometry, LC, spectrophotofluorometry, TLC

Mycotoxicosis

 \rightarrow ergotism

FURTHER COMMENTS

Stability / Reduction: The lysergic acid derivatives are unstable to heat so significant losses occur during \rightarrow bread proces-



Ergot alkaloids. Ergotamine (lyserg acid), erginine (isolysergic acid), agroclavine (clavine alkaloid)

sing. Baking caused a reduction in alkaloid concentration of up to 100% in whole wheat bread and up to 85% rye bread. During the making of triticale pancakes the losses amounted to 74%. A reduction of ca. 90% in total alkaloid content was observed after treatment of wheat ergot sclerotia with chlorine. During the normal cleaning and \rightarrow milling process for grains, ergots are largely removed with the dockage. An accumulation of 70-80% of the ergot in the bran or shorts fractions was observed during milling. Therefore, these processing steps will usually result in a low alkaloid concentration in flour.

Ergotism Ergotism ("holy fire"), caused by \rightarrow ergot alkaloids as derivatives of lysergic acid found in the sclerotia of \rightarrow Claviceps, is probably the first recognized and best known \rightarrow mycotoxicosis with respect to recorded effects on man. It is evident from history that ergotism has plagued humans and animals for centuries. In 430 B.C. an epidemic occurred among the Spartans that may have been due to ergot. In western and central Europe the use of contaminated \rightarrow rye for \rightarrow bread making led to large-scale epidemics in the Middle Ages. The first clear report of ergotism dates from 1582 but since 857 outbreaks of a disease resembling ergotism have been known in Central Europe. During the Middle Ages the disease was also called Saint Anthony's fire because pilgrims suffering from it reported how they had been miraculously cured after paying homage at St. Anthony's shrine in Dauphiné (France). The recovery of the patients was probably due to a change in diet made at the shrine.

Compared to the Middle Ages, human ergotism is now extremely rare, which is due to the change from rye to \rightarrow wheat consumption and improvements in pre-

Ergotism

venting contaminated grain products from entering the food chain. Serious outbreaks sporadically occur in countries like India in 1975 (red millet / *Claviceps fusiformis*). In one outbreak in Ethiopia (1978), 93 people were gangrenous and 47 died after the consumption of wild \rightarrow oats weeds contaminated with sclerotia of *C. purpurea*.

There are two types of ergotism, convulsive (neurological) and gangrenous (necrotic) ergotism. The latter form is due to the ingestion of sclerotia of C. purpurea and began with lassitude, sometimes accompanied by a prickling or an icy cold sensation in the limbs. Severe muscular pains, especially in the calf, followed. Although appetite and pulse remained constant at the beginning of the disease, the intellect was dulled. Swelling and inflammation of the limbs ensued. Similarly intensive burning pains with sensations of intense heat alternated with those of icy coldness. The pains sometimes ceased suddenly, leaving numbness. The skin was coverd with red to violet vesicles while the unaffected parts (face, white of the eyes) turned yellow indicating \rightarrow jaundice. As gangrene set in, the toes and fingers became necrotic (black). In servere cases, the loss of fingers or toes, or even of all four limbs, occurred.

The impairment of the nervous system is characteristic for convulsive ergotism (\rightarrow convulsions) which is caused by *C. paspali.* The following symptoms are typical: sustained spasms, muscle cramps and twitching, numbness of the hands and feet, a tingling sensation under the skin, constriction of the blood vessels, followed by mortification of the limbs. Hallucination also occurs. Even in nonfatal cases full mental recovery was seldom. The mortality rate of ergotism ranges between 11 and 60%. Death may occur within several hours after ingestion of ergots but recovery is possible although not always completely. A higher susceptibility of previous victims of ergotism to recurrences has been reported. Ergotism occurred in Europe (particularly France and Germany), USA, Ethiopia as well as India and besides humans, \rightarrow cattle, pigs, horses, sheep and \rightarrow poultry are affected.

Ergots Ergot bodies are the \rightarrow sclerotia of \rightarrow Claviceps spp. which contain many toxic \rightarrow ergot alkaloids (see Figure Ergots). About 50 species are known to infect many different grasses. The most widespread and common species is C. purpurea responsible for many cases of \rightarrow ergotism in humans and animals. The tightly-packed masses of fungal mycelium develop instead of kernels in grasses and \rightarrow cereals (mainly \rightarrow rye but also e.g. \rightarrow wheat). The size and shape of the ergots may be roughly that of the kernels of the host plant but larger forms do exist. In general, not more than seven to eight ergots are found on a single spike of rye. Purple-black in colour they contain various pharmacologically active compounds, especially the \rightarrow ergot alkaloids (conc. 0.1-0.8%). Low winter soil temperatures and wet springs stimulate the germination of the sclerotia. Infections of the host plants are enhanced by warm summers preceded by cold wet springs.



Ergots. Ergots in rye

Although large sclerotia are easily removed during grain cleaning, small and broken ones may pass through this processing step.

A maximum level of 0.05% and 0.3% ergot by weight has been suggested as an acceptable level for use in the production of \rightarrow flour in Canada and other countries.

Erythema is characterized by redness of the skin due to congestion of the capillaries.

Esophageal cancer (Abbr.: EC) In certain parts of southern Africa, China, and northern Italy, the incidences of EC are extremely high with substantial varations in EC rates separated by only short geographical distances. In the high incidence areas very high fumonisin concentrations (FB₁, FB₂) have been detected in \rightarrow maize and maize products intended for human consumption. In addition, \rightarrow Fusarium moniliforme Sheldon strains isolated from Chinese maize (Linxian County) produced nitrosamines including N-methylbenzylnitrosamine, one of the most potent nitrosamines inducing esophageal cancer in experimental animals.

It has been concluded that the etiology of human esophageal cancer probably involves not one but several factors (e.g. vitamin and trace elements deficiencies in high risk populations in the Transkei). Although the experimental proof of a causative relationship between fumonisin contamination of corn-based staple diet and EC is still lacking, it is obvious that exposure to \rightarrow fumonisins due to the ingestion of maize and maize products in the high EC areas of Transkei / South Africa, Linxian and Cixian Counties/ northern China, northern Italy and southeastern United States is one etiological factor (of several) for human esophageal cancer.

Eumycota Kingdom of Eukaryota, the true \rightarrow fungi

Eupenicillium \rightarrow Trichocomaceae

Eurotiaceae (Syn.: \rightarrow Trichocomaceae)

Eurotiales → Ascomycota

Eurotium \rightarrow Trichocomaceae, anamorph: \rightarrow Aspergillus

In marginally dried grain ($\rightarrow a_w 0.65$ 0.70) Eurotium spp. besides \rightarrow Aspergillus restrictus G. Sm. and Eurotium halophilicum belong to the earliest developing and most commonly encountered \rightarrow storage fungi. However, in some case Eurotium spp. also occurs on \rightarrow grains pre-harvest. They are a characteristically xerophilic group of fungi showing maximum growth rates at a_w < 1.0. Moisture contents in the range of 14.5-15% (\rightarrow cereals) enable their growth. Their metabolic water increases the a_w of the substrate contributing to the growth of mycotoxin producing fungi like Aspergillus spp. and \rightarrow Penicillium spp. Important species are E. amstelodami, E. chevalieri, E. herbariorum, E. rubrum (see Figure Eurotium).

They are able to synthesize different mycotoxins like \rightarrow ochratoxin A and \rightarrow sterigmatocystin. However, accumulation probably does not reach dangerous concentrations. Some still unknown car-



Eurotium. Eurotium herbariorum

Eurotium

cinogenic compounds should also be produced.

Expansin (Syn.: \rightarrow patulin)

Extracellular mycotoxins like \rightarrow aflatoxins, \rightarrow citrinin, \rightarrow kojic acid, \rightarrow mycophenolic

acid, $\rightarrow \beta$ -nitropropionic acid, \rightarrow ochratoxins, \rightarrow patulin, \rightarrow penicillic acid, \rightarrow PRtoxin, \rightarrow rubratoxins, \rightarrow T-2 toxin, and

 \rightarrow zearal enone diffuse into the substrate.

 \rightarrow Intracelullar mycotoxins, \rightarrow mycotoxins

F-2 toxicosis (Syn.: estrogenic syndrome, hyperestrogenism, vulvo-vaginitis) \rightarrow Zearalenone, mainly produced by \rightarrow Fusarium graminearum Schwabe, and related metabolites (e.g. zearalenol) possess estrogenic activity. They may cause severe reproductive and infertility problems in domestic animals. Pigs are very susceptible, \rightarrow cattle seem less susceptible, and chickens are apparently not affected. The effect of long term exposure of humans to low zearalenone levels in the diet is still unknown but this estrogene may cause hormone-dependent tumors in women.

Zearalenone production by *F. graminearum* is favored by both high moisture content and alternating moderate and low temperatures during \rightarrow maize storage. In consequence, adequate drying of maize and storage at low moisture levels will reduce zearalenone contamination. The use of resistant varieties, as well as dilution of contaminated \rightarrow cereals with sound cereals contribute to avoiding F-2 toxicosis. Clinical reports of hyperestrogenism in swine date as far back as the 1920s.

F-2 toxin \rightarrow zearalenone

Fagicladosporic acid → Cladosporium

Fennel may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 2/10, conc. range: 11 µg/kg, country: India incidence: 6/9, conc. range: 30-275 µg/kg, country: India \rightarrow aflatoxin B₂ incidence: 1/10, conc.: 8 µg/kg, country: India incidence: 6/9, conc. range: 28-173 µg/kg, country: India

 \rightarrow aflatoxin G₁ incidence: 6/9, conc. range: 15-76 µg/kg, country: India \rightarrow aflatoxin G₂ incidence: 6/9, conc. range: 9-69 µg/kg, country: India \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 1/3, conc.: 1.2 µg/kg, country: UK \rightarrow citrinin incidence: 2/9, conc. range: 28-59 µg/kg, Ø conc.: 43.5 µg/kg, country: India \rightarrow ochratoxin A incidence: 1/9, conc.: nc, country: India incidence: 3/3, conc. range: $\leq 0.2 \ \mu g / kg$, country: UK → sterigmatocystin incidence: 1/9, conc.: 142 µg/kg, country: India \rightarrow zearalenone incidence: 1/3, conc.: 7 μ g/kg, country: UK \rightarrow spices

Fenugreek may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 3/6, conc. range: 2-4.3 µg/kg, country: Egypt \rightarrow aflatoxin B₂ incidence: 2/6, conc. range: 2.5-3 µg/kg, country: Egypt \rightarrow aflatoxin G₁ incidence: 1/6, conc.: 1.8 µg/kg, country: Egypt \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 1/41*, conc.: 2.5 µg/kg, country: UK, *miscellaneous \rightarrow spices, imported

Fermented products may contain the following \rightarrow mycotoxins: \rightarrow zearalenone incidence: 6/55, conc. range: 8-53 µg/kg, country: Swaziland \rightarrow miso, \rightarrow oriental fermentations Fibrosis

Fibrosis fibrous tissue formation

Field fungi The original source of these fungi is the field. They infect the developing and mature \rightarrow grains while the plants are still growing in the field, or after the seeds are cut and swathed but before they are threshed. The most common field fungi which are present at the onset of storage like \rightarrow Alternaria spp., \rightarrow Cladosporiumspp., *Epicoccum* spp., \rightarrow Fusarium spp. and *Drechslera* spp. are succeeded by \rightarrow storage fungi with increasing storage time. Field fungi have high water requirements (90-100% relative humidities) which in \rightarrow cereals amount to a moisture content of $\approx 20\%$. At lower moisture levels they do not compete well with the storage fungi and most of them die rapidly. Some of the field fungi produce and cumulate \rightarrow mycotoxins in kernels and chaff, e.g. \rightarrow Alternaria mycotoxins and \rightarrow Fusarium mycotoxins.

Figazzas may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 8/8, conc. range: 212-2800 μ g / kg, Ø conc.: 851 μ g / kg, country: Argentina

 \rightarrow wheat products

Fig paste may contain the following \rightarrow mycotoxins:

→ aflatoxins (no specification) incidence: 105/132, conc. range: 1-10 μ g / kg (86 samples), 11-165 μ g / kg (19 sa), country: Turkey

Figs Although \rightarrow ochratoxin A and \rightarrow kojc acid have been isolated from figs the \rightarrow aflatoxins represent the main important \rightarrow mycotoxins contaminating fig fruits.

During the ripening stage the \rightarrow fruits become susceptible to aflatoxin contami-

nation by \rightarrow Aspergillus flavus Link while immature fig fruits do not support development and aflatoxin formation by A. flavus. Under experimental conditions those fruits taking longest to ripen contained up to 72,000 µg aflatoxin/kg when inoculated in the green stage. Maximum aflatoxin formation occurred on the fourth and sixth day and then decreased gradually. Only very little aflatoxin could be detected in fruits inoculated in the firmripe state if dried immediately. Aflatoxin amount increases with the extension of the drying time of the figs. During sundrying on the tree, figs are very susceptible to fungal infection and development leading to fairly high levels of aflatoxins. Under natural conditions Turkish figs remain on the trees until they are shrivelled ripe. After falling to the ground (occasionally covered with cloths), they are dried in sunlight before the fruits are collected. These conditions seem to promote aflatoxin contamination of the fruits. Infection of the figs with A. flavus and \rightarrow Aspergillus parasiticus Speare, due to gall wasps, during pollination will lead to potential aflatoxin contamination in a rather late stage during ripening. However, it is still being debated, whether aflatoxin contamination occurred only after the fruits had fallen from the trees and were lying on the ground for drying. The pattern of contamination in figs closely resembles that described for \rightarrow peanuts: only some figs show an aflatoxin contamination but these individual fruits usually contain very high concentrations (5000 μ g \rightarrow aflatoxin B₁/kg). It was estimated that the degree of contamination is in the range of ca. 1 in 100. The aflatoxin contamination is restricted to a great extent to that part of the fig showing surface fluorescence.

Although figs intended for retail sale are packed in very close contact with each other in small boxes, only slight cross-

contamination or none at all could be observed among packed fruits. Figs may contain the following \rightarrow mycotoxins: aflatoxin B₁ incidence: 6/25*, conc. range: 0.1-3 µg/kg, country: Switzerland, *dried incidence: 2/4*, conc. range: 2.5-11.8 μg/kg, country: Syria, *dried incidence: 8/206* **, conc. range: 3.6-320 μ g/kg, Ø conc.: 112 μ g/kg, country: Turkey, *dried, **lower grade figs incidence: 94/386*, conc. range: 0.2-5 μ g / kg (85 samples), 5-10 μ g / kg (7 sa), $10-20 \ \mu g/kg \ (1 \ sa), \ 20-30 \ \mu g/kg \ (1 \ sa),$ country: Turkey, *dried, randomly selected incidence: 37/52*, conc. range: 5-76,000 μg / kg, country: Turkey, *dried, selected, fluorescent incidence: 8/16*, conc. range: 0.2-5 μ g / kg (6 samples), 5-10 μ g / kg (1 sa), 10-20 μ g/kg (1 sa), country: Turkey, *dried, discolored incidence: 52/62*, conc. range: 0.2-10 μ g / kg (18 samples), 10-100 μ g / kg (8 sa), 100-1.000 μg/kg (14 sa), 1000-10,000 $\mu g / kg (11 sa), > 10,000 \mu g / kg (1 sa),$ country: Turkey, *fluorescent (BGY) \rightarrow aflatoxin B₂ incidence: 4/206* **, conc. range: 23.5-71.8 μg / kg, Ø conc.: 50.6 μg / kg, country: Turkey, *dried, **lower grade figs \rightarrow aflatoxin G₁ incidence: 3/206* **, conc. range: 12.4-97.5 μg/kg, Ø conc.: 61.4 μg/kg, country: Turkey, *dried, ** lower grade figs incidence: 49/386*, conc. range: 0.2-5 $\mu g\,/\,kg$ (45 samples), 5-10 $\mu g\,/\,kg$ (2 sa), 20-30 μ g/kg (2 sa), country: Turkey, *dried, randomly selected incidence: 15/52*, conc. range: 5-180,000 µg/kg, country: Turkey, *dried, selected, fluorescent incidence: 3/16*, 0.2-5 μg/kg (1 sample), 5-10 μ g / kg (2 sa), country: Turkey, *dried, discolored

incidence: 21/62*, conc. range: 0.2-10 μ g / kg (7 samples), 10-100 μ g / kg (4 sa), 100-1000 µg/kg (4 sa), 1000-10,000 $\mu g / kg (4 sa), > 10,000 \mu g / kg (2 sa),$ country: Turkey, fluorescent (BGY) incidence: 1/106*, conc.: 10 µg/kg, country: USA, *dried, selected, fluorescent \rightarrow aflatoxins (no specification) incidence: 53/103, conc. range: 5-203 μg/kg, country: Sweden incidence: 56/122*, conc. range: 1-10 μg/kg (43 samples), 12-96 μg/kg (13 sa), country: Turkey, *dried incidence: 6/165, conc. range: 2-29 μ g/kg, Ø conc.: 13 μ g/kg, country: USA \rightarrow kojic acid incidence: 52/52*, conc. range: 8-6,900,000 μg/kg, country: Turkey, *dried, selected, fluorescent → ochratoxin A incidence: 1/39*, conc.: \leq 0.6 µg/kg, country: Germany incidence: $3/30^*$, conc.: $\leq 3.3 \ \mu g / kg$, country: Germany incidence: 1/9*, conc.: 160 µg/kg, country: Switzerland, *dried incidence: 12/52*, conc. range: 5-12,000 μg/kg, country: Turkey, *dried, selected, fluorescent \rightarrow fruits

Filberts → hazelnuts

Fish may contain the following \rightarrow mycotoxins:

→ aflatoxin B₁ incidence: 1/1* **, conc.: 679 µg/kg, country: Thailand, *total: 795 µg AFB₁, AFB₂, AFG₁, AFG₂,/kg prepared food, **plaa tuu = Mackerel like, sun dried → aflatoxins incidence: 7*/139**, Ø conc.: 166 µg/kg, country: Thailand, **dried, and shrimps *total: Ø conc.: 722 µg/kg AFB₁, AFB₂, AFG₁, AFG₂

\rightarrow ochratoxin A

incidence: 14/20, conc. range: 1000-2000 μ g / kg (sqd), country: Sierra Leone According to Frisvad (1988) potential mycotoxins in dried fish may be \rightarrow ochratoxin A and \rightarrow citreoviridin.

Flavobacterium aurantiacum removes \rightarrow aflatoxins from fluid and solid foods such as cow and peanut milk, vegetable oil, \rightarrow peanuts and \rightarrow peanut butter as well as \rightarrow maize. Rapid conversion of AFB₁ led to water-soluble degradation products. Release of CO₂ by the living cells of the bacterium contributes to the assumption that AFB₁ is at least in part metabolized.

Flavomycelin → Luteoskyrin

Flour (cereals, no specification) Cereal flours mainly show contamination with species of the genera \rightarrow Aspergillus and \rightarrow Penicillium. The degree of contamination varies from sample to sample and probably reflects different sanitation standards in the mills.

Although mycotoxin-producers may contribute to the mold flora of \rightarrow flour to a small extent, their detection is important because transmission into food products of which flour is an ingredient is possible. Improper processing of these food products may result in growth of the fungi and subsequent mycotoxin formation.

Flour may contain the following \rightarrow mycotoxins:

 \rightarrow citrinin incidence: 11/21, conc. range: 0.2-1.0 ug/kg. Ø conc : 0.55 ug/kg. country

 μ g / kg, Ø conc.: 0.55 μ g / kg, country: Switzerland \rightarrow deoxynivalenol

incidence: 13/56, conc. range: 350-8380 μg/kg, country: India

incidence: 36/36, conc. range: 2-240

μg/kg, country: Japan

incidence: 2/2*, conc. range: 23-720 μ g/kg, Ø conc.: 372 μ g/kg, country: Papua New Guinea, *imported, wholemeal self-raising flour incidence: 2/2*, conc. range: 91-1460 μ g / kg, Ø conc.: 776 μ g / kg, country: Papua New Guinea, *imported, wholemeal plain flour → nivalenol incidence: 6 products analysed, conc. range: 37-190 µg/kg, country: Japan incidence: 1/2*, conc.: 13 µg/kg, country: Papua New Guinea, *imported, wholemeal self-raising flour incidence: 1/2*, conc.: 1375 µg/kg, country: Papua New Guinea, *imported, wholemeal plain flour \rightarrow ochratoxin A incidence: 3/80, conc. range: 0.4 µg/kg, country: Germany incidence: 17/93, Ø conc.: 2.2 μg/kg, country: Germany incidence: 26/52, conc. range: 0.1-0.49 μ g / kg (11 samples), 0.5-1.49 μ g / kg (13 sa), 1.5-9.99 µg/kg (2 sa), country: Germany incidence: 11/11, conc. range: < 2.5-20 μg/kg, country: Japan incidence: 48/215, Ø conc.: 4370 µg/kg, country: Poland incidence: 2/7*, conc. range: 490-2900 μg/kg, country: UK, *moldy incidence: 28/57, conc. range: ≤ 2.0 μ g / kg, country: UK incidence: 49/57, conc. range: ≤ 1.6 μ g / kg, country: UK incidence: 48/61, conc. range: \leq 3.2 μg/kg, country: UK incidence: 21/31, conc. range: ≤ 1.0 μ g / kg, country: UK \rightarrow zearalenone incidence: 2/2*, conc. range: 1450-2150 μ g / kg, Ø conc.: 1800 μ g / kg, country: Papua New Guinea, *imported, wholemeal self-raising flour incidence: 2/2*, conc. range: 1400-2570 μ g/kg, Ø conc.: 1985 μ g/kg, country:

Papua New Guinea, *imported, wholemeal plain flour \rightarrow cereals, \rightarrow barley flour, \rightarrow buckwheat flour, \rightarrow maize flour, \rightarrow rye flour, \rightarrow soybean flour, \rightarrow wheat flour

Food Estimations of the FAO stated that ca. 25% of the worldwide produced foodstuff contains mycotoxin(s) at detectable levels. The contamination of food (and feeds) mainly depends on the prevailing environmental conditions that favor mold growth and subsequent mytotoxin formation. As a consequence of the import/ export of food (and feeds) the problem of \rightarrow mycotoxicosis is not limited to any one geographical area but represents a real or potential problem in all areas of the world where food (and feeds) are consumed. It is evident that nearly all staple food products consumed anywhere in the world are prone to mycotoxin (\rightarrow mycotoxins) contamination.

Foods (canned, no specification) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: nc/4, conc. range: 0.2-1.4 μ g / kg, country: UK \rightarrow deoxynivalenol incidence: nc/4, conc. range: 4-9 µg/kg, country: UK \rightarrow fusarenon X incidence: 1/4, conc.: 15 µg/kg, country: UK → nivalenol incidence: 1/4, conc.: 18 µg/kg, country: UK \rightarrow ochratoxin A incidence: nc/4, conc. range: 0.1-0.3 μg/kg, country: UK \rightarrow zearalenone incidence: 1/4, conc.: 6.1 µg/kg, country: UK

Fresh cheese \rightarrow cheese, fresh

Frontoethmoidal encephalomenigocele

(Abbr.: FEEM) Teratogens (\rightarrow teratogenic), produced by fungi in grain staples, may be the cause of FEEM in Myanmar (formerly Burma). The disease occurs in countries like Australia, England, Germany, India, South Africa and United States but it is less common than in Myanmar, Russia and Thailand. As a neural tube defect the disease is characterized by a tumor protusion between the eyes or at the base of the nose. The protusion diameter is in the range of ca. 1.5 cm to ca. 8 cm. Although hypertelorism is common and smell as well as vision can be affected, the disease does not lead to serious debilitation or dislablement.

It is speculated that a teratogen consumed with fungus-contaminated \rightarrow rice during a critical period of pregnancy may interfere with the development of the embryo.

As yet there is only one supposed case consumption of blight-affected \rightarrow potatoes by pregnant women - but correct mycotoxicological justification is lacking.

Fruit juices (no specification) may contain the following \rightarrow mycotoxins: \rightarrow patulin incidence: 2/3, conc. range: $\leq 50 \ \mu g / kg$, country: Germany incidence: 12/58, conc. range: 5-15 $\mu g / kg$, country: Italy

Fruit products (no specification) may contain the following \rightarrow mycotoxins: \rightarrow patulin incidence: 18/52, conc. range: 5-32 µg/kg, country: Australia

Fruits (no specification) The high $\rightarrow a_w$ and nutrient content of ripe fruits and \rightarrow vegetables make them highly susceptible to the infection by toxigenic molds. At full maturity fruits are easily injured and predisposed to fungal attack.

The most important mycotoxigenic fungus on fruits is \rightarrow Penicillium expansum Link. The growth of this fungus leads to \rightarrow patulin contamination, especially in apples.

The contamination with \rightarrow Alternaria spp. one of the most common microorganisms responsible for the spoilage of fruits and vegetables may result in the production of copious amounts of \rightarrow mycotoxins. Mycotoxin formation is favored by the high moisture content of fruits so that all three groups of \rightarrow Alternaria mycotoxins are found. The incorporation of contaminated fruits into processed products, e.g. juices, preserves, sauces, due to faulty sorting procedures or neglect, is a potential health hazard.

According to Frisvad (1988), the following mycotoxins may be found in fruits and fruit products: *Alternaria* mycotoxins, \rightarrow Fusarium mycotoxins, patulin. Fruits may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin (no specification)

incidence: $6/157^*$, conc. range: 2-20 μ g / kg, country: Uruguay, *dried patulin

incidence: 4/74*, conc. range: nc, country: India, *dried

incidence: 1/1, conc.: 10 μ g/kg, country: UK

 \rightarrow zearalenone

incidence: 1/99*, conc.: > 200 µg/kg, country: Uruguay, *dried

→ apples, → bananas, → blueberries, → cherries, → cranberries, → lingonberries, → mandarin fruits, → mango, → oranges, → peaches

Fumonisin B₁ (Syn.: macrofusin) is a 2amino-12,16-dimethyl-3,5,10-trihydroxy-14,15-propane-1,2,3-tricarboxy icosane (\rightarrow mycotoxins) which was first isolated from \rightarrow Fusarium moniliforme Sheldon in 1988 (see Figure Fumonisin B₁).

CHEMICAL DATA

Empirical formula: C₃₄H₅₉NO₁₅, molecular weight: 721

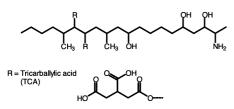
FUNGAL SOURCES see \rightarrow fumonisins

NATURAL OCCURRENCE \rightarrow Asparagus, \rightarrow beans, \rightarrow beer,

 \rightarrow bread, \rightarrow breakfast cereals, \rightarrow corn flakes, \rightarrow maize, \rightarrow maize-based thickeners, \rightarrow maize bran, \rightarrow maize chips, \rightarrow maize flakes, \rightarrow maize flour, \rightarrow maize grits, \rightarrow maize meal, \rightarrow maize products, \rightarrow maize snacks, \rightarrow maize, canned, \rightarrow maize, fiber cereal, \rightarrow maize, hominy, \rightarrow maize, popped, \rightarrow maize, puffed, \rightarrow maize, quality protein, \rightarrow maize, sweet, \rightarrow masa, \rightarrow milk, \rightarrow muffin mix, \rightarrow pop corn, \rightarrow rice, \rightarrow sorghum meal, \rightarrow starch, \rightarrow tortillas, \rightarrow tortilla chips For further information see \rightarrow fumonisins TOXICITY see \rightarrow fumonisins DETECTION see \rightarrow fumonisins Possible Mycotoxicosis see \rightarrow fumonisins

FURTHER COMMENTS

Stability / reduction: At atmospheric pressure chemical ammonia was ineffective for the detoxification of FB1-contaminated maize. However, at high pressure this treatment caused losses of FB₁ to almost 80%. Treatment of fumonisin contaminated maize with 2% ammonia at low pressure for 4 days, a process that successfully decontaminates aflatoxincontaminated maize, did not result in complete destruction of the mycotoxin. Calcium hydroxide was highly effective in removing FB₁ from contaminated maize while potassium hydroxide and hydrochloric acid hydrolyze FB₁ to HFB₁. FB₁ was destroyed by using sodium hypochlorite. The effect of ammoniation on



Fumonisin B₁

FB₁ reduction varies with experimental conditions. Potassium hydroxide and hydrochloric acid caused hydrolyzation of fumonisins to tricarballylic acid and a C₂₂ aminopolyol.

FB₁ losses during **baking** may be related to the nonenzymatic browning reaction. As the heat increases, more FB₁ (the primary amine group) reacts with free aldehyde or ketone groups in reducing sugars (i.e. glucose and fructose). However, commercial drying and baking temperatures in general are not sufficient to significantly reduce the FB₁ concentration in corn muffins whereas the fumonisin content of maize bread was lowered to almost 50%.

Neither drying nor normal food processing and cooking are effective in the destruction of FB₁.

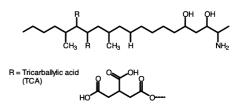
Fumonisin B₂ is a 2-amino-12,16-dimethyl-3,5-dihydroxy-14,15-propane-1,2,3tricarboxy icosane (\rightarrow mycotoxins) which was first isolated from \rightarrow Fusarium moniliforme Sheldon in 1988 (see Figure Fumonisin B_2).

CHEMICAL DATA Empirical formula: C₃₄H₅₉NO₁₄, molecular weight: 705

FUNGAL SOURCES see \rightarrow fumonisins

NATURAL OCCURRENCE

 \rightarrow Asparagus, \rightarrow beer, \rightarrow breakfast cereals, \rightarrow corn flakes, \rightarrow incarpina, \rightarrow mai ze, \rightarrow maize-based thickeners, \rightarrow maize flour, \rightarrow maize grits, \rightarrow maize meal,



Fumonisin B₂

- \rightarrow maize products, \rightarrow maize snacks,
- \rightarrow maize, fiber cereal, \rightarrow maize, hominy,
- \rightarrow maize, puffed, \rightarrow maize, quality protein,
- \rightarrow maize, sweet, \rightarrow masa, \rightarrow muffin mix,
- \rightarrow pop corn, \rightarrow rice, \rightarrow starch, \rightarrow tortillas, \rightarrow tortilla chips

For further information see \rightarrow fumonisins TOXICITY

see \rightarrow fumonisins

DETECTION

see \rightarrow fumonisins

Possible Mycotoxicosis see \rightarrow fumonisins

Fumonisin B₃ is a 2-amino-12,16-dimethyl-3,10-dihydroxy-14,15-propane-1,2,3tricarboxy icosane (mycotoxins) which was first isolated from \rightarrow Fusarium moniliforme Sheldon in 1988 (see Figure Fumonisin B₃).

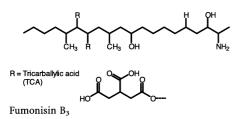
CHEMICAL DATA Empirical formula: C₃₄H₅₉NO₁₄, molecular weight: 705

FUNGAL SOURCES see \rightarrow fumonisins

NATURAL OCCURRENCE \rightarrow breakfast cereals, \rightarrow maize, \rightarrow maize flour, \rightarrow maize meal, \rightarrow maize, quality protein, \rightarrow rice, \rightarrow tortillas For further information see \rightarrow fumonisins. TOXICITY

see \rightarrow fumonisins

DETECTION see \rightarrow fumonisins



Possible Mycotoxicosis see \rightarrow fumonisins

Fumonisins are long-chain polyhydroxyl alkylamines containing two propane tricarboxyclic acid moieties which are esterified to hydroxyl groups on adjacent carbon atoms. \rightarrow Fumonisin B₁ is the most important of the fumonisins. \rightarrow Fumonisin B_2 and \rightarrow fumonisin B_3 are homologs but FB₂ lacks the hydroxyl at C-10 while FB₃ lacks the hydroxyl group at C-5. These three fumonisins account for most of the fumonisins that are both found in naturally contaminated \rightarrow maize as well as under cultivated conditions. At least 13 fumonisins, four B's (B_1, B_2, B_3) B_3 , B_4) having a free amine and three A's (A_1, A_2, A_3) which are amides, fumonisin C_1 , C_3 and FC_4 (analogs of FB_1 , FB_3 and FB₄, respectively) as well as fumonisin P_1 , P_2 , P_3 have been isolated from *F*. moniliforme. FA₁, FA₂ and FA₃ are the Nacetyl derivatives of FB₁, FB₂ and FB₃, respectively. Within each series differing hydroxyl substitution results in different fumonisins. FC₁, FC₃ and FC₄ lacking the C-1 terminal methyl group which is characteristic for the other fumonisins. In comparison to FC₁ the hydroxylated FC₁ (OH-FC₁) has one more hydroxy group at the C-3 position.

The fumonisins were first reported in South Africa (1988) and belong to the most recently described \rightarrow Fusarium mycotoxins. High rates of \rightarrow esophageal cancer in the rural population of South Africa and the death of many horses due to \rightarrow equine leukoencephalomalacia, mainly in New Caledonia, led to their detection. The involved feed was highly infested with \rightarrow Fusarium moniliforme Sheldon. Because researches were unable to find toxic substances in the usual organic extracts of *F. moniliforme* cultures, they concentrated on the aqueous fractions. The isolation of fumonisin B₁ succeeded in South Africa while independently fumonisin B₁ was isolated under the name \rightarrow macrofusin from culture material of *F. moniliforme*, which is responsible for equine leukoencephalomalacia in New Caledonia in 1989 (see Figure Fumonisins).

FUNGAL SOURCES

F. moniliforme, \rightarrow Fusarium proliferatum (Matsushima) Nirenberg (the main producers); *F. anthophilum*, *F. dlamini*, *F. globosum*, *F. napiforme*, *F. nygamai*, and *F. subglutinans* seems to be a non-consistent producer. *A. alternata* f. sp. *lycopersici* is also known for FB₁ production.

NATURAL OCCURRENCE

 \rightarrow beer, \rightarrow bread, \rightarrow breakfast cereals, \rightarrow chilli pickles, \rightarrow corn flakes, \rightarrow curry, \rightarrow curry paste, \rightarrow maize muffin, \rightarrow maize pops cereals, \rightarrow maize starch, \rightarrow maize, infant cereal, \rightarrow maize, infant cream corn, \rightarrow noodles, \rightarrow spices, \rightarrow tandoori, For further information see fumonisin B_1 , fumonisin B_2 , fumonisin B_3 These so-called "aflatoxins of the nineties" are widespread in maize and maizebased products in numerous countries of the world. FB₁, FB₂ and FB₃ are the major compounds produced in nature (\rightarrow food and feed). FB_1 is the predominating fumonisin in naturally-contaminated maize kernels with a ratio of 3:1 (FB₁:FB₂) and 12:1 (FB₁:FB₃) which corresponds to ca. 70% of the total fumonisin concentration detected. However, in vitro there are some isolates of F. moniliforme producing more FB₂ than FB₁. FB₄, FC₁ and FA₁-FA₃ are synthezised in relatively

minor quantities while the three latter ones do not occur naturally. In contrast, FC_1 , FC_3 and FC_4 as well as FB_4 have been detected in Korean moldy maize samples intended for animal consumption.

Fumonisin contamination of maize may be related to dry weather at or just prior to pollination. The contamination may occur world-wide but higher levels in food and feedstuff may be present in countries with a warm, dry climate. In countries having a cool, damp climate only low fumonisin levels are expected. However, some studies indicate that the contamination levels of maize and maize products are similar from country to country.

Whole kernel maize, grits and flour that undergo the mildest forms of processing are most frequently affected, usually showing medium (grits, flour) to high (kernels) fumonisin concentrations while maize \rightarrow bran is also affected. Low contamination or none at all occurred in highly processed maize-based productssuch as corn flakes, maize chips, corn pop cereals, tortillas and tortilla chips but in part recovery problems may be responsible.

This pattern of fumonisin distribution in maize and maize-based products agrees with the growth characteristics of F. moniliforme, which colonizes the tip and germ area of the kernel, just beneath the pericarp. In dry milled fractions the bran and germ fractions were highly contaminated with the fungus and fumonisins while the flour and flaking grit fractions contain low to medium mycotoxin concentrations. Almost no contamination was found in maize and tortilla chips, which may be explained in part by recovery problems, while hominy corn, tortillas and popcorn showed low contamination levels. It is under discussion whether the (apparent) loss of fumonisins by heating is due to degradation and loss of

toxicity or may result from ineffective detectability by current methods of extraction and analysis. It seems that corn starch as a product of the wet milling process is usually free of fumonisins. The widespread occurrence of fumonisins in Eastern and Southern Africa is documented by a positive of 92.5% of the maize samples analyzed. In these countries the daily maize intakes amounts to more than 200 g/person/day (FAO 1992) with peaks in Malawi (468.8 g), Zambia (418.6 g), and Zimbabwe (330.9 g). An daily average intake of 245 µg fumonisins was calculated for Zimbabwe's population on the basis of maize consumption. The high natural contamination of maize with fumonisins is of concern, particularly with respect to the much lower levels of other mycotoxins, like aflatoxin, T-2 toxin or zearalenone, present in food and feedstuffs (although these mycotoxins possess a relatively higher toxicity). Serious health implications may arise, taking into account that 10 and 100 µg fumonisin/g are dangerous to horses and pigs, respectively.

In north-eastern Italy an increasing risk of developing human esophageal cancer with increasing consumption of maize $(\rightarrow \text{ polenta})$ was observed. In animal tissues, so far, only trace amounts of fumonisins have been found. It seems that residues in \rightarrow meat, \rightarrow milk and eggs are not a problem.

TOXICITY

FB₁ causes severe animal diseases like leukoencephalomalacia (LEM, "hole in the head syndrome") in horses (ca. 10,000 µg FB₁ + FB₂/kg bw), pulmonary → edema syndrome (PES) in pigs (ca. 100,000 µg FB₁ + FB₂/kg bw), and liver cancer in rats (15,000 µg/kg bw). In the last case, FB₁ acts as a cancer initiator and promoter. Cattle seem to be less susceptible than pigs which are less susceptible than horses. Besides hepatotoxicity

FB₁ caused nephrotoxicity and diverse effects on the immunsystem in rodents. Toxic reactions also occurred in the case of turkey \rightarrow poultry and broiler chickens but laying hens seem to be not sensitive to low levels of fumonisin. Therefore, the FDA (Center for Veterinary Medicine) recommended that products with fumonisin levels greater than 1, 10, 30, and 50 mg/kg should not be fed to horses, pigs, beef \rightarrow cattle, and poultry, respectively. FB₂ and FB₃ showed hepatotoxic effects similar to FB1 and similar, although weak, cancer-initiating potential. In addition, esophageal cancer (EC) in humans has been observed in distinct areas of the world (Transkei / South Africa, Linxian and Cixian Counties/ northern China, northern Italy and south-eastern United States) where extremely high levels of fumonisins occurred in moldy home-grown maize and maizebased food products.

Since FB₁ inhibits the uptake of folate, it is also under discussion whether the fumonisins are involved in malformations of the central nervous system, e.g. neural tube defects. Such birth defects may be related to dietary exposure to fumonisin. In contrast to AFB₁, FB₁ is not \rightarrow mutagenic or \rightarrow genotoxic, whereas the cytotoxicity is low.

The fumonisins bear a remarkable structural similarity to the long-chain base sphingosine as a component of the longchain backbone of various sphingolipids. These are highly active components of cell membranes. The disruption of their metabolism may result in serious effects on cell behaviour, differentiation and growth. FB1 and FB2 were the first naturally occurring specific inhibitors of sphingolipid synthesis to be discovered. Fumonisins inhibit ceramide synthetase (sphingosine and sphinganine N-acyltransferase) resulting in an alteration in sphingolipid base ratios (sphinganine). This alteration causes massive liquefactive necrosis of the cerebral hemisphere. Neurological manifestations occur in horses, such as abnormal movements, aimless circling, lameness, etc. Interference with sphingolipid biosynthesis is the main cause of their toxicity in horses and probably also in pigs, as well as their tumor-promoting effects. In chicks (\approx 14 days old) fumonisins, perhaps together with other metabolites, may cause "spiking mortality syndrome" involving several neurological signs, reduced growth and mortality. The nixamalization product, the hydrolyzed fumonisin B_1 (HFB₁), resulting from cleavage of the tricarballylic side chains at C-14 and C-15, appears to be more toxic to rats than FB₁ itself since 50 mg/kg of FB_1 or 10 mg/kg of HFB_1 (maize, canned) possessed almost equal toxicity in rat feeding studies. However, the fact that HBF₁ did not initiate cancer in liver may be due to lack of absorption.

DETECTION

ELISA, GC-MS, HPLC, LC, TLC Besides chromatographic, mass spectrometric and immunochemical methods liquid chromatography is most commonly used in analysing food extracts for fumonisins.

FURTHER COMMENTS

The fumonisins are unusual \rightarrow mycotoxins in that they do not contain cyclic or ring groups. They are not unique in nature since structural similarities have been observed with the AAL-toxins, sphingofungins (antifungal agents isolated from \rightarrow Aspergillus fumigatus Fres.) and sphingosine (see above).

In addition, they are relatively water soluble (more soluble in acetonitrile-water or methanol, insoluble in organic solvents) but are as heat-stable as many other mycotoxins (see below).

Unfortunately visual assessment and subsequent separation of the ears into good and moldy lots is not sufficient to prevent fumonisin intake by humans because mycotoxins may also be present in visibly undamaged homegrown Transkeien maize.

Commercial maize hybrids in the US differ in their degree of fumonisin accumulation. Higher concentrations were detected in hybrids grown outside their adapted range.

20 °C was the best temperature for the production of FB_1 on corn.

Suitable storage conditions (e.g. low oxygen tension, kernel moisture content < 22%), reduce or prevent toxin production in stored maize.

Within 24 h, FB₁ is eliminated to more than 99% in the unmetabolized form in the faeces of rat. Traces are found only in the liver, kidney, urine and red blood cells. It is assumed that the adsorption of FB₁ is poor or there is a rapid elimination by biliary excretion.

Stability / Reduction: Fumonisins are appreciably stable during beer fermentation. If contaminated maize grits are used as brewing adjuncts only small decreases in FB₁ and FB₂ concentrations (\approx 20-30%) occurred during the fermentation. Fumonisin uptake by yeast was negligible. Although the distilled ethanol was free of FB₁ all the other fermentation products contained FB₁.

Like other mycotoxins, fumonisins are heat stable. Minor losses occurred after **heating** aqueous solutions of FB₁ and FB₂ at temperatures < 150 °C. Only higher temperatures (150 °C) were effective. Temperatures of ≈ 200 °C (60 min) are necessary to cause substantial fumonisin reduction in dry or moist corn meal. A partial reduction of the fumonisin concentration was detected in muffins that had been baked at 220 °C for 25 min. No reduction in FB₁ and FB₂ levels could be detected in whole milk heated for 30 min at 62 °C. **Canning** (121 °C for various times) of different maize products did not result in significant losses of fumonisins (\leq 15%). However, the apparent loss of fumonisin content in thermally processed foods may be due to matrix-related problems of recovery and detection by analytical methods.

During the nixtamalization process for manufacturing masa or tortilla flour at 100 °C, calcium hydroxide (0.01 M) causes the loss of the two propane-1,2,3-tricarboxylic acid (tricarballylic acid) moieties of FB₁, leading to hydrolyzed FB₁ (HFB₁). This amino pentol chain is found in commercial masa, tortilla chips and canned sweet corn, formed as a result of alkaline conditions and heating during processing. The toxicity of both HBF₁ and HBF_2 was higher than that of FB_1 and FB₂ when mammalian cell cultures and jimsonweed leaf bioassays were used. Removal of the corn fines (or screenings) from bulk shipments of corn by sieving reduced the total fumonisin levels down to almost 30%. This is due to the accumulation of fumonisins in the outer pericarp layers of broken kernels in these screenings (61,000-268,000 µg FB₁/kg) and (19,000-86,000 µg FB₂ / kg). The removal of fine particulate matter from bulk shipments of maize, prior to processing, might be an effective procedure for the preliminary decontamination of affected maize.

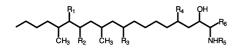
A further reduction is achieved by the **milling** process. Increasing refinement of maize meal means that combined fumonisin levels could be lowered by as much 95% in fine maize meal compared to maize screenings. Dry milling caused an accumulation of fumonisins in the bran, germ and fines fractions that are widely used in the production of animal feed, although bran is sometimes also used in certain breakfast cereals. Flaking grits, widely used in breakfast cereals and \rightarrow snack foods were relatively free of con-

tamination. However, decreasing grit size led to an increase in fumonisin concentration. During wet milling most of fumonisin was found in steep water, gluten fiber, and germ, whereas no detectable levels occurred in the starch fractions.

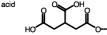
Steeping naturally contaminated corn in water or solutions of sodium bisulfite may reduce fumonisin levels.

Regulations: Based on their toxicological potential (carcinogenic in experimental animals) *F. moniliforme* toxins, including \rightarrow Fusarin C, have been classified as potential carcinogens for humans (class 2B carcinogens) by the IARC. In the EU there is still no legislation on fumonisin B₁ levels. Switzerland is the only country with a maximum tolerated level for fumonisins in maize produced for human consumption (sum of FB₁ and FB₂ \leq 1000 µg/kg). Since maize constitutes only a small component in the diet of

the first world population considerably higher tolerance levels may be adequate for sufficient for protecting of the population against these mycotoxins. However, in areas where maize is a staple food



Tricarballylic acid (TCA)



	R ₁	R ₂	R ₃	R_4	R ₅	R ₆
FB ₄	TCA	TCA	Н	Н	Н	CH3
FA ₁	TCA	TCA	OH	OH	COCH ₃	CH_3
FA ₂	TCA	TCA	Н	OH	COCH ₃	CH3
FA ₃	TCA	TCA	OH	Н	$COCH_3$	CH_3
FC ₁	TCA	TCA	OH	ОН	Н	Н
HHFB _{1a}	TCA	OH	OH	OH	Н	CH ₃
HHFB _{1b}	OH	TCA	OH	OH	Н	CH_3
AP ₁	ОН	OH	OH	OH	Н	CH ₃

Fumonisins. Structure and substituents of fumonisins

levels even lower than 1000 μ g/kg seem to be necessary.

Fungi Kingdom of Eukaryota, the true fungi

Fusaproliferin is a bicyclic sesterterpene $(3-[2-(acetyloxy)-1-methylethyl]-4,7,8,9,12,13,16,16a-octahydro-2,7-dihydroxy-6,10,14,16a-tetramethyl-1(3aH)-cyclopentacyclopentadecenone), characterized from a toxigenic strain maize culture of <math>\rightarrow$ Fusarium proliferatum (Matsushima) Nirenberg in 1993/1995 (see Figure Fusaproliferin).

CHEMICAL DATA Empirical formula: $C_{27}H_{40}O_5$, molecular weight: 444

Fungal Sources F. proliferatum

NATURAL OCCURRENCE

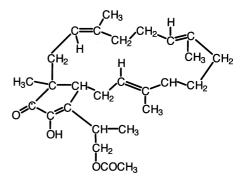
 \rightarrow maize

TOXICITY

 \rightarrow teratogenic, toxic to Artemia salina and mammalian cells

DETECTION HPTLC, TLC

Fusarenon X (Syn.: 4-acetylnivalenol, nivalenolmonoacetate, fusarenon) belongs to the group of naturally-occurring B \rightarrow trichothecenes (3 α ,7 α ,15-trihydroxy-



Fusaproliferin

 4β -acetoxy-12,13-epoxythrichothec-9-en-8-one). Fusarenon X was first isolated in 1967 and is produced by different species of the genus \rightarrow Fusarium (see Figure Fusarenon X).

CHEMICAL DATA

Empirical formula: $C_{17}H_{22}O_8$, molecular weight: 354

FUNGAL SOURCES

→ Fusarium equiseti (Corda) Sacc. sensu Gordon, → Fusarium graminearum Schwabe, → Fusarium oxysporum Schlecht., → Fusarium semitectum Berk. & Rav., → Fusarium sporotrichioides Sherb., → Fusarium sambucinum Fuckel (= *F. sul-phureum*),

NATURAL OCCURRENCE

 \rightarrow foods, \rightarrow garlic, \rightarrow maize, \rightarrow oats, \rightarrow wheat

Тохісіту

 LD_{50} (po): 4.4 mg/kg bw rat \rightarrow immunosuppressive, carcinogenic, cytotoxic, emetic, causes diarrhea, \rightarrow hypothermia, decreased respiratory rate (experimental animals)

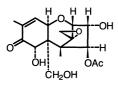
DETECTION

GC, MS, spectroscopy, TLC

FURTHER COMMENTS

Fusarenon X might occur more often in the warmer and subtropical parts of the world.

Fusarin C consists of a polyene chromophore with all the olefinic bonds in the *trans* configuration, linked in position C_{13} to a 2-pyrrolidone moiety and with a C_{13} - C_{14} epoxide group (2-ethylidene-11-[4-hydroxy-4-(2-hydroxyethyl)-2-oxo-6-





oxa-3-azabicyclo[3.1.0]-hex-1-yl]-4,6,10trimethyl-11-oxo-3,5,7,9-undecatetraenoic acid; methyl ester). It is the most important mycotoxin (\rightarrow mycotoxins) in the group of fusarins which include Fusarin A, D, E, F, X, Z. Fusarin C was first described in 1981, isolated from \rightarrow Fusarium moniliforme Sheldon (see Figure Fusarin C).

CHEMICAL DATA

Empirical formula: C₂₃H₂₉NO₇, molecular weight: 431

FUNGAL SOURCES

Different \rightarrow Fusarium species (13) such as \rightarrow Fusarium avenaceum (Fr.) Sacc., *F. crookwellense*, \rightarrow Fusarium culmorum (Wm. G. Smith) Sacc., \rightarrow Fusarium graminearum Schwabe, *F. moniliforme*, \rightarrow Fusarium sambucinum Fuckel, \rightarrow Fusarium sporotrichioides Sherb.

NATURAL OCCURRENCE

 \rightarrow maize

Visibly *Fusarium*-infected as well as healthy looking corn kernels in South Africa were affected. It was also found in maize from Linxian county, China.

TOXICITY

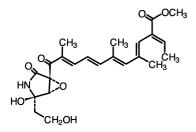
mutagenic as \rightarrow aflatoxin B₁ and \rightarrow sterigmatocystin, genotoxic, \rightarrow immunosuppressive, production and functioning of macrophages are inhibited Fusarin A and D are two less-toxic and non-mutagenic forms.

Although the biological activity of fusarin E is unknown (first described in 1991), its chemical structure may impart a comparable activity to that of fusarin C.

Detection HPLC

FURTHER COMMENTS

Fusarin C is one of the most unstable mycotoxins and therefore the significance of this fungal metabolite to the etiology of human illness is questionable. Although a moderate stability was established after storing contaminated ground



Fusarin C

maize at room temperature for 7 days in the dark, the high thermal instability led to an almost complete loss in \rightarrow maize meal as well as \rightarrow wheat flour during cooking / heating. No fusarin C was detected in maize meal muffins (230 °C) (\rightarrow maize muffin) made from contaminated maize kernels. Because stability of fusarin C decreases with increasing pH, thorough cooking at a slightly basic pH will effectively destroy most of this mycotoxin.

Fusarin C may be produced on soybeans and other cereals.

Fusariogenins → Fusarins

Fusariotoxicoses (in China)

Since 1961 the consumption of moldy \rightarrow wheat and \rightarrow maize in China has been linked with 35 outbreaks of toxicosis in man. Symptons like nausea, diarrhea, dizziness, and headache were accompanied by fever and disturbances of the nervous system 5 to 30 min after ingesting the incriminated \rightarrow grains. 26 outbreaks definitely occurred in the time between March and July and it seems that \rightarrow deoxynivalenol contaminated grain (wheat \leq 40,000 µg DON/kg, maize \leq 92,800 µg DON/kg) was the causal agent.

Fusarium anamorphic \rightarrow Hypocreaceae, teleomorphs \rightarrow Gibberella, \rightarrow Nectria Fusaria are adapted to a wide range of environmental conditions, which explains their ubiquitous distribution in diverse soil and organic substrates. They belong to the (ecological) group of \rightarrow field fungi requiring minimum $\rightarrow a_w$ values of 0.88-0.91 for growth in soil. However, they occasionally develop in stored \rightarrow cereals (especially in Scandinavia) when the moisture content is high (22-33%) and temperature is low. During wet summers, resulting in late harvest cereal, \rightarrow grains are primarily affected by Fusaria infections. The invaded kernels are characterized by shriveled, discolored kernels called scab, tombstone, or head blight. Due to climatic conditions and inadequate drying techniques in Scandinavian grains, Fusaria are quite common in stored cereals.

Toxigenic species often synthesize more than one mycotoxin, e.g. trichothecenes, with one or two of them being dominant. Strains from cold areas are usually more toxic than those from tropical and subtropical regions. Higher yields of toxic metabolites are produced after periods of low temperature. In the temperate countries of northern Europe, Canada and the northern regions of the USA, trichothecenes and zearalenone are more common than \rightarrow aflatoxins.

With respect to human and/or animal health problems \rightarrow Fusarium graminearum Schwabe, \rightarrow Fusarium moniliforme Sheldon, and \rightarrow Fusarium sporotrichioides Sherb. are the most important but more than 20 problematic species are known. The Fusaria produce more than 100 biologically active secondary metabolites which belong to different structural groups. Grains damaged by Fusarium spp. are considered as significantly toxic. Grain toxicity may be measured by the percentage of such damaged kernels in a given lot. From the standpoint of human exposure, mycotoxins such as \rightarrow trichothecenes (e.g. \rightarrow deoxynivalenol, \rightarrow nivalenol, \rightarrow T-2 toxin), \rightarrow zearalenone, as

well as the \rightarrow fumonisins, have attracted the most attention.

The enhanced production of trichothecenes at low temperatures led to the wrong assumption that these mycotoxins served as an agent of war ("yellow rain" = bee faeces containing a mixture of trichothecenes) in Southeast Asia. However, these trichothecenes as well as zearalenone have been isolated from grains grown in this tropical part of the world.

Fusarium avenaceum (Fr.) Sacc. teleomorph: *Gibberella avenacea* Cook is of worldwide distribution and possesses a very broad host range such as \rightarrow cereals, broad bean (\rightarrow beans), \rightarrow potatoes. This species may produce \rightarrow mycotoxins such as antibiotic Y,

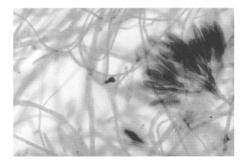
 \rightarrow moniliformin, \rightarrow zearalenone.

Fusarium crookwellense Burgess, Nelson &

Toussoun may produce \rightarrow mycotoxins such as acetylnivalenol, \rightarrow fusarin C, \rightarrow nivalenol, zearalenols, \rightarrow zearalenone.

Fusarium culmorum (W. G. Smith) Sacc. teleomorph: unknown

is a pathogen of \rightarrow wheat, \rightarrow rye, \rightarrow barley, \rightarrow oats and \rightarrow maize, often co-occurring with \rightarrow Fusarium graminearum Schwabe. *F. culmorum* is of worldwide distribution but prefers cooler climatic regions such as northern Europe and southern Australia (see Figure *Fusarium*



Fusarium culmorum (W.G. Smith) Sacc.

culmorum (W.G. Smith) Sacc.). It cooccurs with *F. graminearum* as a causal agent of *Fusarium* head blight, invading cereal heads at the time of flowering. \rightarrow Deoxynivalenol and \rightarrow zearalenone are the primarily produced \rightarrow mycotoxins but some more toxic metabolites such as \rightarrow butenolide, culmorin, \rightarrow diacetoxyscirpenol, \rightarrow fusarenon X, \rightarrow HT-2 toxin, \rightarrow neosolaniol, \rightarrow nivalenol, \rightarrow T-2 toxin may be synthesized.

Fusarium equiseti (Corda) Sacc. sensu Gordon teleomorph: Gibberella intricans Wollenw. as a weak pathogen of \rightarrow cereals, \rightarrow vegetables, legumes, and \rightarrow fruits prefers subtropical and tropical climates. \rightarrow Grains harvested and stored with high moisture contents are likely to be invaded. Several animal diseases like degnala disease, fescue foot, \rightarrow bean hulls poisoning, and tibial dyschondroplasia are probably due to the \rightarrow mycotoxins of F. equiseti. Leukemia in man may be another disease caused by this fungus. F. equiseti may produce mycotoxins such as \rightarrow 15-acetylscirpentriol, \rightarrow butenolide, \rightarrow diacetoxyscirpenol, equisetin, \rightarrow fusarenon X, \rightarrow fusarochromanone, \rightarrow HT-2 toxin, \rightarrow neosolaniol, \rightarrow nivalenol, \rightarrow scirpentriol, \rightarrow T-2 toxin, \rightarrow zearalenone.

Fusarium graminearum Schwabe teleomorph: Giberella zeae (Schw.) Petch. F. graminearum produces several dozen metabolites, four or five accumulate in quantity most often in \rightarrow cereals grown in warmer climates (see Figure Fusarium graminearum Schwabe). This species is divided into Groups I and II. Strains of Group II commonly infest cereal \rightarrow grains especially \rightarrow wheat and \rightarrow maize (scab or head blight) and produce significant amounts of $B \rightarrow$ trichothecenes and \rightarrow zearalenone (up to 60,000,000 μ g/kg). They have been separated into the "NIV-chemotype" (\rightarrow nivalenol and \rightarrow fusarenon X producers) and



Fusarium graminearum Schwabe. Macroconidia of Fusarium graminearum

the "DON-chemotype" which is further separated into chemotype IA (\rightarrow deoxynivalenol and \rightarrow 3-acetyldeoxynivalenol) and "chemotype IB" (DON and \rightarrow 15acetyldeoxynivalenol producers). \rightarrow T-2 toxin production occurs at a low optimal temperature of 6-12 °C.

The following further \rightarrow mycotoxins such as 4-acetamido-2-butenoic acid, \rightarrow butenolide, \rightarrow diacetoxyscirpenol, 3,15-dihydroxy-12,13-epoxythrichothec-9-ene-8one, \rightarrow HT-2 toxin, \rightarrow monoacetoxyscirpenol, \rightarrow neosolaniol, \rightarrow nivalenol, and T-2 toxin may be produced.

F. graminearum is involved in the following \rightarrow mycotoxicosis:

feed refusal and emetic syndromes, \rightarrow F-2 toxicosis, \rightarrow red mold disease

Fusarium moniliforme Sheldon teleo-

morph: Gibberella fujikuroi (Swada) Ito in Ito & K. Kimura is widespread in humid and subhumid temperate zones. It is found also in subtropical and tropical zones, but is uncommon in cooler temperate zones. It is one of the most prevalent seedborne fungi of \rightarrow maize in most dry and warm (corn growing) areas of the world, but crops like \rightarrow peanuts, \rightarrow rice, \rightarrow sorghum, \rightarrow soybeans, sugar-cane, \rightarrow bananas etc. are also attacked.

The most important \rightarrow mycotoxins, the \rightarrow fumonisins, appear to be less common

in \rightarrow maize grown in cooler climates, e.g. northern Europe and Canada, but are of general significance in maize of warm and dry climatic regions, e.g. South Africa, China, Italy. Mating population A of *F. moniliforme* as well as the D mating population of *G. fujikuroi* (*F. proliferatum*) are much better FB₁ producers than the F population of *F. moniliforme*. Several animal diseases like ELEM (horses), PES (swine), hepatocarcinogenicity in rats etc. are caused by these mycotoxins, while EC (\rightarrow esophageal cancer) in man is probably due to these toxic fungal metabolites.

The following mycotoxins such as \rightarrow diacetoxyscirpenol, fusaric acid, fusarins (\rightarrow fusarin C), \rightarrow moniliformin, \rightarrow T-2 toxin, \rightarrow zearalenone may also be produced.

Fusarium mycotoxicosis \rightarrow akakabi-byo disease, \rightarrow alimentary toxic aleukie, \rightarrow Kashin-Beck disease, \rightarrow moldy corn toxicosis, \rightarrow onyalai, \rightarrow pellagra, \rightarrow premature thelarche. These diseases are predominantly found in the temperate regions of the world due to \rightarrow Fusarium mycotoxins. Temperatures of 8 °C and grain humidities between 20-25 °C, especially in cold rainy summers contribute to the occurrence of these \rightarrow mycotoxicosis.

Fusarium mycotoxins \rightarrow Fusarium spp. are well known producers of the \rightarrow thrichothecenes, as well as the estrogenic mycotoxin, \rightarrow zearalenone. Food-relevant *Fusarium* \rightarrow mycotoxins are e.g. \rightarrow 3acetyldeoxynivalenol, \rightarrow 15-acetyldeoxynivalenol, \rightarrow 15-acetylscirpentriol, \rightarrow butenolide, \rightarrow deoxynivalenol, \rightarrow diacetoxyscirpenol, \rightarrow fumonisins, \rightarrow fusarenon X, \rightarrow HT-2 toxin, \rightarrow moniliformin, \rightarrow neosolaniol, \rightarrow nivalenol, \rightarrow T-2 toxin, \rightarrow zearalenone

Fusarium nivale (Fr.) Ces. teleomorph: unknown

is a (seedborne) pathogen of cereal \rightarrow grains, particularly under snow cover, prefering colder to temperate climates as found in e.g. Asia, Australia, Europe, and North America. This "snow mold" may produce \rightarrow mycotoxins such as \rightarrow deoxynivalenol, \rightarrow 3-acetyldeoxynivalenol, \rightarrow zearalenone.

Fusarium oxysporum Schlecht. emend. Snyd.

& Hansen teleomorph: unknown is the most economically important member of the genus *Fusarium* with a cosmopolitan distribution. Pathogenic strains are involved in damping-off diseases and cause vascular wilts in different crop plants (e.g. \rightarrow cereals). This fungus also plays a role in the \rightarrow Moldy sweet potato toxicosis. *F. oxysporum* may produce \rightarrow mycotoxins such as \rightarrow diacetoxyscirpenol, diacetylnivalenol, \rightarrow 7 α ,8 α -dihydroxydiacetoxyscirpenol, \rightarrow fusarenon X, enniatins, fusaric acid, 7-hydroxydiacetoxyscirpenol, \rightarrow moniliformin, \rightarrow neosolaniol, \rightarrow T-2 toxin?, \rightarrow zearalenone.

Fusarium poae (Peck) Wollenw. teleo-

morph: unknown

This species is of wide geographical distribution (predominantly temperate regions) often co-occurring with \rightarrow Fusarium sporotrichioides Sherb.. It has numerous hosts (cereal \rightarrow grains) and is a weak parasite or saprophyte after the death of cereal host plants (see Figure Fusarium poae (Peck) Wollenw.) \rightarrow T-2 toxin and other \rightarrow trichothecenes are produced at low optimal temperatures (6-12 °C), especially during freezing and thawing conditions in overwintering unharvested crops or during storage. Probably due to the production of type A trichothecenes it might be involved in \rightarrow Alimentary toxic aleukia, \rightarrow Moldy corn toxicosis and \rightarrow Kashin-Beck disease (Urov Disease).



Fusarium poae (Peck) Wollenw.

F. poae may produce \rightarrow mycotoxins such as \rightarrow butenolide, \rightarrow diacetoxyscirpenol, \rightarrow HT-2 toxin, \rightarrow neosolaniol, "poin" (water soluble substance, no structure elucidation, contamination with trichothecenes), \rightarrow T-2 toxin, T-2 tetraol.

Fusarium proliferatum (Matsushima) Niren-

berg teleomorph: unknown This taxon was distinguished only recently (1976) from what may now be considered the \rightarrow Fusarium moniliforme Sheldon complex. In consequence, there are similarities with that fungus concerning hosts, pathogenic associations with \rightarrow maize, fumonisin production and toxicity on \rightarrow maize.

This species, often misidentified as *F.* moniliforme, is cosmopolitan but predominant in tropical and subtropical countries, as well as in greenhouses in temperate zones and in a wide range of host plants (e.g. \rightarrow rice, \rightarrow fruits). *F. proliferatum* may produce \rightarrow mycotoxins such as \rightarrow fumonisins, fusaric acid, \rightarrow fusarin C, \rightarrow moniliformin, naphthoquinone pigments.

Fusarium sambucinum Fuckel (Syn.: *Fusarium sulphureum*) teleomorph: *Gibberela pulicaris* (Fr.) Sacc.

This ubiquitous species, which is more common in the northern but less frequently in the southern hemisphere has a wide host range, including stored → fruits and → potatoes. The involvement in human → esophageal cancer is discussed. The following → mycotoxins such as 4-→ acetoxyscirpenol, 4-acetoxyscirpenediol, 8-acetylneosolaniol, → butenolide?, → diacetoxyscirpenol, → fusarenon X, → monoacetoxyscirpenol, → nivalenol?, → sambutoxin, triacetoxyscirpenol, → zearalenone may be produced.

Fusarium sporotrichioides Sherb. teleomorph: unknown

This species is almost exclusively found in temperate to cold areas of the world on a wide variety of host plants, e.g. \rightarrow cereals and their products, stone \rightarrow fruits. It often co-occurs with \rightarrow Fusarium poae (Peck) Wollenw. in overwintered cereals. F. sporotrichioides is the principal agent of \rightarrow Alimentary toxic aleukia (ATA) and involved in \rightarrow Moldy corn toxicosis, fescue foot, \rightarrow Akakabi byo disease, \rightarrow Bean hulls poisoning. Mycotoxin production occurs at low temperatures, between 4 and 1.5 $^\circ \mathrm{C}$ but the optimum temperature seems to be 1.5-4 °C. The following \rightarrow mycotoxins such as acetyl T-2 toxin, \rightarrow butenolide, \rightarrow deoxynivalenol, \rightarrow diacetoxyscirpenol, \rightarrow diacetylnivalenol, \rightarrow fusarenon X, \rightarrow HT-2 toxin, \rightarrow neosolaniol, \rightarrow nivalenol, NT-1 toxin (= T-1 toxin: 4β, 8α-diacetoxy-3α,15-dihydroxy-12,13-epoxytrichothec-9-ene), NT-2 toxin (4β-acetoxy-3α, 8α,15-trihydroxy-12,13epoxytrichothec-9-ene), \rightarrow T-2 toxin, T-2 tetraol, \rightarrow zearalenone may be produced.

Fusarium sulphureum → Fusarium sambucinum Fuckel

Fusarochromanone (Syn.: TDP-1) is a water-soluble chromone derivative con-

taining an amino group at C-5 and a side chain at C-6 (5-amino-6-(3-amino-4hydroxy-1-oxobutyl)-2,3-dihydro-2,2dimethyl-4H-1-benzopyran-4-one). As a metabolite of \rightarrow Fusarium equiseti (Corda) Sacc. Sensu Gordon, it was first isolated and described in 1986 (see Figure Fusarochromanone).

CHEMICAL DATA

Empirical formula: $C_{15}H_{20}N_2O_4$, molecular weight: 292

Fungal Sources F. equiseti

NATURAL OCCURRENCE It should be present in \rightarrow cereals, viz. \rightarrow maize and \rightarrow wheat.

Тохісіту

tibial dyschondroplasia in cattle, chickens, dogs, horses, pigs, and turkeys; hatching reduction of fertile eggs (experimental conditions)

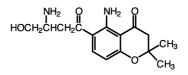
Detection

fluorescence detection, HPLC

Possible Mycotoxicosis It may be involved in the \rightarrow Kashin-Beck disease.

Further Comments

There are two derivatives: TDP-2 the C-3'-N-acetyl derivative, TDP-6 containing a hydroxyl group on C-3' and a methoxyl group on C-4'.



Fusarochromanone

G

111

Gabi → tubers

Galgant (Alpinia officinarum Hance) is a ginger-like spice. Glagant may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/4, conc.: $\leq 5 \ \mu g / kg$, country: Germany \rightarrow spices

Garlic may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/6, conc.: 12 µg/kg, country: India \rightarrow aflatoxin B₂ incidence: 1/6, conc.: 15 µg/kg, country: India \rightarrow aflatoxin G₁ incidence: 1/6, conc.: 10 µg/kg, country: India \rightarrow deoxynivalenol incidence: 1/4 conc.: $14 \mu g/kg$, country: UK \rightarrow fusarenon X incidence: 1/4, conc.: 5 µg/kg, country: UK \rightarrow nivalenol incidence: 1/4, conc.: 21 µg/kg, country: UK \rightarrow spices

Garlic onions may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 2*/58, Ø conc.: 67 µg/kg, country: Thailand, *total Ø conc.: 60 µg/kg AFB₁, AFB₂, AFG₁, AFG₂ \rightarrow spices

Garlic pickle may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂)

incidence: nc/4, conc. range: 0.2-0.6 $\mu g / kg$, country: UK \rightarrow ochratoxin A incidence: 3/4, conc. range: 0.9-2.5 $\mu g / kg$, country: UK \rightarrow zearalenone incidence: 1/4, conc.: 3.8 $\mu g / kg$, country: UK \rightarrow spices

Garlic powder may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 1/41*, conc.: 3.3 µg/kg, country: UK, *imported, miscellaneous \rightarrow spices

genotoxic changes the genom

Gigantic acid (Syn.: \rightarrow patulin)

Ginger is a dried rhizome of tropical origin. Ways of mycotoxin contamination have not yet been elucidated. Ginger may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 8/15, conc. range: < 2.5-25 μg/kg, country: India incidence: 3/5, conc. range: 1.4-6.5 μ g/kg, Ø conc.: 4.03 μ g/kg, country: USA \rightarrow aflatoxin B₂ incidence: 1/5, conc.: 0.2 µg/kg, country: USA \rightarrow aflatoxin G₁ incidence: 1/5, conc.: 2.5 µg/kg, country: USA \rightarrow aflatoxin G₂ incidence: 1/5, conc.: $0.2 \mu g/kg$, country: USA \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 3/41*, conc. range: 1.3-8.4 μ g / kg, Ø conc.: 3.9 μ g / kg, country: UK, *imported, miscellaneous \rightarrow spices incidence: nc/4, conc. range: 4.2-13.5 μg/kg, country: UK

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Ginger

incidence: 2/3, conc. range: $\leq 2 \ \mu g / kg$, country: USA aflatoxins (no specification) incidence: $1/3^*$, conc.: 2 µg/kg, country: USA, *imported → deoxynivalenol incidence: 1/4, conc.: 9 μ g/kg, country: UK \rightarrow neosolaniol incidence: 1/4, conc.: 23 μ g/kg, country: UK \rightarrow nivalenol incidence: 1/4, conc.: $34 \mu g / kg$, country: UK \rightarrow ochratoxin A incidence: 2/4, conc. range: 2.1-7.5 μg/kg, country: UK \rightarrow T-2 toxin incidence: 1/4, conc.: 18 µg/kg, country: UK \rightarrow spices

Goose may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A incidence: 5/12, conc. range: ≤ 0.1 μ g/kg, Ø conc.: 0.03 μ g/kg, country: Denmark incidence: 4/12*, conc. range: ≤ 0.06 μ g/kg, Ø conc.: 0.02 μ g/kg, country: Denmark, *goose liver \rightarrow meat

Grains (no specification) Grinding destroys the protective outer testa of \rightarrow cereals and thus enables the rich nutrients inside to be colonized by mycotoxin-producing fungi. Therefore, ground \rightarrow grains are often more contaminated than intact grains. Ca. 25% of the strains of \rightarrow Aspergillus and \rightarrow Penicillium isolated from grain are able to produce \rightarrow ochratoxin A besides other \rightarrow mycotoxins. OTA levels seem to be a good indicator of proper storage of grain. Grains may contain the following \rightarrow mycotoxins:

→ aflatoxins

incidence: 19/3489, Ø conc.: 5 μg/kg, country: USA \rightarrow deoxynivalenol incidence: 2/17, conc. range: 20-130 μg/kg, country: Germany incidence: 4/11, conc. range: 420-520 μ g/kg, Ø conc.: 470 μ g/kg, country: Sweden → nivalenol incidence: 57/190*, conc. range: 20-290 μg/kg, country: Germany, *moldy \rightarrow ochratoxin A incidence: 2/49, conc.: 18-22 µg/kg, country: Germany \rightarrow T-2 toxin incidence: 9/230, conc. range: 10-50 µg/kg, country: Finland \rightarrow zearalenone incidence: 9/114, conc. range: 5-30 μg/kg, country: Austria incidence: 18/51, conc. range: 10-500 μg/kg, country: Germany incidence: 3/584, conc. range: 200-1200 μ g / kg, country: Poland incidence: 26/1417, conc. range: > 20 μ g / kg, country: UK \rightarrow barley, \rightarrow buckwheat, \rightarrow cereals, \rightarrow cereal products, \rightarrow maize, \rightarrow millet, \rightarrow oats, \rightarrow rice, \rightarrow rye, \rightarrow sorghum, \rightarrow triticale, \rightarrow wheat

Grape juice may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A incidence: 1/6*, conc.: 0.73 µg/kg, country: Germany, *red incidence: 12/14*, conc.: $\leq 4.7 µg/kg$, country: Germany, *white incidence: 6/7*, Ø conc.: 0.218 µg/l, country: Switzerland, *red, imported incidence: 2/3*, Ø conc.: ca. 0.004 µg/kg, country: Switzerland, *white, partly imported incidence: 6/18*, conc. range: < 0.005-0.11 µg/l, country: Switzerland, *white, red, rosé → patulin incidence: 8/8*, conc. range: 360-4200 μ g / kg, Ø conc.: 1500 μ g / kg, country: Canada, *moldy incidence: 21/55, conc. range: 1-230 μ g / l, country: Germany incidence: 8/16, conc. range: 1-8 μ g / l, country: UK → apple juice, → breakfast drinks, → fruit juice, → fruits, → soft drinks

Groundnut toffee is an Indian peanut based snack. It consists of the crashed kotyledons of the \rightarrow peanuts, without seedcoat, which are cooked mild in hot concentrated jaggery syrup. The aflatoxin contamination may be lower than that of \rightarrow bondakaledkai. This may result from a certain degree of cleaning of the infested seeds. Groundnut toffees may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 19/67, conc. range: 2-400 μ g / kg, country: India \rightarrow aflatoxin B₂ incidence: 3/67, conc. range: 3-120 μ g / kg, country: India \rightarrow congressbele

Groundnuts → peanuts

Gushing It could be shown that commercial beers (\rightarrow beer) suspected of gushing, had significantly higher concentrations of \rightarrow deoxynivalenol compared with non-gushing beers.

H

Ham may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/1, conc.: 100 µg / kg, country: Germany \rightarrow ochratoxin A incidence: 60/206*, conc. range: 40-70 µg / kg, country: Yugoslavia, *total of smoked \rightarrow meat

Hare (wild)

may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ \rightarrow incidence: 89/168*, conc. range: 0.3-1.421 µg / kg, Ø conc.: 0.407 µg / kg, country: Czechoslovakia, *liver incidence: 94/168*, conc. range: 0.3-3.21 µg / kg, Ø conc.: 0.658 µg / kg, country: Czechoslovakia, *kidney \rightarrow meat

Hazelnuts (no specification) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/199*, conc.: 325 µg/kg, country: Finland, *imported incidence: 18/29*, conc. range: 5-50,000 μg/kg, country: Germany, *moldy incidence: 1/3*, conc.: 0.6 µg/kg, country: UK, *shelled incidence: 11/142*, conc. range: 2-100 μ g / kg, Ø conc.: 33 μ g / kg, country: USA, *imported \rightarrow aflatoxin B₂ incidence: 1/199*, conc.: 29 µg/kg, country: Finland, *imported \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 3/35*, conc. range: 6-10 μg/kg, country: Sweden *imported; edible, possibly edible and inedible \rightarrow nuts incidence: 2/18*, conc. range: 0.5-5 μ g/kg with a maximum of 0.7 μ g/kg, country. UK, *in-shell

→ aflatoxins (no specification) incidence: 18/20, conc. range: 25-175 μ g/kg, country: Egypt → ochratoxin A incidence: 3/57, conc. range: \leq 4.7 μ g/kg, country: Germany incidence: 2/11, conc. range: \leq 1.49 μ g/kg, country: Germany nuts

Hematuria blood in the urin

Hemorrhage bleeding, escape of blood

Hemorrhagic aleukia (Syn.: \rightarrow Alimentary toxic aleukia)

Hemorrhagic syndrome \rightarrow Alimentary toxic aleukia, \rightarrow Moldy corn toxicosis

Hens may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A Levels up to 29 µg/kg were found in the muscle of hens and chickens collected at a slaughterhouse. The birds had been rejected because of \rightarrow nephropathy. \rightarrow meat

hepatic pertaining to the liver

Hepatitis inflammation of the liver

hiptagenic acid $\rightarrow \beta$ -nitropropionic acid

Holy fire \rightarrow Ergotism

Hot dog The \rightarrow aflatoxins detected in hot dogs derive from the use of mycotoxin-contaminated \rightarrow spices and/or the incorporation of aflatoxin producers. Hot dog may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/25, conc.: 5 µg/kg, country: Egypt \rightarrow aflatoxin B₂ incidence: 1/25, conc.: 2 µg/kg, country: Egypt \rightarrow meat

M. Weidenbörner, *Encyclopedia of Food Mycotoxins* © Springer-Verlag Berlin Heidelberg 2001 **HT-2 toxin** is a 3α , 4β -dihydroxy-4, 15diacetoxy-8a-(3-methylbutyryloxy)-12,13-epoxytrichothec-9-ene which belongs to the trichothecene (\rightarrow trichothecenes) \rightarrow mycotoxins as a metabolite of \rightarrow Fusarium spp. (see Figure HT-2 toxin).

CHEMICAL DATA

Empirical formula: C₂₂H₂₃O₈, molecular weight: 424

FUNGAL SOURCES

Fusarium acuminatum, → Fusarium graminearum Schwabe, \rightarrow Fusarium poae (Peck) Wollenw., → Fusarium sporotrichioides Sherb.

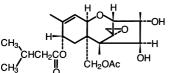
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NATURAL OCCURRENCE
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- \rightarrow barley, \rightarrow chilli powder, \rightarrow curry,
- \rightarrow maize, \rightarrow oats, \rightarrow rye, \rightarrow soybean,
- \rightarrow wheat
- TOXICITY

dermatoxic (similar to \rightarrow T-2 toxin) inhibition of the initiation step in protein synthesis LD $_{50}$ (ip): 9 mg/kg bw mice DETECTION

GC, MS, spectroscopy, TLC

Human breast milk The ingestion of aflatoxin-contaminated (\rightarrow aflatoxins) foods by humans will result in the elimination of variable levels of the toxin in body fluids or the accumulation in the tissue. This is currently a considerable problem for people living in tropical and subtropical countries because \rightarrow aflatoxin B₁ and the corresponding metabolites in human blood and breast \rightarrow milk represent a serious health hazard to the mother, to the fetus, and to newborn infants. The



HT-2 toxin

 \rightarrow aflatoxin M₁ contamination of breast milk is mainly caused by the consumption of food of plant origin, e.g. \rightarrow peanuts. \rightarrow maize. EU data indicate an ochratoxin A contamination between 0.007-0.58 µg/l human milk. Breast-fed infants may ingest (very) high levels of OTA. Human breast milk may contain the following \rightarrow mycotoxins: \rightarrow aflatoxicol incidence: 3/264, conc. range: 0.64-0.27 µg/l, country: Ghana, Nigeria aflatoxin B_1 incidence: 17/264, conc. range: 0.13-8.218 µg/l, country: Ghana, Nigeria \rightarrow aflatoxin B₂ incidence: 2/264, conc. range: 0.04-0.05 µg/l, country: Ghana, Nigeria aflatoxin M₁ incidence: 2/2, conc. range: 0.17-0.79 μ g/l, Ø conc.: 0.48 μ g/l, country: Algeria incidence: 11/73, conc. range: 0.028-1.031 μ g / l, country: Australia incidence: 1/1, conc.: 0.158 µg/l, country: Bahrain incidence: 6/6, conc. range: 0.006-0.174 μ g/l, Ø conc.: 0.061 μ g/l, country: Bangladesh incidence: 48/48, conc. range: 0.004-0.72 μ g/l, Ø conc.: 0.092 μ g/kg, country: Egypt incidence: 90/264, conc. range: 0.02-1.816 μ g/l, country: Ghana incidence: 163/510, conc. range: 0.005-1.379 µg/l, country: Ghana incidence: 48/48, conc. range: 0.004-0.6 μ g/l, Ø conc.: 0.099 μ g/l, country: India incidence: 2/2, conc. range: 0.003-0.051 μ g/l, Ø conc.: 0.027 μ g/l, country: Indonesia incidence: 3/3, conc. range: 0.051-1.6 μ g/l, Ø conc.: 0.58 μ g/l, country: Iran incidence: 2/2, conc. range: 0.008-0.014 $\mu g/l, \emptyset$ conc.: 0.011 $\mu g/l$, country: Iraq

incidence: 42/42 conc. range: 0.002-0.88 μ g/l, Ø conc.: 0.122 μ g/kg, country: Jordan incidence: 53/191, conc. range: 0.005-1.379 µg/l, country: Kenya incidence: 15/15, conc. range: 0.014-1.0 μ g/l, Ø conc.: 0.181 μ g/l, country: Lebanon incidence: 3/3, conc. range: 0.007-0.15 μ g/l, Ø conc.: 0.056 μ g/l, country: Morocco incidence: 6/6, conc. range: 0.07-0.978 μ g/l, Ø conc.: 0.35 μ g/l, country: Oman incidence: 44/44, conc. range: 0.002-1.1 μ g/l, Ø conc.: 0.178 μ g/l, country: Pakistan incidence: 54/55, conc. range: 0-0.84 μ g/l, Ø conc.: 0.115 μ g/l, country: Palestine incidence: 2/2, conc. range: 0.25-0.58 μ g/l, Ø conc.: 0.415 μ g/l, country: Philippines incidence: 3/7, conc.: nc, country: Philippines incidence: 2/2, conc. range: 0.058-0.395 μ g/l, Ø conc.: 0.227 μ g/l, country: Saudi Arabia incidence: 18/18, conc. range: 0.002-1.0 μ g/l, Ø conc.: 0.217 μ g/l, country: Somalia incidence: 37/99, conc. range: 0.005-1.379 μg/l, country: Sudan incidence: 44/44, conc. range: 0.003-2.1 μ g/l, Ø conc.: 0.285 μ g/l, country: Sudan incidence: 13/99, conc. range: 0.005-0.064 μ g/l, country: Sudan incidence: 36/36, conc. range: 0.003-0.8 μ g/l, Ø conc.: 0.204 μ g/l, country: Syria incidence: 10/64, conc. range: 0.3-1.3 μ g/l, country: UAE incidence: 37/37, conc. range: 0.009-3.0 μ g/l, Ø conc.: 0.412 μ g/l, country: UEA incidence: 5/11, conc. range: 0.039-1.736 μ g / l, country: Thailand incidence: 1/1, conc.: 0.02 µg/l, country: The Netherlands

incidence: 27/28, conc. range: 0-1.6 μ g/l, \emptyset conc.: 0.17 µg/l, country: Yemen incidence: 6/64, conc. range: $\leq 0.05 \ \mu g / l$, country: Zimbabwe \rightarrow aflatoxin M₂ incidence: 18/264, conc. range: 0.016-2.075 μg / l, Ghana, Nigeria incidence: 11/99, conc. range: 0.003-0.020 µg/l, country: Sudan aflatoxin M1 & M2 incidence: 13/99, conc. range: 0.003-0.084 µg/l, country: Sudan \rightarrow ochratoxin A incidence: 4/36, conc. range: 0.017-0.03 μ g/l, Ø conc.: 0.024 μ g/l, country: Germany incidence: 9/50, conc. range: 1.7-6.6 μg/l, country: Italy incidence: 22/111, conc. range: 0.1-12 μg/l, country: Italy incidence: 38/115, conc. range: 0.001-0.13 µg/l, country: Norway incidence: 23/40, conc. range: 0.01-0.04 μ g / l, country: Sweden ochratoxin A methyl ester incidence: 4/40, conc. range: 0.01-0.04 μ g/l, country: Sweden \rightarrow dairy products

Human hepacellular carcinoma \rightarrow Aflatoxin B₁ as an extremely potent hepatocarcinogen, is distributed in human foodstuffs especially in sub-Saharan African countries (e.g. Kenya, Mozambique, Swaziland) and southeast Asia (Thailand), where a high incidence of liver cancer can be found. Epidemiological studies showed a highly significant positive correlation between the liver-cancer rate and the level of dietary aflatoxin intake. However, infection with hepatitis B virus may predispose people for primary hepatocellular carcinoma. Although some other agents may also be involved in the development of this disease, interaction between the hepatitis B virus and aflatoxin appears the most plausible explanation available.

Human milk → human breast milk

Human serum About 50% of the European human sera investigated showed contamination with \rightarrow ochratoxin A (< 0.1-57 ng OTA / ml serum), with a slightly higher incidence in rural areas. OTA positive human blood sera also occurred in Canada. There are three main causes for this high contamination rate:

- long biological half-time of OTA which is bound to serumalbumines,
- intake of OTA-contaminated foodstuff,
- inhalation of OTA-contaminated conidia.

Hydnocarpus laurifolia (medicinal seeds) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: nc/nc, conc. range: 20-650 µg/kg, country: India \rightarrow citrinin incidence: nc/nc, conc. range: 10-490 µg/kg, country: India

Hydrolyzed fumonisin B₁ (Abbr.: HFB₁, \rightarrow fumonisins)

4-Hydroxyochratoxin A Although this mycotoxin is produced by \rightarrow Penicillium viridicatum Westling this compound primarily seems to be a detoxification product in animals (e.g. rats) dosed with \rightarrow ochratoxin A.

3'-Hydroxy HT-2 toxin is a metabolite of \rightarrow HT-2 toxin and a contaminant of \rightarrow milk, plasma and the excreta of cows. There is no accumulation in any organ.

3'-Hydroxy T-2 toxin is a metabolite of \rightarrow T-2 toxin and a contaminant of \rightarrow milk, plasma and the excreta of cows. There is no accumulation in any organ.

Hyperemia engorgement of blood

Hyperestrogenism \rightarrow F-2 toxicoses

Hyperplasia an abnormal increase in the number of cells.

Hypocreaceae → Hypocreales

Hypocreales → Ascomycota

Hypothermia an unusually low body temperature

I

Ice cream Manufacturing of naturally contaminated \rightarrow milk will result in \rightarrow aflatoxin M₁ contamination of ice \rightarrow cream because no toxin destruction occurred after 8 month of frozen storage. \rightarrow coconut ice

Icterus → Jaundice

immunosuppressive increased susceptibility to diseases caused by bacteria, viruses and fungi

Incarpina is a product consisting of \rightarrow maize plus cottonseed \rightarrow flour. Incarpina may contain the following \rightarrow mycotoxins: \rightarrow fumonisin B₂ incidence: 1/1, conc.: 140 µg/kg, country: Guatemala

Indian cassia (Cinnamomum tamala (Bush.-Ham.) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/6, conc.: 13 µg/kg, country: India \rightarrow aflatoxin B₂ incidence: 1/6, conc.: 11 µg/kg, country: India \rightarrow aflatoxin G₁ incidence: 1/6, conc.: $8 \mu g / kg$, country: India \rightarrow aflatoxin G₂ incidence: 1/6, conc.: 4 µg/kg, country: India \rightarrow spices

Indian childhood cirrhosis This disease caused vague gastrointestinal symptoms and \rightarrow anorexia. The subsequent hepatomegaly often resulted in \rightarrow icterus, \rightarrow ascites and \rightarrow hepatic coma, mainly in children with a peak incidence at 3 years, in certain areas of India. The detection of

aflatoxin-like fluorescent substances succeeded in the mother's breast milk (\rightarrow Human breast milk), the urine of affected children, parboiled \rightarrow rice and the \rightarrow peanut oil used for frying most foods. However, because no chemical confirmation of the identity of these compounds was carried out, the etiology of this lethal disease is unresolved.

Ingwer may contain the following → mycotoxins: → aflatoxin B₁ incidence: 8/15, conc. range: < 2.5-12.5 µg / kg, country: Canada incidence: 1/12, conc.: $\leq 5 \mu g / kg$, country: Germany incidence: 3/5, conc. range: 1.4-6.5 µg / kg, country: USA → spices

Intracellular mycotoxins like \rightarrow penitrem A, \rightarrow roquefortine C, \rightarrow sterigmatocystin, verrrucosidin are mainly intracellular. \rightarrow Extracellular mycotoxins, \rightarrow mycotoxins

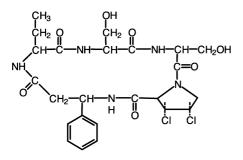
Islanditoxin is composed of L-serine, L- β -phenyl- β -aminopropionic acid, L- α amino-n-butyric acid, and L-dichloroproline in the mole ratio of 2:1:1:1 (\rightarrow mycotoxins). This cyclic, water soluble, colorless chloropeptide was first isolated in 1955 and structurally elucidated in 1959 (see Figure Islanditoxin).

Снемісаl Data Empirical formula: $C_{24}H_{31}O_7N_5Cl_2$, molecular weight: 571

FUNGAL SOURCES Pencillium islandicum Sopp

NATURAL OCCURRENCE It may be a contaminant of "yellow rice". TOXICITY

LD₅₀ (po): 6.55 mg/kg bw mice Clinical signs include respiratory and circulatory disturbances, low body temperature, decrease of muscle and skin tension,



Islanditoxin

enlargement of the liver (significant decline in \rightarrow hepatic glycogen content, concomitant decrease in hepatic glycogen synthetase activity), hemorrhagic chan-

ges (\rightarrow hemorrhage) in the small intestines.

DETECTION TLC

POSSIBLE MYCOTOXICOSIS

In combination with \rightarrow luteoskyrin, islanditoxin should be responsible for the \rightarrow Yellow rice disease

FURTHER COMMENTS

Compared to luteoskyrin it is more toxic. If the chlorine atoms are removed, the toxicity of islanditoxin is significantly reduced.

Isofumigaclavine A, B (Syn.: \rightarrow roquefortine A & B)

J

Jam (no specification) may contain the following \rightarrow mycotoxins: \rightarrow patulin incidence: 10/20, conc. range: 5-50 µg/ kg, country: Italy incidence: 15/35, conc. range: 2-20 µg/ kg, country: Germany \rightarrow fruits

```
\rightarrow aflatoxin B<sub>1</sub>
incidence: 34/144, conc. range: 0.1-14.9
μg/kg, country: Japan
\rightarrow aflatoxin B<sub>2</sub>
incidence: 34/144, conc. range: tr-1.8
μg/kg, country: Japan
\rightarrow aflatoxin G<sub>1</sub>
incidence: 34/144, conc. range: 0.3-0.7
μg/kg, country: Japan
\rightarrow deoxynivalenol
incidence: 2/12, conc. range: 48-496
μg/kg, country: Japan
\rightarrow nivalenol
incidence: 11/12, conc. range: 3-920
μg/kg, country: Japan
\rightarrow zearalenone
incidence: 7/7, conc. range: 10-440
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μg/kg, country: Japan

Jaundice is characterized by very high levels of bilirubin in the blood while bile pigment is deposited in the skin and mucous membrane, causing to a yellow appearance.
Job's-tears is an oriental kind of seed. Job's-tears may contain the following → mycotoxins:

K

Kashin-Beck disease (Syn.: Urov disease) is neither heritable nor congenital. Its etiology is still unknown KBD is endemic to northern China, North-Korea, Russia (Siberia) and was first described along the Urov river in Russia in 1861. Especially in Russia certain climatic conditions, like significant temperature changes during the day, major rainfall during late summer and / or early fall at grain maturing and harvesting in connection with this disease have been reported. More recently KBD (named after the two Russian scientists Kashin and Beck who studied the disease from the 1860s) has also been detected in Taiwan, Japan, Sweden, and Holland. In China about two million people are affected, predominantly peasants in rural areas. The consumption of \rightarrow maize and \rightarrow wheat infected with \rightarrow Fusarium spp. may be responsible (→ Fusarium equiseti (Corda) Sacc. sensu Gordon / → fusarochromanone). Besides the possible involvement of different Fusarium species (F. equiseti, → Fusarium oxysporum Schlecht. emend. Snyd. & Hansen, \rightarrow Fusarium poae (Peck) Wollenw.), selenium deficiency as well as trace metal toxicity have been discussed as possible etiological agents. Pre-adolescent and adolescent children are primarily affected. They show bone and joint deformation, typically in the elbows, knees, and ankles, which leads to impaired mobility; disproportionate dwarfism may also occur. At an early stage, reversibility of the disease is possible if the patients leave the endemic area. The decline of KBD in some areas may be the result of improved hygienic conditions, together with the import of grain from non-endemic regions. Further studies are needed to elucidate the causative agents of this widespread, crippling disease.

Kodua poisoning occurs in India and may be due to the ingestion of kodo \rightarrow millet seeds (*Paspalum* spp.) by cattle and humans that are contaminated with \rightarrow Aspergillus spp. / \rightarrow cyclopiazonic acid. Cattle show symptoms of nervousness, lack of muscular coordination, depression and spasms, death sometimes occurs. The accidental consumption of the contaminated, dehusked grains cooked like \rightarrow rice or used in \rightarrow bread baking caused tremors (\rightarrow tremorgenic mycotoxins), giddiness, and sleepiness.

Kojic acid is a 2-hydroxymethyl-5hydroxy-2- γ -pyrone (\rightarrow mycotoxins) produced by several \rightarrow Aspergillus and \rightarrow Penicillium species (see Figure Kojic acid).

Снемісаl Data Empirical formula: C₆H₆O₄, molecular weight: 142

FUNGAL SOURCES Aspergillus spp. (e.g. \rightarrow Aspergillus candidus Link, \rightarrow Aspergillus flavus Link, \rightarrow Aspergillus oryzae (Ahlburg) Cohn, \rightarrow Aspergillus parasiticus Speare, A. tamarii group, A. wentii group), Penicillium spp. (\rightarrow Penicillium citrinum Thom, P. lanosum, P. rubrum) and Verticillium

NATURAL OCCURRENCE \rightarrow figs, \rightarrow maize

TOXICITY

dahliae.

convulsive (\rightarrow convulsions), \rightarrow mutagenic insecticidal

 LD_{50} (ip): 30 mg/mice

Large amounts are necessary to produce server intoxication or death in animals. Up to now, no natural cases of kojic acid

СН₂ОН

Kojic acid

M. Weidenbörner, *Encyclopedia of Food Mycotoxins* © Springer-Verlag Berlin Heidelberg 2001 toxicosis have appeared in animals or humans.

Detection TLC

Koshk → yoghurt

Kubeba is an Egyptian meat product. Detection of \rightarrow aflatoxins in kubeba results from the use of mycotoxin contaminated \rightarrow spices and/or the incorporation of aflatoxin producers. Kubeba may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/25, conc.: 150 µg/kg, country: Egypt \rightarrow aflatoxin B₂ incidence: 1/25, conc.: 25 µg/kg, country: Egypt \rightarrow meat

Kulen is a Yugoslavian \rightarrow meat speciality and may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A incidence: 27/206* conc. range: 10-460 μ g/kg, country: Yugoslavia, *total of smoked \rightarrow meat

Kwashiorkor is common in tropical and subtropical countries / areas like parts of Brazil, Central America, southern India, Mexico, South Africa, Uganda, and parts of Zaire. The local main staple foods, e.g. \rightarrow maize, \rightarrow rice and / or plantains (high in starch, low in protein), are often contaminated with \rightarrow aflatoxins. Accumulation of these \rightarrow mycotoxins in the body fluids and tissues of very young children suffering from kwashiorkor has been reported. The symptoms shown by these children may in part also be due to protein malnutrition. The disease is characterized by several clinical signs like hypoalbuminaemia, \rightarrow edema, immunosuppression (\rightarrow immunosuppressive), and fatty liver. These symptons are also caused by aflatoxins in experimental animals (guinea pigs). Although an association between aflatoxin and kwashiorkor has been established, conclusive evidence is still lacking.

L

125

Lemons (pickled in salt) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 19/40*, Ø conc.: 195 μg/kg, country: India incidence: 3/18**, conc. range: 20-60 μg / kg, country: Germany, **moldy \rightarrow aflatoxin B₂ incidence: 19/40*, Ø conc.: 42 μg/kg, country: India \rightarrow aflatoxin G₁ incidence: $19/40^*$, Ø conc.: $110 \mu g/kg$, country: India \rightarrow aflatoxin G₂ incidence: 19/40*, Ø conc.: 25 μg / kg, country: India *stored in polythene bags \rightarrow fruits

Lentils may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/6, conc.: 3.1 µg/kg, country: Egypt incidence: 1/4, conc.: 72 μ g/kg, country: Egypt incidence: 1/20, conc.: 8-10 µg/kg, country: Italy incidence: 1/2*, conc.: 1.8 µg/kg, country: Syria, *ground \rightarrow aflatoxins incidence: 1/20, conc.: 20 µg/kg, country: Egypt \rightarrow ochratoxin A incidence: 2/10, conc. range: 0.1-0.19 μg/kg, country: Germany \rightarrow beans, \rightarrow cabbage, \rightarrow cowpeas, \rightarrow pigeon peas, \rightarrow peas, \rightarrow soybeans, \rightarrow vegetables

Leucopin (Syn.: \rightarrow patulin)

Leukocytosis transient increase in the amount of the white blood cells in the blood.

Lewia → Pleosporaceae

Libritos may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 4/4, conc. range: 210-1023 $\mu g / kg$, \emptyset conc.: 581 $\mu g / kg$, country: Argentina \rightarrow wheat products

Lima beans → Beans, lima

Lineseed oil may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/10, conc.: 1.2 µg/kg, country: Germany

Lineseeds may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1*/6, conc.: 1.1 µg / kg, country: Germany \rightarrow aflatoxin G₁ incidence: 1*/6, conc.: 0.9 µg / kg, country: Germany *soaked for 36 h

Lingonberries may contain the following \rightarrow mycotoxins: \rightarrow patulin incidence: 1/2, conc.: 265 µg/kg, country: Sweden \rightarrow fruits

Losses Worldwide losses in the export market due to mycotoxin contamination in only five crops (\rightarrow barley, cottonseed, \rightarrow maize, \rightarrow peanuts, and \rightarrow rice) were

Losses

estimated for 1985 at about 1.5 billion dollars. These losses did not consider human costs due to mycotoxin (\rightarrow mycotoxins) contamination, losses in pigs/ abortions, dairy \rightarrow cattle/ \rightarrow milk production and \rightarrow poultry/egg production, losses in domestic animals due to reduced weight gain because of mycotoxin-contaminated animal feeds, losses due to lower prices for lower quality \rightarrow grains etc. In view of all these costs, a major research effort in mycotoxin prevention and control is necessary.

Low water activity foods Direct mycotoxin (\rightarrow mycotoxins) contamination in foods characterized by low water activities is most unlikely. The minimum \rightarrow a_w for mycotoxin production is a_w 0.80 (*Aspergillus ochraceus l* \rightarrow penicillic acid).

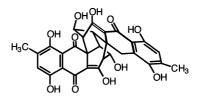
Luteoskyrin (Syn.: flavomycelin) is a 2,2',4,4',5,5',8,8'-octahydroxy-2,2',3,3'tetrahydro-7,7'-dimethyl-1,1'-bianthraquinone (\rightarrow mycotoxins). This yellow anthraquinone-like pigment is produced by \rightarrow Penicillium islandicum Sopp which was first isolated in 1912 from skyr, an Iceland kind of \rightarrow yogurt (see Figure Luteoskyrin). Due to pyrolysis, this bisanthraquinone decomposes into cateniarin and \rightarrow islanditoxin in a molar ratio of 1:1.

CHEMICAL DATA Empirical formula: $C_{30}H_{22}O_{12}$, molecular weight: 574

Fungal Sources P. islandicum

NATURAL OCCURRENCE

P. islandicum is promoted in tropical and subtropical climates especially in the \rightarrow rice-growing areas of Asia and Africa where high temperatures and humid con-



Luteoskyrin

ditions are common. This yellow rice may be contaminated with luteoskyrin. A high incidences of diseases such as liver cirrhosis and carcinoma occurs in such areas.

P. islandicum grows on contaminated rice, \rightarrow maize, and other \rightarrow cereals. Because of the lipophilic nature of luteoskyrin, \rightarrow oil processed from contaminated rice husks might be a high risk foodstuff. In Europe animal feed is mainly affected by luteoskyrin contamination, while food contamination is rare.

TOXICITY

hepatotoxic: the liver shows yellow discoloration, centrilobular necrosis, fatty degeneration, liver tumors (mice); carcinogenic

 LD_{50} (po): 221 mg/kg bw mice, significant toxicological variation depending on the route of administration. Chemically luteoskyrin is very similar to \rightarrow rugulosin which caused the same clinical signs.

Detection TLC

Possible Mycotoxicosis \rightarrow Yellow rice disease

FURTHER COMMENTS

 \rightarrow Apples and \rightarrow grape juice are very good substrates for luteoskyrin production.

Lymphocytosis exessive increase in the number of lymph cells

Μ

Macrofusin (Syn.: fumonisin $B_1 \rightarrow$ fumonisins)

Maize Among cereal \rightarrow grains maize as a staple food is considered as a high risk crop for mycotoxin production. The toxin levels found are generally higher than those of other small-grain \rightarrow cereals. \rightarrow Fusarium graminearum Schwabe, → Fusarium proliferatum (Matsushima) Nirenberg and \rightarrow Fusarium moniliforme Sheldon are extremely common on maize ears and their \rightarrow mycotoxins represent a serious health hazard to man and domestic animals. In addition, F. subglutinans is also very common on maize. F. graminearum and F. moniliforme colonize maize seeds on the cob or in the ear. The developing corn is invaded by F. graminearum at the silking stage, especially in periods of heavy rainfall. Wet or insufficiently dried stored kernels promote mycelial growth, while low temperatures $(\leq 15 \ ^{\circ}C)$ are essential for \rightarrow zearalenone production. Ears stored in cribs are most frequently affected by zearalenone contamination.

F. moniliforme is primarily an internally seed-borne fungus but it is also soilborne and survives in plant residues. F. moniliforme requires a minimum moisture content of 18.4% (maize) for vegetative growth. The fungus invades the seed through the pedicle to colonize the internal section of the kernel, including the embryo. In consequence, fumonisin contamination may occur. Since fumonisin production is favored by high seed moisture contents, maximum formation probably occurs during harvest or before drying and storage. In the field \rightarrow fumonisin B₁ formation in maize succeeded FB₂ and FB₃ production while most of the fumonisin is located in the pericarp layer of maize kernels.

In Argentinian maize a good agreement between fungal contamination (F. moniliforme, F. proliferatum) at the medium and late stages of maturity and fumonisin contamination was found. Conventional grading of corn for human consumption is not effective to lower fumonisin levels because "first-grade" maize may also contain high fumonisin concentrations. In general, processing does not remove or inactivate fumonisin B_1 in maize and maize products. So, these foodstuffs are regarded as the main sources for human and animal FB1 intake. However, it could be shown that \rightarrow milling caused the accumulation of \rightarrow fumonisins in maize screenings and \rightarrow bran. Therefore, increasing the level of refinement of corn meal may cause a decrease in fumonisin levels. \rightarrow Fusarium mycotoxins occur in different parts of the maize plant, e.g. stalk, leaf axis, peduncle, rachis and kernels but they are unevenly distributed. It seems that the kernels are one of the least contaminated parts of the plant. Therefore, mycotoxin contamination of maize grain may be minimized by prompt, undelayed, harvesting. In addition, maize may be prone to field

In addition, maize may be prone to field infection with \rightarrow Aspergillus flavus Link and \rightarrow Aspergillus parasiticus Speare resulting in significant aflatoxin contamination before harvesting. It is suggested that insect damage and inoculum spread is the major cause of maize infections in the USA. Colonization of the base of the kernels is due to the inoculum present on the infected maize silks. The high carbohydrate and low nitrogen content of the seeds favor aflatoxin production but regional differences in contamination (higher levels: e.g. southeastern States USA, western and northern regions India) do occur.

The aflatoxigenic fungi are primarily surface-borne but occasionally internally seed-borne. The maturing kernels are

Maize

highly susceptible during the late milk and dough stage, but infection is also possible at a later stage before harvesting. Integrity of the kernel in general prevents aflatoxin contamination but direct colonization of intact kernels through the silk scars by the fungus without harming the process of fertilization and growth of the developing embryo is possible. However, breaks in the pericarp tissue due to insects (European corn borer = Ostrinia nubialis, corn earworm = Heliothis zea and the rice weevil = Sitophilus oryzae) which also transmit spores of A. flavus, plant stress (e.g. drought, low soil fertility, weed competition) and / or mechanical damage major contribute to infection of maize kernels.

During warm weather at harvest there is a high risk of aflatoxin contamination of maize. Temperatures of 24 °C or a moisture content of 17.5% are necessary for aflatoxin B₁ production in stored maize. \rightarrow Aflatoxins as well as zearalenone formation occurs during the development of "hot spots".

Single kernels or pieces of kernels of a maize sample may contain very high levels of \rightarrow aflatoxin B₁ (88,500-101,000 μ g / kg). In naturally contaminated maize \rightarrow aflatoxin G₁ is always found to a lower extent than AFB₁ and never occurs in the absence of AFB₁. Aflatoxins may also be present in all corn fractions including sound kernels, damaged and discolored kernels, fluorescing kernels, kernels with visible fluorescence beneath the seed coat, broken corn-foreign material. Aflatoxin-containing particles can be removed to a different degree by cleaning processes, e.g. blowers and sieves, because the contaminated fragments shatter easily. Electronic sorting devices are also helpful.

Wet- or dry-milling of maize will result in the accumulation of aflatoxins mainly in the feed fractions. This phenomenon is even more pronounced in the wet milling process.

Besides the simultaneous contamination of maize with different *Fusarium* mycotoxins co-contamination with aflatoxin and fumonisin B_1 has been detected. Under favorable conditions the growth and mycotoxin production of aflatoxigenic fungi as well as *F. moniliforme* and / or *F. proliferatum* is possible but negative relationships between these fungi and mycotoxin production have also been reported. It is assumed that fumonisins are more evenly distributed in maize kernels compared to the distribution of aflatoxins.

The excellent mycotoxin-promoting nature of maize is confirmed by the fact that maize kernels contained nearly 10 times more moniliformin than *Fusarium* damaged wheat kernels (\emptyset ca. 16,000 µg/kg).

- Maize may contain the following \rightarrow mycotoxins:
- \rightarrow 3-acetyldeoxynivalenol

incidence: 1/1, conc.: 100 μg/kg, country: Austria

incidence: 5/24, conc. range: 30-185 μ g/kg, Ø conc.: 113 μ g/kg, country:

China

incidence: 12/36*, conc. range: 20-1500 μg / kg, Ø conc.: 200 μg / kg, country: Korea, *moldy

incidence: 2/35*, conc. range: 50-200 μ g / kg, Ø conc.: 200 μ g / kg, country: Korea, *healthy

incidence: 1/9*, conc.: 300 μ g/kg, country: Poland, *healthy and damaged kernels

→ 15-acetyldeoxynivalenol

incidence: 5/24, conc. range: 160-1435 μ g/kg, Ø conc.: 495 μ g/kg, country: China

incidence: $30/36^*$, conc. range: $20-4600 \mu g/kg$, \emptyset conc.: $900 \mu g/kg$, country: Korea, *moldy

incidence: 6/35*, conc. range: 2-100 μ g / kg, Ø conc.: 40 μ g / kg, country: Korea, *healthy incidence: 4/9*, conc. range: 2800-7700 μ g / kg, Ø conc.: 4725 μ g / kg, country: Poland, *healthy and damaged kernels incidence: 7/20*, conc. range: 900-7900 μ g/kg, Ø conc.: 1800 μ g/kg, country: USA *moldy \rightarrow 4-acetylnivalenol incidence: 14/36*, conc. range: 2-2200 μ g / kg, Ø conc.: 400 μ g / kg, country: Korea, *moldy incidence: 3/35*, conc. range: 4-30 μ g / kg, Ø conc.: 10 μ g / kg, country: Korea, *healthy aflatoxicol I incidence: 2/2, conc. range: 12.9-25.4 μ g / kg, Ø conc.: 19.15 μ g / kg, country: Thailand aflatoxicol II incidence: 2/2, conc. range: 7.9-15.7 μ g/kg, Ø conc.: 11.8 μ g/kg, country: Thailand \rightarrow aflatoxin B₁ incidence: 445/2271, conc. range: \leq 560 μ g / kg, Ø conc.: 11.6 μ g / kg, country: Argentina incidence: 5/150, conc. range: 10-50 μ g/kg, Ø conc.: 24 μ g/kg, country: Argentina incidence: 3/174, conc. range: 1-3 µg/kg, country: Australia incidence: 1/1, conc.: 131 µg/kg, country: Burma incidence: 30/36, conc. range: 0.54-76.32 μ g/kg, Ø conc.: 15.8 μ g/kg, country: Costa Rica incidence: 1/6, conc.: 2.6 µg/kg, country: Egypt incidence: 2/75, conc. range: 10 μ g/kg, Ø conc.: 10 µg/kg, country: France incidence: 1*/3, conc.: 25 µg/kg, country: Germany, *moldy incidence: 975/2074, conc. range: > 5-666 μg/kg, country: India

incidence: 6/6, conc. range: tr-15,600 μg/kg, country: India incidence: 11/16, conc. range: 4-428 μ g / kg, Ø conc.: 102 μ g / kg, country: Indonesia incidence: 10/12, conc. range: 1-3300 μ g/kg, Ø conc.: 352 μ g/kg, country: Indonesia incidence: 50/111, conc. range: 0.02-1.2 μ g / kg, country: Italy incidence: 2/161, conc.: 0.1 µg/kg, country: Japan incidence: 3/3, conc. range: 8.8-37.5 μ g / kg, Ø conc.: 21.8 μ g / kg, country: Nepal incidence: 44/50, conc. range: 1-430 μ g / kg, Ø conc.: 49 μ g / kg, country: Philippines incidence: 39/155, conc. range: < 5-1500 µg/kg, country: South Africa incidence: 2/2, conc. range: 131-340 μ g / kg, Ø conc.: 236 μ g / kg, country: Thailand incidence: 158/162, conc. range: 500-1200 μg/kg, country: Thailand incidence: 17/27, conc. range: 1-606 μ g/kg, Ø conc.: 63 μ g/kg, country: Thailand incidence: 22*/62, Ø conc.: 400 µg/kg, country: Thailand, *total: Ø conc.: 2730 μ g / kg AFB₁, AFB₂, AFG₁, AFG₂ incidence: 3/38, conc. range: 48-62 μg/kg, country: Tunisia incidence: 27/167, conc. range: 2-73.9 μg/kg, country: Turkey incidence: 24/29, conc. range: $< 5 \mu g / kg$ (21 samples), 6-10 µg/kg (2 sa), 11-15 μ g / kg (1 sa), country: UK incidence: 16/567, conc. range: 20-350 μg/kg, country: USA incidence: 6/283, conc. range: 6-25 μ g / kg, Ø conc.: 15 μ g / kg, country: USA incidence: 21/60, conc. range: 4-308 μ g / kg, Ø conc.: 58.6 μ g / kg, country: USA incidence: 8/293, conc. range: < 6-25 μg/kg, country: USA

Maize

incidence: 27/28, conc. range: 0-321 μ g / kg, Ø conc.: 73 μ g / kg, country: USA incidence: 11*/34, conc. range: 0.7-47 μ g / kg, country: USA, *single damaged kernels contained 88,500-101,000 µg AFB_1/kg incidence: 25/353, conc. range: 3-19 μ g / kg, Ø conc.: 10.4 μ g / kg, country: USA incidence: 10/918, conc. range: 3-19 $\mu g / kg$, Ø conc.: 9 $\mu g / kg$, country: USA \rightarrow aflatoxin B₂ incidence: 92/2271, conc. range: 130 μ g/kg, Ø conc.: 28.15 μ g/kg, country: Argentina incidence: 1/174, conc.: 50 µg/kg, country: Australia incidence: 1/1, conc.: 18 µg/kg, country: Burma incidence: 18/36, conc. range.: 0.16-5.82 μ g / kg, Ø conc.: 1.9 μ g / kg, country: Costa Rica incidence: 1/6, conc.: 3.7 µg/kg, country: Egypt incidence: 9/16, conc. range: 1-160 $\mu g \, / \, kg, \, {\cal O} \, \, conc.:$ 9 $\mu g \, / \, kg, \, country: Indo$ nesia incidence: 8/12, conc. range: 1-680 μ g/kg, Ø conc.: 90 μ g/kg, country: Indonesia incidence: 34/50, conc. range: 1-78 μ g/kg, Ø conc.: 14 μ g/kg, country: Indonesia incidence: 3/3, conc. range: 2.3-5 µg/kg, \emptyset conc.: 4.1 µg/kg, country: Nepal incidence: 2/2, conc. range: 17-47 µg/kg, Ø conc.: 32 μ g/kg, country: Thailand incidence: 135/162, conc. range: 49-260 μg/kg, country: Thailand incidence: 11/27, conc. range: 1-73 μ g/kg, Ø conc.: 14 μ g/kg, country: Thailand incidence: 8/167, conc. range: 1.5-6 μg/kg, country: Turkey incidence: 4/567, conc. range: 52-129 μ g / kg, country: USA incidence: 15/60, conc. range: tr-40 μg/kg, country: USA

 \rightarrow aflatoxin G₁ incidence: 2/150, conc. range: 10-25 μ g / kg, Ø conc.: 17.5 μ g / kg, country: Argentina incidence: 1/174, conc.: 2 µg/kg, country: Australia incidence: 1/3, conc.: 57.6 µg/kg, country: Nepal incidence: 2/50, conc. range: 40-78 μ g / kg, Ø conc.: 59 μ g / kg, country: Philippines incidence: 17/162, conc. range: 50-250 μ g / kg, country: Thailand incidence: 3/27, conc. range: 2-7 µg/kg, \emptyset conc.: 5 µg/kg, country: Thailand incidence: 3/38, conc. range: 8-22 µg/kg, country: Tunisia incidence: 3/167, conc. range: 2-5.4 μg / kg, country: Turkey incidence: 2/283, conc. range: tr-12 μ g / kg, country: USA incidence: 5/60, conc. range: tr-10 µg/kg, country: USA incidence: 3/353, conc. range: 3-8 µg/kg, \emptyset conc.: 5.7 µg/kg, country: USA incidence: 3/918, conc. range: tr-3 µg/kg, country: USA \rightarrow aflatoxin G₂ incidence: 5/16, conc. range: tr-8 µg/kg, country: Indonesia incidence: 2/50, conc. range: 3-33 µg/kg, \emptyset conc.: 18 µg/kg, country: Indonesia incidence: 1/3, conc.: 9.7 µg/kg, country: Nepal incidence: 2/162, conc. range: 49-110 μg/kg, country: Thailand incidence: 2/167, conc. range: 2-3 µg/kg, country: Turkey incidence: 2/60, conc. range: tr-1 µg/kg, country: USA aflatoxin (no specification) incidence: 1/71*, conc.: 2-20 μg/kg, country: Uruguay, *and by-products \rightarrow aflatoxins (no specification) incidence: 1*/36, conc.: < 25 µg/kg, country: Canada, *AFB₁, AFB₂, AFG₁, AFG_2

incidence: 9*/10, conc. range: 2-35 μ g / kg, Ø conc.: 9.7 μ g / kg, country: Gambia, *AFB₁, AFB₂, AFG₁, AFG₂ incidence: 304/364, conc. range: nc, country: Germany incidence: 7/22, conc. range: 12-160 µg AFB₁ / kg, 25-90 μg AFB₂ / kg, 10-95 μg AFG₁ / kg, 65 μ g AFG₂ / kg, country: India incidence: 2/8, conc. range: nc, country: Hong Kong incidence: 2/52, conc. range: nc, country: Mocambique incidence: 22*/49, conc. range: 1-100 μ g / kg (13 samples), 100-1000 μ g / kg (9 sa), country: Uganda, * 19 samples contained AFB₁, 11 AFB₂, 14 AFG₁, 4 AFG₂ incidence: 39/45, conc. range: 1-2300 μ g / kg, Ø conc. 252 μ g / kg, country: USA incidence: 717/4651, conc. range: 20-100 μg/kg, country: USA incidence: 40/1594, conc. range: 3-37 μ g / kg, Ø conc.: 9 μ g / kg, country: USA incidence: 21/60, conc. range: 6-348 μ g / kg, Ø conc.: 66 μ g / kg, country: USA incidence: 235/2866, conc. range: 15 µg/ kg, country: USA incidence: 281/743, Ø conc.: 135 μg/kg, country: USA incidence: 46/123*, Ø conc.: 130 μg/kg, country: USA incidence: 49/101*, Ø conc.: 187 μg/kg, country: USA incidence: 36/99*, Ø conc.: 58 μg/kg, country: USA incidence: 33/114*, Ø conc.: 118 μg/kg, country: USA incidence: 81/99*, Ø conc.: 167 μg/kg, country: USA incidence: 11/90*, \emptyset conc.: 110 µg/kg, country: USA incidence: 24/117*, Ø conc.: 176 μg/kg, country: USA *dent maize incidence: 49/109, conc. range: ≤ 123 μ g / kg, Ø conc.: 30 μ g / kg, country: USA incidence: 12/28, conc. range: \leq 98 μ g / kg, Ø conc.: 20 μ g / kg, country: USA

incidence: 63/197, conc. range: \leq 1019 μ g / kg, Ø conc.: 77 μ g / kg, country: USA incidence: 57/315, conc. range: tr-845 μg/kg, country: USA incidence: 27/28, conc. range: 0-321 μ g / kg, Ø conc.: 73 μ g / kg, country: USA incidence: 218/1669, conc. range: 20-99 μ g / kg (167 samples), 100 μ g / kg (51 sa), country: USA → beauvericin incidence: 6/22*, conc. range: tr-520,000 μ g / kg, Ø conc.: 102,833 μ g / kg, country: Italy, *visibly infected \rightarrow citrinin incidence: 1/1, conc.: 212 µg/kg, country: Burma incidence: 2/2, conc. range: 174-1390 μ g / kg, Ø conc.: 782 μ g / kg, country: Thailand incidence: 1/1, conc.: 450 µg/kg, country: UK \rightarrow cyclopiazonic acid incidence: 23/45, conc. range: < 25-2800 μ g / kg, Ø conc.: 467 μ g / kg, country: USA → deoxynivalenol incidence: 2/20, Ø conc.: 111 µg/kg, country: Argentina incidence: 33/100, conc. range: tr-200 μg/kg, country: Argentina incidence: 1/1*, conc.: 1450 µg/kg, country: Argentina, *flint maize incidence: 14/58, conc. range: 200-400 μg/kg, country: Argentina incidence: 77/78, conc. range: ≤ 6200 μ g / kg, Ø conc.: 790 μ g / kg, country: Austria incidence: 1/1, conc.: 90,000 µg/kg, country: Austria incidence: 46/51, conc. range: 40-3700 μ g/kg, Ø conc.: 730 μ g/kg, country: Austria incidence: 3/6*, conc. range: 550-50,500 μ g / kg, Ø conc.: 17,400 μ g / kg, country: Austria, *visibly moldy (Fusarium spp.) incidence: 3/3, conc. range: 1300-7900 μg/kg, country: Austria

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incidence: 77/78, conc. range: \leq 6200 μ g / kg, Ø conc.: 780 μ g / kg, country: Austria incidence: 1/1, conc.: 960 μ g/kg, country: Canada incidence: 2/2*, conc.: 130-700 μg/kg, Ø conc.: 415 µg/kg, country: Canada, *No. 2 incidence: 243/283, conc. range: 20-4090 μ g/kg, Ø conc.: 610 μ g/kg, country: Canada incidence: 28/28, conc. range: \leq 4500 μ g / kg, Ø conc.: 1960 μ g / kg, country: China incidence: 24/24, conc. range: 360-12,670 μ g / kg, Ø conc.: 5376 μ g / kg, country: China incidence: 4/4, conc. range: 20-100 μg/kg, country: France incidence: 1/1, conc.: 40 µg/kg, country: France incidence: 3/3, conc. range: 20-60 µg/kg, \emptyset conc.: 40 µg/kg, country: France incidence: 9/23, conc. range: 10-1800 μ g / kg, Ø conc.: 900 μ g / kg, country: Germany incidence: 10/35, conc. range: 30-2000 μg/kg, country: Germany incidence: 2/11, conc. range: 200-1300 μg/kg, country: Germany incidence: 2/4*, conc. range: 280-640 μ g / kg, Ø conc.: 460 μ g / kg, country: Germany, *organic produce incidence: 2/16, conc. range: 21-32 μ g/kg, Ø conc.: 27 μ g/kg, country: Indonesia incidence: 2/3, Ø conc.: 402 µg/kg, country: Italy incidence: nc/6, conc. range: 20-670 μ g / kg, country: Italy incidence: 1/1, conc.: 67,000 µg/kg, country: Italy incidence: 1/1*, conc.: 20,000 µg/kg, country: Italy, *visible moldy (Fusarium spp.) incidence: 2/2, conc. range: 101-500 μ g / kg, country: Italy

incidence: 14/15, conc. range: 22-442 μ g/kg, Ø conc.: 145 μ g/kg, country: Korea incidence: 34/36*, conc. range: 6-15,200 μ g/kg, Ø conc.: 4000 μ g/kg, country: Korea, *moldy incidence: 8/35*, conc. range: 10-100 μ g/kg, Ø conc.: 40 μ g/kg, country: Korea, *healthy incidence: 1/3, conc.: $352 \mu g/kg$, country: Korea incidence: 3/9, Ø conc: 541 µg/kg, country: Nepal incidence: 11/20, conc. range: \leq 300 μ g/kg, Ø conc.: 100 μ g/kg, country: New Zealand incidence: 73/91, conc. range: 3500 µg/kg, country: New Zealand incidence: 8/9*, conc. range: 1400-132,000 μ g / kg, Ø conc.: 49,350 μ g / kg, country: Poland, *healthy and damaged kernels incidence: 14/36, conc. range: tr-820 µg/kg, country: South Africa incidence: 1/5, conc.: 140 µg/kg, country: South Africa incidence: 24/24, conc. range: 50-12,100 μ g / kg, Ø conc.: 2900 μ g / kg*, 300 µg/kg**, country: South Africa, *lowprevalence EC area, **high-prevalence EC area incidence: 2*/2, conc. range: 420-2500 μ g/kg, Ø conc.: 1460 μ g/kg, country: South Africa, *moldy incidence: 7/10, conc. range: 20-100 µg/kg, country: South Africa incidence: 50/50, conc. range: 7-7400 μg/kg, country: South Africa incidence: 43/72, conc. range: 10-15,800 µg/kg, country: South Africa incidence: 2/2*, conc. range: 120-180 μ g/kg, Ø conc.: 150 μ g/kg, country: USA, *yellow maize No. 3 incidence: 7/100, conc. range: 95-312 μg/kg, country: USA incidence: 24/52, conc. range: 500-10,000 μ g / kg, Ø conc.: 5000 μ g / kg, country: USA

incidence: 93/198, conc. range: ≤ 2470 μ g / kg, Ø conc.: 400 μ g / kg, country: USA incidence: 44/52, conc. range: 500-10,700 μg/kg, country: USA incidence: 17/20*, conc. range: 400-65,800 μ g/kg, Ø conc.: 19,700 μ g/kg, country: USA, *moldy incidence: 33/33, conc. range: 20-100 μg/kg (2 samples), 101-500 μg/kg (17 sa), > 500 μ g/kg (14 sa), country: USA incidence: 1/1*, conc.: 100 µg/kg, country: USA, *dent maize No. 2 incidence: 1/1*, conc.: 550 µg/kg, country: USA, *waxy maize incidence: 19/19*, conc. range: 69,960-722,450 μg / kg, Ø conc.: 445,790 μg / kg, country: USA, *moldy, tip section of sweet maize ears incidence: 1/12, \emptyset conc.: 6 μ g/kg, country: Yemen \rightarrow diacetoxyscirpenol incidence: 2/100, conc. range: 400-450 μ g / kg, Ø conc.: 425 μ g / kg, country: Argentina incidence: 1/6*, conc.: 400 µg/kg, country: Austria, *Fusarium infected incidence: 1/77, conc.: 31,500 µg/kg, country: Germany incidence: 6/11, conc. range: 500-2100 μg/kg, country: Germany incidence: 1*/nc, conc.: 14,000 µg/kg, country: India, *moldy incidence: 5/100, conc. range: nc, country: Italy incidence: 6/20, conc. range: \leq 900 μ g / kg, Ø conc.: 350 μ g / kg, country: New Zealand incidence: 8/100, conc. range: nc, country: Yugoslavia \rightarrow fumonisin B₁ incidence: 1/1, conc.: 900 µg/kg, country: Argentina incidence: 17/17*, conc. range: 1110-6695 μ g / kg, Ø conc.: 2877 μ g / kg, country: Argentina, *field-trial corn

incidence: nc/547*, conc. range: \leq 4330 μ g/kg, Ø conc.: 290 μ g/kg, country: Argentina, *export corn for South Africa incidence: 47/47*, conc. range: 50-720 μ g/kg, Ø conc.: 300 μ g/kg, country: Argentina, *export corn for South Africa incidence: 8/8, conc. range: 85-8791 μ g/kg, Ø conc.: 2131 μ g/kg, country: Argentina incidence: 1/1*, conc.: 250 µg/kg, country: Bahrain, *imported from The Netherlands incidence: 9/11*, conc. range: 20-2630 μ g / kg, Ø conc.: 506 μ g / kg, country: Benin, *corn genotypes incidence: 2/2, conc. range: 165-350 μ g/kg, Ø conc.: 258 μ g/kg, country: Botswana incidence: 48/48, conc. range: 600-18,520 μ g / kg, Ø conc.: 5080 μ g / kg, country: Brazil incidence: 6/6, conc. range: 12,200-75,200 μg/kg, country: Burundi incidence: 1/3*, conc.: 120 µg/kg, country: Canada, *fresh maize incidence: 16/48, conc. range: 160-2300 μ g / kg, Ø conc.: 760 μ g / kg, country: Canada incidence: 2/5, conc. range: 5300-8400 μ g / kg, Ø conc.: 6800 μ g / kg, country: China incidence: 16/19*, conc. range: 18,000-155,000 μg/kg, Ø conc.: 74,000 μg/kg, country: China, *moldy corn incidence: 15/15*, conc. range: 20,000-60,000 μg/kg, Ø conc.: 35,300 μg/kg, country: China, *fine corn incidence: 13/27*, conc. range: 186-2964 μ g/kg, Ø conc.: 872 μ g/kg, country: China, *high-EC area incidence: 5/20*, conc. range: 197-1732 μ g / kg, Ø conc.: 890 μ g / kg, country: China, *low-EC area incidence: 7/7, conc. range: 365-3276 μ g / kg, Ø conc.: 1428 μ g / kg, country: China

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incidence: 8/8, conc. range: 1700-4780 μ g / kg, Ø conc.: 2803 μ g / kg, country: Costa Rica incidence: 11/19*, conc. range: 10-60 μ g/kg, Ø conc.: 19.1 μ g/kg, country: Croatia, *corn genotypes incidence: 25/25*, conc. range: tr (< 25 µg/kg) -3350 μg/kg, Ø conc.: 868 μg/kg, country: France, *imported from The Netherlands incidence: 3/3*, conc. range: 100-560 μ g / kg, Ø conc.: 277 μ g / kg, country: Greece, *imported from The Netherlands incidence: 16/16, conc. range: 51-2440 μ g / kg, Ø conc.: 788 μ g / kg, country: Indonesia incidence: 7/12, conc. range: 226-1780 μ g / kg, Ø conc.: 843 μ g / kg, country: Indonesia incidence: 26/26*, conc. range: 10-2330 μ g/kg, Ø conc.: 382 μ g/kg, country: Italy, *corn genotypes incidence: 7/7, conc. range: 100-5310 μ g / kg, Ø conc.: 2807 μ g / kg, country: Italy incidence: 20/22*, conc. range: tr-300,000 μ g / kg, Ø conc.: 74,500 μ g / kg, country: Italy, *visibly infected incidence: 1/1, conc.: 130 µg/kg, country: Kenya incidence: 93/197, conc. range: 110-12,000 μg / kg, Ø conc.: 670 μg / kg, country: Kenya incidence: 33/36*, conc. range: 100-168,800 μg/kg, Ø conc.: 23,200 μg/kg, country: Korea, *moldy incidence: 10/35*, conc. range: 90-12,500 μ g / kg, Ø conc.: 3200 μ g / kg, country: Korea, *healthy incidence: 7/8, conc. range: 20-115 μ g/kg, Ø conc.: 67.1 μ g/kg, country: Malawi incidence: 3/3, conc. range: 240-295 μ g / kg, Ø conc.: 260 μ g / kg, country: Mozambique incidence: 12/24, conc. range: 50-4600 μ g/kg, Ø conc.: 600 μ g/kg, country: Nepal

incidence: 26/50, conc. range: 57-1820 μ g / kg, Ø conc.: 419 μ g / kg, country: Philippines incidence: 2/7*, conc. range: 10-20 μ g/kg, Ø conc.: 15 μ g/kg, country: Poland, *corn genotypes incidence: 9/9*, conc. range: 90-2300 μ g/kg, Ø conc.: 1031 μ g/kg, country: Portugal, *corn genotypes incidence: 3/6*, conc. range: 10-20 μ g/kg, Ø conc.: 13.3 μ g/kg, country: Romania, *corn genotypes incidence: $2/12^*$, conc. range: $\leq 550 \ \mu g/$ kg, \emptyset conc.: 375 µg/kg, country: South Africa, *good corn, low-EC area incidence: 12/12*, conc. range: 50-7900 μ g / kg, Ø conc.: 1600 μ g / kg, country: South Africa, *good corn, high-EC area incidence: 11/11*, conc. range: 450-18,900 μ g/kg, Ø conc.: 6520 μ g/kg, country: South Africa, *moldy corn intended for beer brewing or animal feed, low-EC area incidence: 12/12*, conc. range: 3450-46,900 μg/kg, Ø conc.: 23,900 μg/kg, country: South Africa, *moldy corn intended for beer brewing or animal feed, high-EC area incidence: 5/6*, conc. range: 210-5380 μ g/kg, Ø conc.: 1840 μ g/kg, country: South Africa, *good corn, high-EC area incidence: $6/8^*$, conc. range: ≤ 3310 μ g/kg, Ø conc.: 667 μ g/kg, country: South Africa, *good corn, low-EC area incidence: 7/7*, conc. range: 110-11,340 μ g / kg, Ø conc.: 4050 μ g / kg, country: South Africa, *moldy corn, low-EC area incidence: 6/6*, conc. range: 3020-117,520 μg / kg, Ø conc.: 53,740 μg / kg, country: South Africa, *moldy corn, high-EC area incidence: 1/1, conc.: 600 μ g/kg, country: South Africa incidence: 50/68**, conc. range: < 50-5420 μ g/kg, Ø conc.: 570 μ g/kg (all samples), country: South Africa incidence: 55/66**, conc. range: < 20-5030 μ g/kg, Ø conc.: 380 μ g/kg (all samples), country: South Africa

incidence: nc/77**, conc. range: \leq 3050 μ g / kg, Ø conc.: 320 μ g / kg (all samples), country: South Africa incidence: nc/71**, conc. range: \leq 1810 μ g / kg, Ø conc.: 340 μ g / kg (all samples), country: South Africa incidence: nc/113**, conc. range: \leq 5640 μ g / kg, Ø conc.: 320 μ g / kg (all samples), country: South Africa **white corn incidence: 31/53***, conc. range: < 50-1120 μg / kg, Ø conc.: 180 μg / kg (all samples), country: South Africa incidence: 50/62***, conc. range: < 20-1060 μ g/kg, Ø conc.: 180 μ g/kg (all samples), country: South Africa incidence: nc/82***, conc. range: ≤ 1840 μ g/kg, Ø conc.: 190 μ g/kg (all samples), country: South Africa incidence: nc/76***, conc. range: \leq 740 μ g / kg, Ø conc.: 170 μ g / kg (all samples), country: South Africa incidence: nc/117***, conc. range: \leq 11,700 μ g / kg, Ø conc.: 680 μ g / kg (all samples), country: South Africa ***yellow corn incidence: 24/68*, conc. range: < 50-865 μ g/kg, Ø conc.: 280 μ g/kg, country: South Africa, *export corn for Taiwan incidence: 3/3, conc. range: 400-4440 μ g/kg, Ø conc.: 2447 μ g/kg, country: South Africa incidence: 8/9, conc. range: 25-165 μ g/kg, Ø conc.: 79.4 μ g/kg, country: Tanzania incidence: 16/18, conc. range: 63-18,800 μ g / kg, Ø conc.: 1790 μ g / kg, country: Thailand incidence: 19/27, conc. range: 63-18,800 μ g / kg, Ø conc.: 1580 μ g / kg, country: Thailand incidence: 9/19*, conc. range: 8-380 μ g / kg, Ø conc.: 209 μ g / kg, country: The Netherlands, *intended for bread production incidence: 2/10*, conc. range: 8-110 μg/kg, country: The Netherlands, *intended for popcorn production

incidence: 1/1, conc.: 605 μ g/kg, country: Uganda incidence: 7/7, conc. range: 105-1915 μ g / kg, Ø conc.: 635 μ g / kg, country: USA incidence: 6/7, conc. range: 1100-2600 μ g/kg, Ø conc.: 2083 μ g/kg, country: USA incidence: nc/175, conc. range: \leq 37,900 μ g/kg, Ø conc.: 2984 μ g/kg (all samples), country: USA incidence: 24/28, conc. range: \leq 1820 μ g / kg, Ø conc.: 870 μ g / kg, country: USA incidence: nc/80, conc. range: ≤ 1600 $\mu g / kg$, Ø conc.: 50 $\mu g / kg$ (all samples), country: USA incidence: nc/91, conc. range: \leq 8400 μ g / kg, Ø conc.: 370 μ g / kg, country: USA incidence: 284/886*, conc. range: 1-10 $\mu g / kg$ (276 samples), > 10 $\mu g / kg$ (8 sa), country: USA, *field-trial corn incidence: 13/99, conc. range: 1200-3200 μ g / kg, Ø conc.: 2400 μ g / kg, country: USA incidence: 5/6*, conc. range: < 50-4100 μ g / kg, Ø conc.: 2220 μ g / kg, country: USA, *export corn for Japan incidence: nc/846, conc. range: \leq 7470 μ g / kg, Ø conc.: 950 μ g / kg (all samples), country: USA, *export corn for South Africa incidence: nc/836, conc. range: \leq 7600 μ g / kg, Ø conc.: 960 μ g / kg (all samples), country: USA, *export corn for South Africa incidence: 79/79*, conc. range: 890-3860 μ g / kg, Ø conc.: 2350 μ g / kg, country: USA, *export corn for South Africa incidence: 5/5*, conc. range: 300-3400 μ g / kg, Ø conc.: 2400 μ g / kg, country: USA, *Indian maize incidence: 7/7*, conc. range: 80-16,310 μ g / kg, Ø conc.: 2883 μ g / kg, country: USA, *including 1 white maize sample

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incidence: 7/7, conc. range: 280-33,450 μ g / kg, Ø conc.: 6617 μ g / kg, country: USA incidence: 20/20, conc. range: 20-1420 μ g / kg, Ø conc.: 180 μ g / kg, country: Zambia incidence: 1/2, conc.: $125 \mu g/kg$, country: Zimbabwe incidence: 32/33*, conc. range: 30-1240 μ g / kg, Ø conc.: 488 μ g / kg, country: unknown origin, *imported from The Netherlands \rightarrow fumonisin B₂ incidence: 1/1, conc.: 800 μ g/kg, country: Argentina incidence: 17/17*, conc. range: 325-2680 μ g / kg, Ø conc.: 1137 μ g / kg, country: Argentina, *field-trial corn incidence: nc/547*, conc. range: ≤ 1250 μ g / kg, Ø conc.: 20 μ g / kg, country: Argentina, *export corn for South Africa incidence: 41/47*, conc. range: 50-500 μ g / kg, Ø conc.: 110 μ g / kg, country: Argentina, *export corn for South Africa incidence: 7/8, conc. range: 78-2267 μ g / kg, Ø conc.: 583 μ g / kg, country: Argentina incidence: 7/11*, conc. range: 20-680 µg/ kg, Ø conc.: 147 μ g / kg, country: Benin, *corn genotypes incidence: 2/2, conc. range: 50-105 μ g/kg, Ø conc. 77.5 μ g/kg, country: Botswana incidence: 48/48, conc. range: 1200-19,130 μg/kg, Ø conc.: 4213 μg/kg, country: Brazil incidence: 2/5, conc. range: 2300-4300 μ g / kg, Ø conc.: 3300 μ g / kg, country: China incidence: 3/27*, conc. range: 298-550 μ g/kg, Ø conc.: 448 μ g/kg, country: China, *high-EC area incidence: 2/20*, conc. range: 213-447 μ g / kg, Ø conc.: 330 μ g / kg, country: China, *low-EC area incidence: 4/7, conc. range: 96-2834 μ g / kg, Ø conc.: 1223 μ g / kg, country: China

incidence: 11/19*, conc. range: 10 µg/kg, \emptyset conc.: 10 μ g / kg, country: Croatia, *corn genotypes incidence: 8/16, conc. range: tr-376 μ g/kg, Ø conc.: 182 μ g/kg, country: Indonesia incidence: 3/12, conc. range: 231-556 μ g / kg, Ø conc.: 442 μ g / kg, country. Indonesia incidence: 7/7, conc. range: 30-1480 μ g/kg, Ø conc.: 839 μ g/kg, country: Italy incidence: 13/26, conc. range: 20-520 μ g/kg, Ø conc.: 143 μ g/kg, country: Italy incidence: 1/1, conc.: 275 µg/kg, country: Kenya incidence: 31/36*, conc. range: 70-48,400 μ g / kg, Ø conc.: 7500 μ g / kg, country: Korea, *moldy incidence: 8/35*, conc. range: 100-5400 μ g / kg, Ø conc.: 1100 μ g / kg, country: Korea, *healthy incidence: 1/8, conc.: 30 μ g / kg, country: Malawi incidence: 3/3, conc. range: 75-110 μ g/kg, Ø conc.: 90 μ g/kg, country: Mozambique incidence: 7/24, conc. range: 100-5500 μ g/kg, Ø conc.: 1600 μ g/kg, country: Nepal incidence: 1/7*, conc.: 10 µg/kg, country: Poland, *corn genotypes incidence: 6/50, conc. range: 58-1210 μ g / kg, Ø conc.: 286 μ g / kg, country: Philippines incidence: 8/9*, conc. range: 250-4450 μ g/kg, Ø conc.: 1211 μ g/kg, country: Portugal, *corn genotypes incidence: 1/6*, conc.: 10 µg/kg, country: Romania, *corn genotypes incidence: 1/1, conc.: 300 µg/kg, country: South Africa incidence: 3/12*, conc. range: 0-150 μ g/kg, Ø conc.: 83 μ g/kg, country: South Africa, *good corn, low-EC area

incidence: 10/12*, conc. range: $\leq 2250 \ \mu g/kg$, Ø conc.: 610 $\mu g/kg$, country: South Africa, *good corn, high-EC area incidence: 11/11*, conc. range: 150-6750 $\mu g/kg$, Ø conc.: 2500 $\mu g/kg$, country: South Africa, *moldy corn intended for beer brewing or animal feed, low-EC area incidence: 12/12*, conc. range: 900-16,300 $\mu g/kg$, Ø conc.: 7550 $\mu g/kg$, country: South Africa, *moldy corn intended for beer brewing or animal feed, high-EC area

incidence: $2/8^*$, conc. range: ≤ 970 μ g / kg, Ø conc.: 515 μ g / kg, country: South Africa, *good corn, low-EC area incidence: 5/6*, conc. range: 150-1320 μ g / kg, Ø conc.: 508 μ g / kg, country: South Africa, *good corn, high-EC area incidence: $6/7^*$, conc. range: ≤ 3700 μ g/kg, Ø conc.: 1277 μ g/kg, country: South Africa, *moldy corn, low-EC area incidence: 6/6*, conc. range: 750-22,960 μ g / kg, Ø conc.: 13,680 μ g / kg, country: South Africa, *moldy corn, high-EC area incidence: 50/68**, conc. range: < 50-1600 μ g/kg, Ø conc.: 190 μ g/kg (all samples), country: South Africa incidence: 55/66**, conc. range: < 20-1670 μg/kg, Ø conc.: 140 μg/kg (all samples), country: South Africa incidence: nc/77**, conc. range: \leq 270 μ g / kg, Ø conc.: 30 μ g / kg (all samples), country: South Africa incidence: nc/71**, conc. range: \leq 740 μ g / kg, Ø conc.: 50 μ g / kg (all samples), country: South Africa incidence: nc/113**, conc. range: \leq 1430 μ g/kg, Ø conc.: 80 μ g/kg (all samples), country: South Africa **white corn incidence: 31/53***, conc. range: < 50-700 μg/kg, Ø conc.: 50 μg/kg (all samples), country: South Africa incidence: 50/62***, conc. range: < 20-320 μ g/kg, Ø conc.: 70 μ g/kg (all samples), country: South Africa

incidence: nc/82***, conc. range: ≤ 690 μ g / kg, Ø conc.: 30 μ g / kg (all samples), country: South Africa incidence: nc/76, conc. range: ≤ 540 μ g / kg, Ø conc.: 30 μ g / kg (all samples), country: South Africa incidence: nc/117***, conc. range: \leq 5690 μ g / kg, Ø conc.: 220 μ g / kg (all samples), country: South Africa ***yellow corn incidence: 24/68*, conc. range: < 50-250 μ g / kg, Ø conc.: 130 μ g / kg, country: South Africa, *export corn for Taiwan incidence: 3/3, conc. range: 150-1300 μ g / kg, Ø conc.: 833 μ g / kg, country: South Africa incidence: 1/9, conc.: 60 µg/kg, country: Tanzania incidence: 12/18, conc. range: 50-1400 μ g / kg, Ø conc.: 251 μ g / kg, country: Thailand incidence: 12/27, conc. range: 50-1400 μ g / kg, Ø conc.: 251 μ g / kg, country: Thailand incidence: 1/1, conc.: 155 µg/kg, Uganda incidence: 6/7, conc. range: 70-460 μ g/kg, Ø conc.: 182 μ g/kg, country: USA incidence: 6/7, conc. range: 600-10,200 μ g / kg, Ø conc.: 2867 μ g / kg, country: USA incidence: nc/175, conc. range: \leq 12,300 μ g/kg, Ø conc.: 821 μ g/kg, country: USA incidence: 5/6*, conc. range: < 100-10,200 μ g/kg, Ø conc.: 3120 μ g/kg, country: USA incidence: nc/846*, conc. range: ≤ 2470 μ g / kg, Ø conc.: 120 μ g / kg (all samples), country: USA incidence: nc/836*, conc. range: \leq 3120 μ g / kg, Ø conc.: 140 μ g / kg (all samples), country: USA incidence: 79/79*, conc. range: 260-1120 μ g / kg, Ø conc.: 670 μ g / kg, country: USA, *export corn for South Africa

Maize

incidence: 7/7*, conc. range: 30-4020 μ g / kg, Ø conc.: 811 μ g / kg, country: USA, *including 1 white maize sample incidence: 5/5, conc. range: 32-4200 μ g / kg, Ø conc.: 1187 μ g / kg, country: USA incidence: 15/20, conc. range: 10-290 μ g / kg, Ø conc.: 50.7 μ g / kg, country: Zambia incidence: 1/2, conc.: 40 µg/kg, country: Zimbabwe \rightarrow fumonisin B₃ incidence: $17/17^*$, conc. range: $\leq 110-855$ μ g / kg, Ø conc.: 372 μ g / kg, country: Argentina, *field-trial corn incidence: 28/47*, conc. range: 50-500 μ g / kg, Ø conc.: 80 μ g / kg, country: Argentina, *export corn for South Africa incidence: 6/8, conc. range: 50-980 μ g / kg, Ø conc.: 348 μ g / kg, country: Argentina incidence: 2/2, conc. range: 40-70 µg/kg, \emptyset conc.: 55 μ g/kg, country: Botswana incidence: 2/7, conc. range: 230-545 μ g/kg, Ø conc.: 388 μ g/kg, country: China incidence: 4/16, conc. range: 57-222 μ g/kg, Ø conc.: 108 μ g/kg, country: Indonesia incidence: 1/1, conc.: 780 μg/kg, country: Kenya incidence: 31/36*, conc. range: 50-10,600 μ g / kg, Ø conc.: 6300 μ g / kg, country: Korea, *moldy incidence: 7/35*, conc. range: 50-500 μ g/kg, Ø conc.: 300 μ g/kg, country: Korea, *healthy incidence: 55/66**, conc. range: < 20-400 μ g / kg, Ø conc.: 40 μ g / kg (all samples), country: South Africa incidence: nc/77**, conc. range: \leq 340 μ g/kg, Ø conc.: 10 μ g/kg (all samples), country: South Africa incidence: nc/71**, conc. range: ≤ 180 μ g / kg, Ø conc.: 10 μ g / kg (all samples), country: South Africa

incidence: nc/113**, conc. range: ≤ 400 μ g / kg, Ø conc.: 30 μ g / kg (all samples), country: South Africa **white corn incidence: 50/62***, conc. range: < 20-200 μ g/kg, Ø conc.: 20 μ g/kg (all samples), country: South Africa incidence: nc/82***, conc. range: ≤ 120 μ g / kg, Ø conc.: < 10 μ g / kg (all samples), country: South Africa incidence: nc/76, conc. range: \leq 330 μ g / kg, Ø conc.: 10 μ g / kg (all samples), country: South Africa incidence: nc/117, conc. range: \leq 1960 μ g / kg, Ø conc.: 110 μ g / kg (all samples), country: South Africa ***yellow corn incidence: 1/1, conc.: 85 μ g/kg, country: Uganda incidence: nc/175, conc. range: 2800 μ g / kg, Ø conc.: 290 μ g / kg, country: USA incidence: 79/79*, conc. range: 80-550 μ g / kg, Ø conc.: 250 μ g / kg, country: USA, *export corn for South Africa fumonisin (no specification) (\rightarrow fumonisins) incidence: 17/17*, conc. range: 500-48,500 μ g/kg, Ø conc.: 17,864 μ g/kg, country: USA, *hybrid maize incidence: 4/4*, conc. range: 37-1400 μ g/kg, Ø conc.: 411 μ g/kg, country: Germany, *organic produce incidence: 3/3, conc. range: 25-50 µg/kg, \emptyset conc.: 40 µg/kg, country: Mozambique \rightarrow fusaproliferin incidence: 9/22*, conc. range: 600-500,000 μ g / kg, Ø conc.: 72,222 μ g / kg, country: Italy, *visibly infected \rightarrow fusarenon X incidence: 2/2*, conc. range: 400-900 μ g / kg, Ø conc.: 650 μ g / kg, country: Austria, *visibly moldy (Fusarium spp.)

incidence: 5/15, conc. range: 15-72 μ g/kg, Ø conc.: 27 μ g/kg, country: Korea \rightarrow fusarin C incidence: 2/2, conc. range: 20-280 μ g / kg, Ø conc.: 150 μ g / kg, country: South Africa \rightarrow HT-2 toxin incidence: 2/52, conc. range: 500-800 μ g/kg, Ø conc.: 650 μ g/kg, country: Germany incidence: 1/56, conc.: 600 µg/kg, country: Germany incidence: 2/11, conc. range: 500-700 μg/kg, country: Hungary incidence: 3*/162, conc. range: 53,000-645,000 μg / kg, Ø conc.: 294,333 μg / kg, country: Poland, *heavily damaged kernels → kojic acid incidence: 3/155, conc.: nc, country: South Africa \rightarrow moniliformin incidence: 2/12, conc. range: 60-200 μ g/kg, Ø conc.: 130 μ g/kg, country: Canada incidence: 23/58, conc. range: 80-650 μg/kg, country: Germany incidence: 15/29, conc. range: \leq 280 μg/kg, country: New Zealand incidence: 20/20*, conc. range: 4200-399,300 μg / kg, Ø conc.: 97,710 μg / kg, country: Poland incidence: 57/57*, conc. range: 16,800-425,000 μg/kg, Ø conc.: 172,000 μg/kg, country: Poland *hand selected, visible fungal damage incidence: 15/36, conc. range: tr-12,000 µg/kg, country: South Africa incidence: 24/24, conc. range: 350-11,570 μg/kg, Ø conc.: 3500 μg/kg*, 800 μg / kg**, country: South Africa, *low-prevalence EC area, **high-prevalence EC area incidence: 2*/2, conc. range: 16,000-25,000 μg/kg, Ø conc.: 20,500 μg/kg, country: South Africa, *moldy

incidence: 64/64, conc. range: < 50-3160 μ g / kg, country: different countries, mainly Africa → neosolaniol incidence: 1/100, conc.: traces, country: Argentina incidence: 2*/162, conc. range: 19,400-27,200 μg / kg, Ø conc.: 23,300 μg / kg, country: Poland, *heavily damaged kernels \rightarrow nivalenol incidence: 5/100, conc. range: tr-500 µg/kg, country: Argentina incidence: 2/2, conc. range: 700-2200 μ g / kg, Ø conc.: 1450 μ g / kg, country: Austria incidence: 2/2*, conc. range: 500-1800 μg/kg, country: Austria, *visibly moldy (Fusarium spp.) incidence: 1/1, conc.: $12 \mu g / kg$, country: Canada incidence: 28/28, conc. range: \leq 4050 μ g / kg, Ø conc.: 1960 μ g / kg, country: China incidence: 24/24, conc. range: 54-2760 μ g / kg, Ø conc.: 757 μ g / kg, country: China incidence: 2/16, conc. range: 49-169 μ g/kg, Ø conc.: 109 μ g/kg, country: Indonesia incidence: 8/15, conc. range: 26-332 μ g/kg, Ø conc.: 168 μ g/kg, country: Korea incidence: 32/36*, conc. range: 6-15,600 μ g / kg, Ø conc.: 1700 μ g / kg, country: Korea, *moldy incidence: 6/35*, conc. range: 20-200 μ g/kg, Ø conc.: 80 μ g/kg, country: Korea, *healthy incidence: 1/3, conc.: 624 μ g/kg, country: Korea incidence: 6/9, \emptyset conc.: 892 µg/kg, country: Nepal incidence: 73/91, conc. range: \leq 3600 μ g / kg, country: New Zealand incidence: 7/50, conc. range: 18-102 μ g / kg, Ø conc.: 43 μ g / kg, country: Philippines

Maize

incidence: 6/36, conc. range: tr-240 μg/kg, country: South Africa incidence: 24/24, conc. range: 880-15,200 μ g / kg, Ø conc.: 4600 μ g / kg*, 1800 μg/kg**, country: South Africa *low-prevalence EC area, **high-prevalence EC area \rightarrow ochratoxin A incidence: 3/27, conc. range: 5-100 μg/kg, country: Austria incidence: 1/12, conc.: $32 \mu g/kg$, country: Brazil incidence: 12/52, conc. range: 25-35 μg/kg, country: Bulgaria incidence: 87/151*, conc. range: 0.2-1418 μ g / kg, country: Bulgaria, *area with endemic nephropathy incidence; 30/113, conc. range: 0.2-235 μg/kg, country: Bulgaria incidence: 1/28, conc.: 55 µg/kg, country: Chile incidence: 1/3, conc.: $12 \mu g / kg$, country: Egypt incidence: 18/924, conc. range: 15-200 μg/kg, country: France incidence: 2/75, conc.: 10 μg/kg, Ø conc.: 10 µg/kg, country: France incidence: 2/49*, conc. range: 18-22 μ g / kg, Ø conc.: 20 μ g / kg, country: Germany, *moldy incidence: 3/40, conc. range: 1.7-82 μ g/kg, Ø conc.: 80.3 μ g/kg, country: Germany incidence: 1/7, conc.: 0.1 µg/kg, country. Germany incidence: 38/112, conc. range: $\leq 0.7 \ \mu g/$ kg, country: Italy incidence: 14/90, conc. range: $\leq 2.0 \ \mu g/$ kg, country: Italy incidence: 39/111, conc. range: 0.1-1.02 μg/kg, country: Italy incidence: 1/22, conc.: nc, country: India incidence: 1/26, conc.: $3 \mu g/kg$, country: Indonesia incidence: 2/123, conc. range: 25-400 μ g / kg, Ø conc.: 213 μ g / kg, country: Poland

incidence: 1/30, conc.: 2.5 µg/kg, country: Spain incidence: 2/167, conc. range: ca. 10 μg / kg, country: Turkey incidence: 11/29, conc. range: < 50-500 μg/kg, country: UK incidence: 5/39, conc. range: \leq 4.9-11.2 μ g / kg, country: UK incidence: 4/11, conc. range: $\leq 0.8 \ \mu g/$ kg, country: UK incidence: 11/19, conc. range: $\leq 0.7 \ \mu g/$ kg, country: UK incidence: 3/293, conc. range: 83-166 μ g / kg, Ø conc.: 123 μ g / kg, country: USA incidence: 1/283, conc. range: 130 µg/kg, country: USA incidence: 50/542, conc. range: 6-140 µg/kg, country: Yugoslavia incidence: 50/191, conc. range: 45-5100 μ g / kg, Ø conc.: 490 μ g / kg, country: Yugoslavia incidence: 2/48, conc. range: 14-90 μ g / kg, Ø conc.: 40.6 μ g / kg, country: Yugoslavia \rightarrow ochratoxin B incidence: 2/293, conc. range: traces, country: Canada \rightarrow penicillic acid incidence: 7/20*, conc. range: 5-231 μ g / kg, Ø conc. 59 μ g / kg, country: USA, *mold damaged incidence: 48/48*, conc. range: 5-184 μ g / kg, Ø conc.: 46 μ g / kg, country: USA, *mold damaged \rightarrow sterigmatocystin incidence: 4/155, conc.: nc, country: South Africa incidence: 10/167, conc. range: ca. 20 μg / kg, country: Turkey incidence: 2/29, conc. range: > 10 μ g/kg, country: UK \rightarrow T-2 toxin incidence: 1/52, conc.: 10 µg/kg, country: Germany incidence: 4/56, conc. range: 100-200 μg/kg, country: Germany

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incidence: 7/11, conc. range: 100-4400 μg/kg, country: Hungary incidence: 5/150, conc. range: 500-5000 μg/kg, country: Hungary incidence: 1*/nc, conc.: 4000 µg/kg, country: India, *moldy incidence: 1/4, conc.: 0.8 μ g/kg, country: Italy incidence: 3/100, conc. range: nc, country: Italy incidence: 3/162, conc. range: 47,000-992,000 μg / kg, Ø conc.: 411,333 μg / kg, country: Poland incidence: 9/118, conc. range: 78-650 μg/kg, country: Taiwan incidence: 8/100, conc. range: nc, country: Yugoslavia incidence: 1/1, conc.: 2000 µg/kg, country: USA incidence: 15/100, conc. range: 900-2400 μg/kg, country: USA incidence: 9/118, conc. range: 78-650 μg/kg, country: USA incidence: 93/173, conc. range: 0.2-1 μg/kg, country: USA incidence: 13/20, conc. range: ≤ 200 μ g / kg, Ø conc.: 74 μ g / kg, country: New Zealand T-2 tetraol incidence: 1*/162, conc.: 36,200 µg/kg, country: Poland, *heavily damaged kernels T-2 triol incidence: 2/56, conc. range: 300 µg/kg, \emptyset conc. 300 μ g/kg, country: Germany incidence: 2*/162, conc. range: 9700-14,500 μg / kg, Ø conc.: 12,100 μg / kg, country: Poland, *heavily damaged kernels → zearalenols incidence: nc/6, conc. range: 20-90 μ g / kg, country: Italy zearalenone incidence: 15/20, Ø conc.: 6 µg/kg, country: Argentina incidence: 676/2271, conc. range: \leq 2000 μ g/kg, Ø conc.: 165 μ g/kg, country: Argentina

incidence: 16/55, conc. range: 200-750 µg/kg, country: Argentina incidence: 9/150, conc. range: 40-350 μ g / kg, Ø conc.: 210 μ g / kg, country: Argentina incidence: 148/174, conc. range: ≤ 2070 μ g / kg, Ø conc.: 230 μ g / kg, country: Australia incidence: 3/3, conc. range: 1100-1300 μ g/kg, Ø conc.: 1200 μ g/kg, country: Austria incidence: 27/51, conc. range: 1-200 μ g / kg, Ø conc.: 70 μ g / kg, country: Austria incidence: 41/78, conc. range: \leq 70 µg/kg, \emptyset conc.: 9 µg/kg, country: Austria incidence: 3/6, conc. range: 420-1000 μ g / kg, Ø conc.: 740 μ g / kg, country: Austria incidence: 1/1, conc.: 33 μ g/kg, country: Canada incidence: 23/81*, conc. range: 130-475 μg/kg, country: Canada, *domestic, maize and maize products incidence: 1/61*, conc.: 200 µg/kg, country: Canada, *imported, maize and maize products incidence: 62/75, conc. range: 10,000-175,000 µg/kg, country: France incidence: 16/59, conc. range: 1-260 μ g / kg, Ø conc.: 50 μ g / kg, country: Germany incidence: 2/174, conc. range: 10-1200 μg/kg, country: Germany incidence: 2/4*, conc. range: 49-92 μ g/kg, Ø conc.: 70.5 μ g/kg, country: Germany, *organic produce incidence: 8/11, conc. range: 700-7500 μg/kg, country: Hungary incidence: 1*/nc, conc.: 16,000 µg/kg, country: India, *moldy incidence: 2/16, conc. range: 11-12 μ g/kg, Ø conc.: 11.5 μ g/kg, country: Indonesia incidence: 1/3, \emptyset conc.: 35 µg/kg, country: Italy incidence: 31/111, conc. range: 51-670 μ g / kg, country: Italy

Maize

incidence: nc/6, conc. range: 400-7400 μ g / kg, country: Italy incidence: 3/41, conc. range: 40-2000 μg/kg, country: Italy incidence: 1/1, conc.: 40 µg/kg, country: Kenya incidence: 1/15, conc.: 71 µg/kg, country: Korea incidence: 32/36*, conc. range: 2-7300 μ g / kg, Ø conc.: 600 μ g / kg, country: Korea, *moldy incidence: 7/35*, conc. range: 2-300 μ g / kg, Ø conc.: 70 μ g / kg, country: Korea, *healthy incidence: 1/1, conc.: 400 µg/kg, country: Malawi incidence: 6/139, conc. range: nc, country: Mexico incidence: 5/9, \emptyset conc.: 819 µg/kg, country: Nepal incidence: 69/91, conc. range: \leq 500 μg/kg, country: New Zealand incidence: 2/50, conc. range: 59-505 μ g / kg, Ø conc.: 282 μ g / kg, country: Philippines incidence: 5/9*, conc. range: 50-2050 μ g / kg, Ø conc.: 1048 μ g / kg, country: Poland, *healthy and damaged kernels incidence: 8/8, conc. range: 620-72,000 μg/kg, country: Portugal incidence: 2/44, conc. range: 20-503 µg/kg, country: South Africa incidence: 14/24, conc. range: 120-3280 $\mu g / kg$, Ø conc.: 1200 $\mu g / kg^*$, 400 $\mu g /$ kg**, country: South Africa, *low-prevalence EC area, **high-prevalence EC area incidence: 2*/2, conc. range: 4000-8000 μ g /kg, Ø conc.: 6000 μ g / kg, country: South Africa, *moldy incidence: 9/155, conc.: nc, country: South Africa incidence: 2/9, conc. range: 40-80 µg/kg, \emptyset conc.: 60 µg/kg, country: Tanzania incidence: 1/27, conc.: 923 µg/kg, country: Thailand incidence: 6/76*, conc. range: 100-200 μ g / kg (2 samples), > 200 μ g / kg (4 sa), country: Uruguay, *and by-products

incidence: 4/nc, conc. range: 2310-35,600 µg/kg, country: Yugoslavia incidence: 54/116, conc. range: 10-275,800 µg/kg, country: Yugoslavia incidence: 5/191, conc. range: 43-10,000 μg/kg, country: Yugoslavia incidence: 23/54, conc. range: 700-37,500 µg/kg, country: Yugoslavia incidence: 4/29, conc. range: ca. 2000 μ g / kg, country: UK incidence: 7/73, conc. range: 49-303 μg/kg, country: USA incidence: 6/576, conc. range: 450-800 μ g / kg, Ø conc. 624 μ g / kg, country: USA incidence: 38/223, conc. range: 100-5000 μ g / kg, Ø conc. 900 μ g / kg, country: USA incidence: 2/283, conc. range: 800- > 1250 µg/kg, country: USA incidence: 5/293, conc. range: 450-750 μ g / kg, country: USA incidence: 17/20*, conc. range: 200-13,200 μ g/kg, Ø conc.: 2700 μ g/kg, country: USA, *moldy incidence: 6/26, conc. range: 200-500 μg/kg, country: USA incidence: 19/315, conc. range: < 100-210 μg/kg, country: USA incidence: 18/315, conc.: 400 µg/kg, country: USA incidence: 4/12, Ø conc.: 10 μg/kg, country: Yemen \rightarrow cereals

Maize, boiled may contain the following \rightarrow mycotoxins: aflatoxin (\rightarrow aflatoxins) incidence: 16/24, Ø conc.: 9 µg/kg, country: Philippines

Maize, brewers may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 8/11, conc. range: 20-100 µg/kg (6 samples), 101-500 µg/kg (1 sa), country: UK → zearalenone incidence: nc/17, Ø conc.: 290 µg/kg, country: Zambia incidence: nc/13, Ø conc.: 680 µg/kg, country: Zambia

Maize, brewers flaked may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 5/6, conc. range: 10-110 µg/kg, Ø conc.: 50 µg/kg, country: UK

Maize, brewers grits may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 2/3, conc. range: 40-140 µg/kg, Ø conc.: 90 µg/kg, country: UK

Maize, canned may contain the following \rightarrow mycotoxins: aflatoxin (\rightarrow aflatoxins) incidence: 3/4, conc. range: $\leq 25 \ \mu\text{g}/\text{kg}$, \emptyset conc.: $6 \ \mu\text{g}/\text{kg}$, country: Philippines \rightarrow fumonisin B₁ incidence: 1/1, conc.: 26 $\ \mu\text{g}/\text{kg}$, country: USA hydrolyzed fumonisin B₁ (HBF₁) incidence: 1/1, conc.: nc, country: USA

Maize, dried may contain the following \rightarrow mycotoxins: aflatoxin (\rightarrow aflatoxins) incidence: 33/660, conc. range: \leq 1152 μ g/kg, \emptyset conc.: 76 μ g/kg, country: Philippines

Maize, fiber cereal may contain the following \rightarrow mycotoxins: \rightarrow fumonisin B₁ incidence: 1/1, conc.: 130 µg/kg, country: USA incidence: 1/1, conc.: 60 µg/kg, country: Venezuela \rightarrow fumonisin B₂ incidence: 1/1, conc.: 30 µg/kg, country: Venezuela **Maize, hominy** may contain the following \rightarrow mycotoxins: \rightarrow fumonisin B₁ incidence: 1/1, conc.: 60 µg/kg, country: USA \rightarrow fumonisin B₂ incidence: 1/1, conc.: 20 µg/kg, country: USA

Maize, infant cereal may contain the following \rightarrow mycotoxins: \rightarrow fumonisins (no specification) incidence: 1/1, conc.: 200 µg/kg, country: USA

Maize, infant cream corn may contain the following \rightarrow mycotoxins: \rightarrow fumonisins (no specification) incidence: 1/1, conc.: 200 µg/kg, country: USA

Maize, popped may contain the following \rightarrow mycotoxins: \rightarrow fumonisin B₁ incidence: 3/5, conc. range: \leq 300 µg/kg, country: The Netherlands

Maize, preharvest may contain the following \rightarrow mycotoxins: \rightarrow zearalenone incidence: 1/116, conc.: < 5000 µg/kg, country: Spain

Maize, puffed may contain the following \rightarrow mycotoxins: \rightarrow fumonisin B₁ incidence: 6/6, conc. range: 790-6100 $\mu g / kg, \emptyset$ conc.: 3145 $\mu g / kg$, country: Italy \rightarrow fumonisin B₂ incidence: 6/6, conc. range: 110-740 $\mu g / kg, \emptyset$ conc. 397 $\mu g / kg$, country: Italy

Maize, quality-protein may contain the following \rightarrow mycotoxins:

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 \rightarrow fumonisin B₁ incidence: nc/12, conc. range: ≤ 2040 μ g / kg, Ø conc.: 410 μ g / kg (all samples), country: South Africa incidence: nc/59, conc. range: \leq 4400 μ g / kg, Ø conc.: 340 μ g / kg (all samples), country: South Africa \rightarrow fumonisin B₂ incidence: nc/12, conc. range: \leq 1090 μ g / kg, Ø conc.: 120 μ g / kg (all samples), country: South Africa incidence: nc/59, conc. range: \leq 1290 μ g / kg, Ø conc.: 110 μ g / kg (all samples), country: South Africa \rightarrow fumonisin B₃ incidence: nc/12, conc. range: ≤ 60 μ g / kg, Ø conc.: < 10 μ g / kg (all samples), country: South Africa incidence: nc/59, conc. range: \leq 800 μ g / kg, Ø conc.: 40 μ g / kg (all samples), country: South Africa

Maize, shelled may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins

incidence: 36/1594, conc. range: \leq 37 μ g / kg, Ø conc.: 9 μ g / kg, country: USA incidence: 21/60, conc. range: \leq 348 μ g/kg, Ø conc.: 66 μ g/kg, country: USA incidence: 394/1283, conc. range: \leq 306 μ g / kg, Ø conc.: 35 μ g / kg, country: USA incidence: 152/297, conc. range: \leq 3190 μ g / kg, Ø conc.: 50 μ g / kg, country: USA incidence: 35/81, conc. range: \leq 710 μ g/kg, Ø conc.: 49 μ g/kg, country: USA incidence: 16/34, conc. range: ≤ 145 μ g / kg, Ø conc.: 17 μ g / kg, country: USA incidence: 4/169, conc. range: $\leq 5 \ \mu g / kg$, \emptyset conc.: 2 µg/kg, country: USA incidence: 432/1385, conc. range: \leq 3300 μ g / kg, Ø conc.: 242 μ g / kg, country: USA incidence: 44/62, conc. range: ≤ 1524 μ g/kg, Ø conc.: 142 μ g/kg, country: USA incidence: 18/31, conc. range: ≤ 631 μ g / kg, Ø conc.: 82 μ g / kg, country: USA

incidence: 26/11, conc. range: $\leq 16 \mu g/kg$, \emptyset conc.: 8 $\mu g/kg$, country: USA incidence: 116/1395, conc. range: $\leq 1290 \mu g/kg$, \emptyset conc.: 47 $\mu g/kg$, country: USA incidence: 24/148, conc. range: $\leq 364 \mu g/kg$, \emptyset conc.: 57.8 $\mu g/kg$, country: USA \rightarrow deoxynivalenol incidence: 64/96, conc. range: tr-500 $\mu g/kg$ (53 samples), 500-1000 $\mu g/kg$ (10 sa), 1000-2000 $\mu g/kg$ (1 sa), country: USA

Maize, steeped may contain the following \rightarrow mycotoxins: \rightarrow fumonisins Experimental studies showed an increased fumonisin concentration in the steeping water with a similar decrease in the \rightarrow maize kernels. The different rates of interchange of fumonisin B₁ and FB₂ between the solid matrix and the water solution may be explained by the different polarities of the fumonisins. Although FB₁ and FB₂ have been detected in both the germ and the remaining ker-

nel, the germ fraction contained lower fumonisin levels.

Maize, sweet may contain the following \rightarrow mycotoxins: \rightarrow fumonisin B₁ incidence: 11/40, conc. range: < 10-190 μg/kg, country: Germany incidence: 5/5, conc. range: 60-790 μ g/kg, Ø conc.: 298 μ g/kg, country: Italy incidence: 1/7, conc.: 70 µg/kg, country: Switzerland incidence: $12/24^*$, conc. range: ≤ 1089 μ g / kg, Ø conc.: 400 μ g / kg, country: Thailand, *canned \rightarrow fumonisin B₂ incidence: $6/24^*$, conc. range: ≤ 658 μ g / kg, Ø conc.: 64.5 μ g / kg, country: Thailand, *canned \rightarrow fumonisins (FB₁, FB₂, FB₃)

incidence: 1/22, conc.: 11 μ g / kg, country: UK

Maize-based thickeners may contain the following \rightarrow mycotoxins: \rightarrow fumonisins (FB₁, FB₂, FB₃) incidence: 4/21, conc. range: 14-110 µg / kg, Ø conc.: 23 µg / kg, country: UK

Maize bran may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin incidence: 2/2, conc. range: 37-71 µg/kg, \emptyset conc.: 54 µg/kg, country: Philippines \rightarrow fumonisin B₁ incidence: 3/4, conc. range: 60-330 μ g / kg, Ø conc.: 168 μ g / kg, country: USA incidence: 1/1, conc.: 290 µg/kg, country: USA \rightarrow fumonisin B₂ incidence: 3/3, conc. range: 10-40 µg/kg, \emptyset conc.: 23.3 μ g / kg, country: USA incidence: 1/1, conc.: 70 µg/kg, country: USA \rightarrow bran

 $\begin{array}{ll} \mbox{Maize chips} & may contain the following \\ \rightarrow mycotoxins: \\ \rightarrow fumonisin B_1 \\ incidence: 2/2, conc.: tr-37 \ \mu g / kg, country: Canada \\ incidence: 3/9, conc. range: <math>\leq 160 \ \mu g / kg$, country: The Netherlands \\ \mbox{Maize flakes} & may contain the following \\ \end{array}

 \rightarrow mycotoxins:

→ fumonisin B₁ incidence: 2/5, conc. range: 10 µg/kg, Ø conc.: 10 µg/kg, country: USA → cereal flakes, → corn flakes, → oat flakes

Maize flour Pattern of \rightarrow zearalenone distribution in \rightarrow maize kernels is the same as in the case of aflatoxin (\rightarrow aflatoxins). After dry \rightarrow milling the largest

amounts were found in the high fat fractions (oil and feed usage). Processing of zearalenone-contaminated corn (120 μ g/kg) led to 15 μ g zearalenone/kg in the starch. The starch of wetmilled maize did not show any zearalenone contamination but gluten and solubes contained about 50% and ca. 20% zearalenone from the whole maize. Wet milling of maize contaminated with \rightarrow nivalenol, \rightarrow deoxynivalenol and zearalenone caused a significant accumulation of the highly water-soluble NIV and DON in the concentrated steep liquor fractions (\leq 8800 µg/kg). Low levels were observed in the solid (germ, fibre and gluten) fractions (< $300 \mu g/kg$). In contrast, the relatively water insoluble zearalenone accumulated in the solids (2200-4800 μ g/kg), while only 600 μ g/kg were found in the concentrated steep liquor. The starch fractions were almost free of \rightarrow mycotoxins.

Dry milling of deoxynivalenol contaminated maize resulted in the accumulation of the mycotoxin in the maize germ meal (animal feed).

Most of \rightarrow T-2 toxin (> 60%) was removed by wet milling with the steep and process water. The starch contained less than 5% while the remainder is found in the germ, gluten und fibre. In a laboratory simulated scale aflatoxin, \rightarrow fumonisins, T-2 toxin, and zearalenone accumulated in the solubes fractions during milling. While the fumonisins and zearalenol also concentrated in the gluten, aflatoxin and fumonisins were found in the fibre fraction, too. Maize flour may contain the following

Maize flour may contain the following mycotoxins: \rightarrow aflatoxin B₁

incidence: 1/5, conc.: 15 μg/kg, country: Japan

incidence: 1/4, conc.: 31 μg/kg, country: Japan

Maize flour

incidence: 11/11, conc. range: 3.7-37 μ g / kg, Ø conc.: 18.95 μ g / kg, country: Thailand \rightarrow aflatoxin B₂ incidence: 1/5, conc.: 5.2 µg/kg, country: Japan incidence: 1/4, conc.: 5.3 µg/kg, country: Japan incidence: 11/11, conc. range: 2.3-9.9 μ g / kg, Ø conc.: 5.96 μ g / kg, country: Thailand \rightarrow citrinin incidence: 1/5 conc.: 27 µg/kg, country: Japan incidence: 1/4, conc.: 73 µg/kg, country: Japan incidence: 14/23, conc. range: \leq 1390 μg/kg, country: Japan incidence: 11/11, conc. range: 10-98 μ g/kg, Ø conc.: 58.9 μ g/kg, country: Thailand deoxynivalenol incidence: nc, Ø conc.: 180 μg/kg, country: Canada incidence: 1/2, conc.: 240 µg/kg, country: UK incidence: 5/5, conc. range: 20-50 µg/kg, country: UK incidence: nc/4, conc. range: 17-67 μ g / kg, country: UK \rightarrow fumonisin B₁ incidence: 4/4, conc. range: 35-255 μ g/kg, Ø conc.: 185 μ g/kg, country: Botswana incidence: 11/39, conc. range: < 100-1600 μ g / kg, Ø conc.: 550 μ g / kg, country: Canda incidence: 3/4, conc. range: 60-200 μ g/kg, Ø conc.: 100 μ g/kg, country: China incidence: 1/1, conc.: 3540 μ g/kg, country: Italy incidence: 2/2, conc. range: 60-70 µg/kg, \emptyset conc.: 65 µg/kg, country: South Africa incidence: nc/3, conc. range: 0-310 μ g / kg, Ø conc.: 100 μ g / kg (all samples), country: South Africa

incidence: nc/13, conc. range: 40-3910 μ g / kg, Ø conc.: 550 (all samples), country: South Africa incidence: 1/3, conc. range: 50-70 µg/kg, country: Spain incidence: 5/25, conc. range: < 30-330 μ g / kg, Ø conc.: 130 μ g / kg, country: Spain incidence: 1/2, conc.: 608 µg/kg, country: Thailand incidence: 6/6, conc. range: 480-880 μ g / kg, Ø conc.: 660 μ g / kg, country: Thailand incidence: 5/7*, conc. range: 40-90 μ g / kg, Ø conc.: 58 μ g / kg, country: The Netherlands, *mixes incidence: 2/6*, conc. range: 8-25 µg/kg, country: The Netherlands, *mixes incidence: 1/1, conc.: 740 µg/kg, country: Zambia incidence: 4/4, conc. range: 55-1910 μ g/kg, Ø conc.: 625 μ g/kg, country: Zimbabwe \rightarrow fumonisin B₂ incidence: 2/4, conc. range: 75-85 µg/kg, \emptyset conc.: 80 μ g/kg, country: Botswana incidence: 1/1, conc.: 840 µg/kg, country: Italy incidence: nc/13, conc. range: 0-810 μ g/kg, Ø conc.: 90 (all samples), country: South Africa incidence: nc/25, conc. range: 50-60 μg/kg, country: Spain incidence: nc/6, conc. range: 120-240 μ g / kg, Ø conc.: 160 μ g / kg, country: Thailand incidence: 1/1, conc.: 380 μ g/kg, country: Zambia incidence: 2/4, conc. range: 150-620 μ g / kg, Ø conc.: 385 μ g / kg, country: Zimbabwe \rightarrow fumonisin B₃ incidence: 1/4, conc.: 30 μ g / kg, country: Botswana incidence: nc/13, conc. range: 0-470 μ g/kg, Ø conc.: 40 (all samples), country: South Africa incidence: 1/1, conc.: 85 µg/kg, Zambia

incidence: 2/4, conc. range: 55-205 μ g / kg, Ø conc.: 130 μ g / kg, country: Zimbabwe \rightarrow fumonisins (FB₁, FB₂) incidence: 1/4, conc.: 218 μ g/kg, country: UK \rightarrow moniliformin incidence: 6/6, conc. range: < 50-250 μg/kg, country: UK, USA \rightarrow ochratoxin A incidence: 4/13, conc. range: 50-200 μ g / kg, country: UK incidence: 1/4, conc.: 0.6 µg/kg, country: UK zearalenone incidence: 1/4, conc.: 100 μ g/kg, country: Botswana incidence: nc/4, conc. range: 6.5-40.8 μg/kg, country: UK \rightarrow flour

Maize grits (Syn.: polenta, semolina) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 14/35, conc. range: 0.5-1 μ g / kg (8 samples), 1-3 μ g / kg, (6 sa), country: Switzerland → deoxynivalenol incidence: 3/3, conc. range: 130-910 μ g / kg, Ø conc.: 640 μ g / kg, country: Germany incidence: 1/1*, conc.: 170 µg/kg, country: Germany, *organic produce \rightarrow fumonisin B₁ incidence: 1/3, conc.: 800 μ g/kg, country: Canada incidence: 1/1, conc.: 3760 µg/kg, country: Italy incidence: 6/6, conc. range: 420-3730 μ g / kg, Ø conc.: 2152 μ g / kg, country: Italy incidence: 6/6, conc. range: 45.6-1230 μ g / kg, country: Italy incidence: 20/20, conc. range: 150-3760 μ g / kg, Ø conc.: 1380 μ g / kg, country: Italy

incidence: 2/2, conc. range: < 10-20.8 μg/kg, country: Germany incidence: 5/5, conc. range: < 10-33.1 μg/kg, country: Germany incidence: 14/17, conc. range: 200-2600 μ g / kg, Ø conc.: 500 μ g / kg, country: Japan incidence: 10/18, conc. range: 0-190 μ g/kg, Ø conc.: 125 μ g/kg, country: South Africa incidence: nc/8, conc. range: 0-740 μ g/kg, Ø conc.: 130 μ g/kg (all samples), country: South Africa incidence: nc/73, conc. range: 0-1380 μ g / kg, Ø conc.: 140 μ g / kg (all samples), country: South Africa incidence: 3/15, conc. range: 50-90 µg/kg, country: Spain incidence: 34/55, conc. range: 0-790 μ g / kg, Ø conc.: 260 μ g / kg, country: Switzerland incidence: 5/5, conc. range: 250-1820 μ g/kg, Ø conc.: 830 μ g/kg, country: Thailand incidence: 2/3, conc. range: \leq 40 µg/kg, country: The Netherlands incidence: 10/10, conc. range: 105-2545 μ g/kg, Ø conc.: 601 μ g/kg, country: USA incidence: 4/4, conc. range: 140-270 μ g/kg, Ø conc.: 198 μ g/kg, country: USA incidence: 1/3, conc.: 80 μ g / kg, country: USA incidence: 5/5, conc. range: 140-270 μ g / kg, Ø conc.: 200 μ g / kg, country: USA \rightarrow fumonisin B₂ incidence: 1/1, conc.: 910 µg/kg, country: Italy incidence: 6/6, conc. range: 80-840 μ g / kg, Ø conc.: 477 μ g / kg, country: Italy incidence: nc/20, conc. range: 60-910 μ g / kg, Ø conc.: 370 μ g / kg, country: Italy

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Maize grits

incidence: 5/17, conc. range: 300-2800 μ g / kg, Ø conc.: 1000 μ g / kg, country: Japan incidence: 4/18, conc. range: 0-120 μ g/kg, Ø conc.: 85 μ g/kg, country: South Africa incidence: nc/8, conc. range: 0-70 µg/kg, country: South Africa incidence: nc/73, conc. range: 0-420 μ g / kg, Ø conc.: 20 μ g / kg (all samples), country: South Africa incidence: 13/55, conc. range: 0-160 μ g / kg, Ø conc.: 100 μ g / kg, country: Switzerland incidence: nc/5, conc. range: 70-400 μ g / kg, Ø conc.: 190 μ g / kg, country: Thailand incidence: 5/10, conc. range: 0-1065 μ g / kg, Ø conc.: 375 μ g / kg, country: USA incidence: 3/4, conc. range: 60-110 μ g / kg, Ø conc.: 86.6 μ g / kg, country: USA incidence: nc/5, conc. range: 10-111 μ g / kg, Ø conc.: 80 μ g / kg, country: USA \rightarrow fumonisin B₃ incidence: nc/73, conc. range: 0-160 µg/kg, country: South Africa \rightarrow fumonisins (FB₁, FB₂, FB₃) incidence: 4/4, conc. range: 20-1200 μ g/kg (HPLC), Ø conc.: 400 μ g/kg, country: Germany incidence: 16/20, conc. range: 16-2124 μ g / kg, Ø conc.: 531 μ g / kg, country: UK fumonisins (no specification) incidence: 3/3, conc. range: 3.6-2600 μ g / kg, Ø conc.: 869 μ g / kg, country: Germany \rightarrow ochratoxin A incidence: 1/4, conc.: $< 5 \mu g / kg$, country: UK \rightarrow barley grits, \rightarrow rye grits, \rightarrow wheat grits **Maize malt** may contain the following → mycotoxins:

aflatoxin (\rightarrow aflatoxins)

incidence: 1/13, conc.: 1.71 μ g/kg, country: Zambia \rightarrow zearalenone incidence: nc/13, conc. range: 800-4000 μ g/kg, Ø conc.: 680 μ g/kg, country: Zambia \rightarrow barley malt

Maize meal For the US-market it could be shown that maize meal may contain mean levels up to and above 1 mg/kg \rightarrow fumonisin B₁ while other maize products e.g. \rightarrow maize grits usually show a lower contamination. Maize meal spiked with \rightarrow fumonisins was completely free of fumonisins after heating to 220 °C for 25 min. Maize meal may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: nc/4, conc. range: 0.2-0.7 μ g / kg, country: UK aflatoxins incidence: 1/2*, conc.: 129 µg/kg, country: USA, *imported \rightarrow deoxynivalenol incidence: 45/50, conc. range: 0-250 μg/kg, country: USA incidence: nc, \emptyset conc.: 110 µg/kg, country: Canada incidence: 2/2*, conc. range: 500-870 μ g/kg, Ø conc.: 685 μ g/kg, country: Germany, *organic product fumonisin B₁ incidence: nc/3, conc. range: < 50-1150 μg/kg, country: Austria incidence: 5/5, conc. range: 180-450 μg/kg, country: Botswana incidence: nc/15, conc. range: < 50-210 μg/kg, country: Bulgaria incidence: 1/2, conc.: 50 µg/kg, country: Canada incidence: 18/53*, conc. range: < 100-3500 μ g/kg, Ø conc.: 530 μ g/kg, country: Canada *and semolina

incidence: 2/2, conc. range: 1780-2980 μ g / kg, Ø conc.: 2380 μ g / kg, country: Egypt incidence: 1/1, conc.: 1240 µg/kg, country: France incidence: nc/3, conc. range: < 50-110 μg/kg, country: Kenya incidence: 1/2, conc.: 660 µg/kg, country: Peru incidence: 46/52, conc. range: < 50-475 μ g / kg, Ø conc.: 138 μ g / kg, country: South Africa incidence: nc/81, conc. range: 0-3900 μ g / kg, Ø conc.: 200 μ g / kg (all samples), country: South Africa incidence: nc/127, conc. range: 0-2850 μ g / kg, Ø conc.: 290 μ g / kg (all samples), country: South Africa incidence: 2/7, conc. range: 0-110 µg/kg, \emptyset conc.: 85 µg/kg, country: Switzerland incidence: 27/27*, conc. range: < 10-2200 μ g / kg, Ø conc.: 260 μ g / kg, country: Switzerland, *and grits incidence: 15/16, conc. range: < 50-2790 μ g / kg, Ø conc.: 1048 μ g / kg, country: USA incidence: 2/2*, conc. range: ca. 210-360 μ g / kg, Ø conc.: 290 μ g / kg, country: USA incidence: 3/3*, conc. range: 600-1200 μ g/kg, Ø conc.: 800 μ g/kg, country: USA incidence: 10/12*, conc. range: < 100-1200 μg/kg, Ø conc.: 550 μg/kg, country: USA, *maize meal, white incidence: 4/4*, conc. range: ca. 560-840 μg/kg, country: USA incidence: 3/3*, conc. range: 500-1000 μ g/kg, Ø conc.: 700 μ g/kg, country: USA incidence: 5/6*, conc. range: < 100-1710 μ g / kg, Ø conc.: 980 μ g / kg, country: USA, *maize meal yellow incidence: 10/13, conc. range: 430-2050 μ g / kg, Ø conc.: 848 μ g / kg, country: USA

incidence: 11/13, conc. range: < 80-2800 μ g / kg, Ø conc.: 970 μ g / kg (all samples), country: USA incidence: 16/16, conc. range: 280-2050 μ g / kg, Ø conc.: 860 μ g / kg, country: USA incidence: 1/1*, conc.: 2850 µg/kg, country: USA, *maize meal white, self rising incidence: 5/5*, conc. range: 400-1300 μ g / kg, Ø conc.: 890 μ g / kg, country: USA, *maize meal yellow incidence: 1/1*, conc.: 6320 µg/kg, country: USA, *maize meal blue incidence: 1/1*, conc.: 40 µg/kg, country: Venezuela, *maize meal white incidence: 4/4, conc. range: 70-1880 μ g / kg, Ø conc.: 718 μ g / kg, country: Venezuela incidence: 3/3, conc. range: 1060-3630 μg/kg, country: Zimbabwe \rightarrow fumonisin B₂ incidence: 5/5, conc. range: < 50-120 μg/kg, country: Botswana incidence: nc/15, conc. range: 50-150 μg/kg, country: Bulgaria incidence: 2/2, conc. range: 470-780 μ g/kg, Ø conc.: 625 μ g/kg, country: Egypt incidence: 1/1, conc.: 390 μg/kg, country: France incidence: nc/3, conc. range: < 50-140 μg/kg, country: Kenya incidence: 1/2, conc.: 135 µg/kg, country: Peru incidence: 11/52, conc. range: < 50-131 μ g / kg, Ø conc.: 83 μ g / kg, country: South Africa incidence: nc/81, conc. range: 0-760 μ g / kg, Ø conc.: 100 μ g / kg (all samples), country: South Africa incidence: nc/127, conc. range: 0-910 μ g / kg, Ø conc.: 70 μ g / kg (all samples), country: South Africa incidence: nc/27*, conc. range: < 10-590 μ g / kg, Ø conc.: 60 μ g / kg, country: Switzerland, *and grits

Maize meal

incidence: 13/16, con. range: 0-920 μ g / kg, Ø conc.: 298 μ g / kg, country: USA incidence: 2/2*, conc. range: ca. 33-58 μ g / kg, Ø conc.: 40.5 μ g / kg, country: USA incidence: nc/12*, conc. range: < 100-520 μ g / kg, Ø conc.: 210 μ g / kg, country: USA, *maize meal white incidence: 4/4*, conc. range: ca. 120-414 μg/kg, country: USA incidence: nc/6*, conc. range: < 100-470 μ g / kg, Ø conc.: 380 μ g / kg, country: USA, *maize meal yellow incidence: 10/13, conc. range: 50-360 μ g / kg, Ø conc.: 192 μ g / kg, country: USA incidence: nc/13, conc. range: < 100-1000 μ g / kg, Ø conc.: 300 μ g / kg (all samples), country: USA incidence: nc/16, conc. range: 50-530 μ g / kg, Ø conc.: 200 μ g / kg, country: USA incidence: $1/1^*$, conc.: 10 µg/kg, country: Venezuela, *maize meal white incidence: 4/4, conc. range: 20-530 μ g / kg, Ø conc.: 198 μ g / kg, country: Venezuela incidence: nc/3, conc. range: 240-910 μg / kg, country: Zimbabwe \rightarrow fumonisin B₃ incidence: 5/5, conc. range: < 50-120 μg/kg, country: Botswana incidence: 1/1, conc.: 180 μ g/kg, country: France incidence: nc/81, conc. range: 0-150 μ g / kg, Ø conc.: < 100 μ g / kg (all samples), country: South Africa incidence: nc/127, conc. range: 0-460 μ g / kg, Ø conc.: 30 μ g / kg (all samples), country: South Africa incidence: 2/2*, conc. range: "present", country: USA, *maize meal white incidence: 4/4*, conc. range: "present", country: USA, *maize meal yellow incidence: nc/3, conc. range: 130-230 μg/kg, country: Zimbabwe

fumonisins (FB₁, FB₂) incidence: 1/12, conc.: < 1000 µg/kg, country: UK fumonisins incidence: 1/2, conc.: 7.1 µg/kg, country: Germany incidence: 1/1, conc.: 1300 µg/kg (HPLC), country: Germany incidence: 3/3, conc. range: 1500-4700 μ g / kg, Ø conc.: 2933 μ g / kg, country: USA incidence: 2/2*, conc. range: no exact data, country: USA, *maize meal, blue incidence: 7/12*, conc. range: no exact data, country: USA, *maize meal yellow incidence: 6/6*, conc. range: 450-4750 μ g / kg, Ø conc.: 1558 μ g / kg, country: USA *maize meal yellow incidence: 4/4*, conc. range: 650-7450 μ g/kg, Ø conc.: 3075 μ g/kg, country: USA, *maize meal white \rightarrow moniliformin incidence: 27/27, conc. range: 50-180 μ g/kg, Ø conc.: 85.6 μ g/kg, country: France, UK, USA \rightarrow zearalenone incidence: 2/2*, conc. range: 38-65 μ g/kg, Ø conc.: 56.5 μ g/kg, country: Germany, *organic product incidence: 12/50, conc. range: nc, country: Mexico incidence: 9/11, conc. range: 11-69 μ g/kg, Ø conc.: 33.1 μ g/kg, country: USA incidence: 7/9, conc. range: 3.2-120 μ g / kg, Ø conc.: 23 μ g / kg, country: USA \rightarrow sorghum meal

Maize muffin may contain the following \rightarrow mycotoxins: \rightarrow fumonisins incidence: 1/1, conc.: 300 µg/kg, country: USA

Maize pop cereal may contain the following \rightarrow mycotoxins:

 \rightarrow fumonisins (no specification) incidence: 1/1, conc.: 200 µg/kg, country: USA

Maize products (no specification) Fumonisin concentration in refined \rightarrow maize products may be lowered during the process of \rightarrow milling. Maize products may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins incidence: 1/23, conc.: 37 µg/kg, country: Japan incidence: 15/41, conc.: > $30 - \leq 400$ µg/kg, country: Philippines incidence: 19/139, conc. range: \leq 53 μ g / kg, Ø conc.: 19.6 μ g / kg, country: USA \rightarrow fumonisin B₁ incidence: 5/71, conc. range: < 100-1200 μ g/kg, Ø conc.: 330 μ g/kg, country: Canada incidence: 2/2, conc. range: 1780-2980 μ g / kg, Ø conc.: 2380 μ g / kg, country: Egypt incidence: 5/8, conc. range: < 10-60 μ g / kg, Ø conc.: 50 μ g / kg, country: Italy incidence: nc/4, conc. range: 0-660 μ g / kg, Ø conc.: 165 μ g / kg, country: Peru incidence: 2/6, conc. range: 41.4-73 μ g / kg, Ø conc.: 57.2 μ g / kg, country: Thailand incidence: 2/8, conc. range: 0-91 µg/kg, Ø conc.: 84 μ g/kg, country: South Africa incidence: nc/68, conc. range: 0-475 μ g/kg, Ø conc: 105 μ g/kg, country: South Africa incidence: 2/20, conc. range: 60-200 μ g/kg, Ø conc.: 130 μ g/kg, country: Spain incidence: 4/4, conc. range: 85-700 μ g / kg, Ø conc.: 409 μ g / kg, country: USA incidence: 4/4, conc. range: 20-320 μ g / kg, Ø conc.: 170 μ g / kg, country: USA

incidence: 4/9, conc. range: < 10-120 μg / kg, Ø conc.: 70 μ g/kg, country: USA incidence: nc/29, conc. range: 0-2790 μ g/kg, Ø conc.: 711 μ g/kg, country: USA incidence: 3/5, conc. range: < 50-1210 μ g / kg, Ø conc.: 540 μ g / kg, country: USA \rightarrow fumonisin B₂ incidence: 2/2, conc. range: 410-780 μ g / kg, Ø conc.: 595 μ g / kg, country: Egypt incidence: nc/8, conc. range: < 10-20 μg/kg, country: Italy incidence: nc/4, conc. range: 0-135 μ g/kg, Ø conc.: 34 μ g/kg, country: Peru incidence: nc/68, conc. range: 0-120 μ g/kg, Ø conc: 21 μ g/kg, country: South Africa incidence: 3/4, conc. range: 0-240 µg/kg, Ø conc.: 148 µg/kg, country: USA incidence: nc/9, conc. range: 10-30 μ g / kg, Ø conc.: 20 μ g / kg, country: USA incidence: nc/29, conc. range: 0-2790 μ g / kg, Ø conc.: 711 μ g / kg, country: USA \rightarrow ochratoxin A incidence: 1/23, conc.: 73 µg/kg, country: Japan

Maize screenings (Syn.: corn screenings) Compared to the intact corn fumonisin levels in \rightarrow maize screenings can be about 10 times higher (Iowa corn). It seems that there is no size-related segregation of fumonisin contents in corn screenings. The accumulation of \rightarrow fumonisins in corn screenings may be a source of concern since they are used in feed formulas for livestock.

Maize snacks may contain the following \rightarrow mycotoxins: \rightarrow fumonisin B₁ incidence: 2/11, conc. range: 50-200

μg/kg, country: Spain

Maize snacks

incidence: 26/78, conc. range: \leq 2395 µg/kg, Ø conc.: 456 µg/kg, country: Thailand \rightarrow fumonisin B₂ incidence: 16/78, conc. range: \leq 715 µg/kg, Ø conc.: 145 µg/kg, country: Thailand \rightarrow fumonisins (FB₁, FB₂, FB₃) incidence: 31/40, conc. range: 11-220 µg/kg, Ø conc.: 46 µg/kg, country: UK

Maize starch may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin incidence: 6/9, conc. range: $\leq 25 \ \mu\text{g}/\text{kg}$, \emptyset conc.: 12 $\mu\text{g}/\text{kg}$, country: Philippines \rightarrow fumonisins (no specification) incidence: 1/1, conc.: 500 $\mu\text{g}/\text{kg}$, country: USA

Majoran may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A incidence: 1/1, conc.: 28 µg/kg, country: Austria \rightarrow spices

Malaga \rightarrow Wine

Malt (malting)

The mycotoxin contamination of malt with e.g. \rightarrow deoxynivalenol, \rightarrow nivalenol and/or \rightarrow zearalenone is due to the use of natural contaminated \rightarrow grains and/or growth of certain fungi during various stages of the malting production. \rightarrow Mycotoxins may impair malt processing. \rightarrow T-2 toxin, added before malting, inhibited coleoptile and rootlet elongation in germinating acid-dehusked \rightarrow barley depending on the concentration used. To some extent this mycotoxin also retarded de novo synthesis of α -amylase. \rightarrow Diacetoxyscirpenol and deoxynivalenol act in the same way.

The apparent loss of zearal enone (\approx 75%) and T-2 toxin ($\approx\,$ 54%) during

malting might be due to the binding of the mycotoxins to substances extracted from barley into steep liquor and present in kilned malt or might be caused by binding to microoganisms or degradation products of malt sugars. DON losses amounted up to almost 80%. Since substantial to total losses of \rightarrow ochratoxin A and \rightarrow citrinin during malting have been reported, it is concluded that neither OTA nor citrinin are likely to enter the brewing process from malted barley but from brewing adjuncts (OTA). Malt may contain the following mycotoxins: ochratoxin A incidence: 3/11*, conc. range: 0.1-0.92 μg/kg, country: Germany, *partly imported incidence: 1/2, conc. range: 1.5-9.99 μg/kg, country: Germany \rightarrow beer

Mandarin fruits may contain the following \rightarrow mycotoxins: \rightarrow alternariol incidence: 2/3*, conc. range: 1000-5200 $\mu g \, / \, kg, \, {\ensuremath{\not Q}} \,$ conc: 3100 $\mu g \, / \, kg,$ country: Italy \rightarrow alternariol methyl ether incidence: 2/3*, conc. range: 550-1400 μ g / kg, Ø conc.: 975 μ g / kg, country: Italy \rightarrow tenuazonic acid incidence: 3/3*, conc. range: 21,000-173,900 μg/kg, Ø conc.: 94,033 μg/kg, country: Italy *samples visibly affected by \rightarrow Alternaria rot \rightarrow fruits Mango (pickled in salt) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁

incidence: 4/8*, Ø conc.: 52 μg/kg, coun-

try: India

incidence: 26/40**, Ø conc.: 210 μg/kg, country: India \rightarrow aflatoxin B₂ incidence: 4/8*, Ø conc.: 5 µg/kg, country: India incidence: $26/40^{**}$, Ø conc.: $32 \mu g/kg$, country: India \rightarrow aflatoxin G₁ incidence: 4/8*, Ø conc.: 24 µg/kg, country: India incidence: 26/40**, Ø conc.: 184 μg/kg, country: India \rightarrow aflatoxin G₂ incidence: $4/8^*$, \emptyset conc.: traces, country: India incidence: 26/40**, Ø conc.: 15 μg/kg, country: India * stored in bottles, **stored in polythene bags \rightarrow fruits

Manioc may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins (no specification) incidence: 1/8, conc.: nc, country: Mocambique

Marchpane → marzipan

Marzipan (almond paste) Blanched \rightarrow almonds for marzipan manufacture should be processed immediately after blanching. If the period of storage prior to blending with sugar and drying is too long, fungal infection may occur with subsequent aflatoxin contamination. 3 days of storage at 28 °C are almost critical. Marzipan may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/168, conc.: 39 µg/kg, country: Finland incidence: 3/12, conc. range: tr-2 µg/kg, country: Germany \rightarrow aflatoxin B₂

incidence: 1/168, conc.: 7 μ g/kg, country: Finland incidence: 1/16, conc.: < 1 μ g/kg, country: Germany \rightarrow aflatoxin G₁ incidence: 1/12, conc.: traces, country: Germany \rightarrow almonds, \rightarrow persipan

is tortilla \rightarrow flour which has tradi-Masa tionally been treated with Ca(OH)₂ and heat (nixtamalization). This processing, which improves the nutritive value of \rightarrow maize, may decontaminate fumonisincontaminated maize because the fumonisin levels in \rightarrow maize products made with masa usually are low. Masa may contain the following \rightarrow mycotoxins: \rightarrow fumonisin B₁ incidence: 2/3, conc. range: 590-1800 μ g / kg, Ø conc.: 1195 μ g / kg, country: Mexico incidence: 3/3, conc. range: 40-380 μ g/kg, Ø conc.: 163 μ g/kg, country: USA incidence: 8/8, conc. range: 63-689 μ g/kg, Ø conc.: 262 μ g/kg, country: USA / Mexico \rightarrow fumonisin B₂ incidence: 2/3, conc. range: 110-1380 μ g / kg, Ø conc.: 553 μ g / kg, country: Mexico incidence: 1/3, conc.: 60 µg/kg, country: USA hydrolyzed fumonisin B_1 (HBF₁) incidence: 1/2, conc.: 100 μ g/kg, country: Mexico incidence: 2/3, conc. range: 20-100 μ g / kg, Ø conc.: 60 μ g / kg, country: USA incidence: 6/8, conc. range: 21-178 μ g/kg, Ø conc.: 64 μ g/kg, country: USA/Mexico \rightarrow maize

Meat and meat products

Probably there is little or no danger in the transmission of \rightarrow mycotoxins into the muscle tissues of most animals consuming feed contaminated with mycotoxins. This is due to the fact that the transfer ratios are obviously high; the transfer ratio for \rightarrow aflatoxin B₁ (µg/kg mycotoxin in feed: $\mu g / kg$ mycotoxin in tissue) is in the range of 1000-14,000. In general, it is expected that animals exposed to such high mycotoxin concentrations suffer from obvious disease symptoms or even die. It is most unlikely that such animals enter the food chain and therefore animal tissues do not contribute substantially to mycotoxin intake of humans. However, \rightarrow ochratoxin A in kidneys, \rightarrow sausages, and black pudding prepared from pigs may represent an exception. Feeding experiments with aflatoxin B_1 , ochratoxin A, \rightarrow patulin, \rightarrow penicillic acid, \rightarrow sterigmatocystin, \rightarrow T-2 toxin, and \rightarrow zearalenone labeled with radioactive elements show a major excretion rate for the mycotoxins and / or their metabolites within 24 h. Only minor levels could be detected in the edible tissue. Extensive breakdown of the mycotoxins is not expected since a negligible amount of radioactivity could be detected in the expired air. Elaborated data suggest that the structure modifications are due to mixed-function oxidases and the high specific activity implies that the liver and biliary system is mainly responsible for the elimination process.

The only two mycotoxins that might be important in domestic animals are aflatoxin B_1 (\rightarrow milk/ \rightarrow aflatoxin M_1) in cows and ochratoxin A in liver, kidneys and meat products, e.g. certain sausages of pigs. There are three possible ways of mycotoxin contamination of meat/meat products:

i) Mold growth on the product surface leading to direct mycotoxin contamination which is of minor importance. Contamination rate of fermented meat products such as salamis or country cured \rightarrow ham with the most dangerous \rightarrow Aspergillus flavus Link and \rightarrow Aspergillus parasiticus Speare is rare. If present these molds must successfully compete against starter cultures (e.g. *Penicillium nalgiovense*) or the house flora. In addition, relative humidities < 80%, temperatures between 10 and 15 °C, as well as smoking and curing ingredients prevent or reduce aflatoxin production. It is not expected that human exposure to \rightarrow aflatoxins by this route is of great importance.

ii) Meat products may contain mycotoxins due to the use of naturally-contaminated \rightarrow spices and spice mixtures (see \rightarrow meat, luncheon)

iii) However, transmission (\rightarrow carry-over) of mycotoxins by the intake of contaminated feedstuff, especially in the case of OTA, is more serious. A period of approximately four weeks is necessary to reduce OTA to nondetectable levels (< 2 μ g/kg) in swine kidneys.

Among the domestic animals ruminants, e.g. cattle, are not very susceptible to ochratoxin A. Since OTA is a phenylalanine amide of isocoumarin the rumen flora decomposes the mycotoxin by proteolytic enzymes. However, pigs are extremely sensitive to OTA which possesses a relatively high serum half-live of 72-120 h. Blood / plasma contains the highest OTA concentrations followed by kidneys, liver, muscles, fat (in decreasing order). On average, OTA levels in lean muscle is half of that in the kidney. OTA levels in e.g. raw sausages, liver sausagetype, Frankfurter type sausage, ham, bacon depend on the particular recipe of the meat product (proportion of pork, beef, etc.). If liver tissue or blood is used, OTA concentrations greatly increase in the corresponding meat products, such as Bologna-type sausages, blood sausages or black pudding. In contrast, pork as well as poultry meat contain, if at all, low levels of OTA and therefore do not constitute a significant health problem for humans (see Figure Meat). The stability of OTA prevents its reduction during the stages of meat-product manufacturing, such as heating and ripening as well as storage. Only frying or boiling lowered the OTA concentration by as much as 40% (blood-pudding, kidneys, muscular tissues) depending on the water content and the inner temperature of the treated product. No OTA losses occurred in adipose tissues. Compared to food of plant origin, meat and meat products usually show lower levels of OTA contamination.

Contamination problems with the milk aflatoxin M1 arise because feed consumption and lactation are concurrent events without any withdrawal period. However, the four "primary" aflatoxins B_1 , B_2 , G_1 , and G₂ are rapidly metabolized. In consequence, none of them or only low levels are found in animal tissues or milk. In addition, despite the toxicity of AFM₁, the macromolecule-bound AFB1 derivatives in meat are at least 4000 times less active than AFB₁. The water-soluble conjugates are at least 100 times less potent compared to AFB₁. From the present data it is concluded that there is a negligible carcinogenic risk for humans who consume aflatoxin contaminated liver or meat compared with certain foodstuffs of plant origin, e.g. \rightarrow nuts. According to Frisvad (1988) the following mycotoxins may be found in meat (and eggs): aflatoxins, \rightarrow citrinin, \rightarrow cyclopia-

zonic acid, ochratoxin A, patulin, \rightarrow penicillic acid, penitrem A (\rightarrow penitrems), \rightarrow rugulosin, \rightarrow sterigmatocystin, \rightarrow vio-

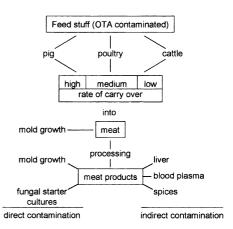
mellein, \rightarrow xanthomegnin.

Meat and meat products may contain the following mycotoxins:

 \rightarrow aflatoxins (no specification) incidence: 2/19, conc. range: < 1 µg/kg, country: UK → citrinin incidence: 9/23, conc. range: < 100 µg/kg, country: UK → ochratoxin A incidence: 7/33, conc. range: 0-4 µg/kg, country: UK incidence: 6/6*, conc. range: 0.1-2.2 µg/kg, country: Tunesia, *and fish (mackerel) → patulin incidence: 7/24, conc. range: 0-200 µg/kg, country: UK

Meat, luncheon Detection of \rightarrow aflatoxins in luncheon \rightarrow meat results from the use of mycotoxin contaminated \rightarrow spices and/or the incorporation of aflatoxin producers. Luncheon meat may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/25, conc.: 4 µg/kg, country: Egypt \rightarrow aflatoxin B₂ incidence: 1/25, conc.: 2 µg/kg, country: Egypt

Mechanical damage of the seed or fruit coat favors the penetration of molds into \rightarrow grains / \rightarrow fruits as a prerequisite for



Meat. Direct and indirect OTA contamination of meat and meat products

mycotoxin (\rightarrow mycotoxins) contamination.

Melon may contain the following \rightarrow mycotoxins: \rightarrow alternariol methyl ether incidence: 1/1*, conc.: 51 µg/kg, country: Italy *sample visibly affected by \rightarrow Alternaria rot \rightarrow fruits

Melon balls snacks may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins incidence: 4*/40, conc. range: 10-40 μ g/kg, country: Nigeria *all samples contained \rightarrow aflatoxin B₁; 2 sa additionally \rightarrow aflatoxin B₂ and / or \rightarrow aflatoxin G₁

Melon seeds may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins (no specification) incidence: 2/4*, conc. range: $\leq 29 \ \mu g/kg$, \emptyset conc.: 26 $\mu g/kg$, country: USA, *imported

Microbial interactions The presence of competing fungi / bacteria and their effects on toxin production are not predictable. In general, development and mycotoxin formation of \rightarrow Aspergillus spp. and \rightarrow Penicillium spp. is considerably reduced if other competing microorganisms are present.

Milk, camel may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 6/20, conc. range: 0.25-0.8 µg/l, country: UAE

Milk, cow Of all animal products milk, one of the best natural foods and the primary nutrient for children, including infants, is most frequently contaminated

with aflatoxin residues. Therefore, a theoretical hazard associated with \rightarrow aflatoxin M₁ in commercially available milk, \rightarrow human breast milk, and milk products does exist mainly because the growing young are very susceptible to the adverse effects of \rightarrow mycotoxins. They usually have a relatively low body weight, showing a high cell activity whereas the immune system is only partially developed. Due to the slower rate of biotransformation of carcinogens in infants a longer circulation time of the chemicals may be the consequence. In the early 1960s a toxic factor in milk was discovered after feeding lactating cows with aflatoxin-contaminated diet. The toxic factor named aflatoxin M occurred in milk 4-5 h after ingestion of the contaminated feed. Structure elucidation of the milk metabolite AFM₁ succeeded in 1966. Subsequent studies revealed that AFM_1 is the major aflatoxin in milk although other hydroxylated \rightarrow aflatoxin B_1 metabolites such as \rightarrow aflatoxin M_2 , \rightarrow aflatoxin M₄, \rightarrow aflatoxin Q₁, and \rightarrow aflatoxicol have been detected. However, these aflatoxin derivatives occur in very low concentrations (two to three orders of magnitude lower compared to AFM₁).

Transmission of other \rightarrow mycotoxins such as \rightarrow deoxynivalenol, \rightarrow fumonisins, \rightarrow ochratoxin A, \rightarrow sterigmatocystin, \rightarrow T-2 toxin, and \rightarrow zearalenone in milk and \rightarrow milk products has been studied / detected. In the case of \rightarrow trichothecenes relatively high concentrations have to be ingested to produce detectable toxin residues in milk. Compared to DON marginally more T-2 toxin appears to be transmitted into milk probably due to its lipophilic nature. However, in cattle this mycotoxin is extensively and very rapidly metabolized. Instead of relatively high oral doses (0.5-3.6 mg/kg) no substantial accumulation of any residues in milk, organs or tissues could be observed.

Their low toxicity and / or limited presence in milk probably makes these mycotoxins of little significance for human health.

Contaminated concentrate feeds are mainly responsible for aflatoxin contamination in milk and dairy products. The increased AFM₁ contamination rate in milk during the winter months was due to the major usage of compound feeds in the cold season imported from tropical and subtropical countries. In contrast, in spring and summer time home grown non contaminated roughage, summer forage, and pasture are available. Strict control measures for locally manufactured and imported feedstuffs are a prerequisite of low aflatoxin levels in milk and \rightarrow dairy products. In this connection the Commission of the European Communities further tightened the acceptable level for AFB₁ in feedstuff in dairy cattle from 20 to 10 μ g/kg in 1984 to 5 μ g/kg in 1991. To prevent AFM₁ contamination in milk feeding of \rightarrow peanuts which are frequently contaminated by AFB₁ to lactating cows has been forbidden by the Swiss legislation. Transmission rate (\rightarrow carry over) of aflatoxin B_1 that is ingested in the feed and excreted as the 4-hydroxylated derivative AFM₁ in milk varies among individual animals but is linearly correlated with milk yield and roughly amounted 1.5% (0.35-3%), e.g. 300 μ g aflatoxin B₁/kg feed will result in $\approx 4.5 \,\mu g$ aflatoxin $M_1/1$ milk. A rapid increase in aflatoxin concentration was observed when a high intake of the mycotoxin reduced the milk yield.

Metabolization of AFB_1 is due to the hepatic microsomal mixed-function oxidase system, but, depending on species, several other metabolic conversions are possible (e.g. metabolism rate in the liver, excretion rate by other routes). AFM_1 is found in cow's milk as early as 4 h after ingestion of the contaminated feed. A significant decrease in aflatoxin concentration occurs 24-48 h after exposition. 3 to 5 days after aflatoxin-free rations are given aflatoxin values decreased to zero in the milk.

Heating, e.g. pasteurization (\rightarrow milk, pasteurized) or sterilization (\rightarrow milk, sterilized), does not cause any destruction of the thermoresistant AFM₁ molecule in milk although different results have been reported (63% pasteurization, 80% sterilization). Data about the reduction of AFM₁ concentration in cold treated or frozen milk are contradictory and not conclusive. AFM₁ contamination of (processed) milk indicates the level of AFB₁ in animal feed.

There is no homogeneous distribution of AFM_1 in milk. Since the semipolar AFM_1 is primarily bound to casein it is estimated that about 30% of AFM1 are associated with the nonfat milk solids. The enrichment of AFM₁ in the nonfat fraction resulted from processes which involve fat (\rightarrow cream) separation. When butter is made from naturally contaminated cream, the AFM₁ concentration in the butter amounted to a little more than 20%, while the major portion of AFM_1 is found in buttermilk (\rightarrow milk-, butter). Skim-milk manufacturing may lead to the accumulation of about 80% of AFM₁ in that portion. Lower levels of AFM₁ (60-75%) may be found in concentrated milk. No AFM₁ reduction was observed during the manufaturing of cheese and yogurt.

Although aflatoxin B_1 is also a contaminant of milk very much lower levels compared to AFM₁ have been found. Compared to raw farm milk, the rate of AFM₁ contamination in commercial milk is often higher, resulting from the addition of a few contaminated samples to uncontaminated bulk milk. However, for the same reason commercial milk in general shows low levels of AFM₁ contamination.

Detectable levels of OTA in cow's milk only result from a daily dose of OTA higher than 1.66 mg/kg bw. Lower amounts did not lead to the detection of OTA in milk mainly due to the hydrolysis of this mycotoxin (ochratoxin α) by the microflora in the rumen of the cow. It seems that milk is not an important contributor to OTA intake but taking into account that e.g. children consume large quantities of milk, even low OTA levels $(0.01-0.04 \ \mu g/l \ milk)$ might significantly increase the daily OTA intake. Milk may contain the following mycotoxins: aflatoxin B₁ incidence: 2/1150, conc. range: 0.28-0.36 μg/l, country: Spain incidence: 5/105, conc. range: ≤ 2500 μg/l, country: Yugoslavia aflatoxin M₁ incidence: 9/12*, conc. range: 0.002-0.05 μg/l, country: Austria, *raw incidence: 32/88*, conc. range: 0.001-0.01 μg/l, country: Austria, *commercial incidence: 42/68, conc. range: 0.02-0.2 μg/l, country: Belgium incidence: 46/145, conc. range: < 0.02-0.5 μ g / l, country: Belgium incidence: 3/6, conc. range: 0.025-0.5 μg/l, country: Brazil incidence: 4/224, conc. range: tr-0.002 μg/l, country: Brazil incidence: 1/100*, conc.: 0.2 µg/l, country: Brazil, *commercial incidence: 9/50*, conc. range: 0.1-1.68 μg/l, country: Brazil, *farm incidence: 22/85, conc. range: > 0.5 μ g/l, country: Cuba incidence: 5/77*, conc. range: tr-0.38 μg/l, country: Czechoslovakia, *raw incidence: 27/89*, conc. range: $< 0.5 \mu g/l$, country: Czechoslovakia, *raw incidence: 25/191, conc. range: 0.05-0.1 µg/l, country: Czechoslovakia incidence: 9/67, conc. range: 0.05-0.1 µg/l, country: Czechoslovakia

incidence: 43/403, conc. range: 0.025-0.1 μ g/l (37 samples), 0.1-0.5 μ g/l (6 sa), country: Czechoslovakia inicdence: 46/376, conc. range: 0.025-0.1 $\mu g/l$ (44 samples), > 0.1 $\mu g/l$ (2 sa), country: Czechoslovakia incidence: 9/117*, conc. range: 0.05-0.1 μg/l, country: Czechoslovakia, *commercial incidence: 11/88, conc. range: < 0.001-0.023 µg/l, country: France incidence: 168/380, conc. range: 0.05-1.15 µg/l, country: France incidence: 32/102, conc. range: 0.5-5 μg/l, country: France incidence: 5489/5489, conc. range: 0-0.05 μg/l (5.284 samples), 0.05-0.5 μg/l (200 sa), > 0.5 µg/l (5 sa), country: France incidence: 757/757, conc. range: 0-0.05 μ g/l (659 samples), 0.05-0.5 μ g/l (84 sa), $> 0.5 \ \mu g/l$ (14 sa), country: France incidence: 70/112, conc. range: < 0.01-16.1 μg/l, country: France incidence: 31/225, conc. range: < 0.001-0.01 µg/l, country: Germany incidence: 16/25, conc. range: 0.04-0.13 μ g/l, country: Germany incidence: 21/48, conc. range: 0.04-0.25 μ g/l, country: Germany incidence: 7/13, conc. range: 0.05-0.13 μg/l, country: Germany incidence: 79/419, conc. range: 0.05-0.54 μ g/l, Ø conc.: 0.12 μ g/l, country: Germany incidence: 118/260, conc. range: 0.05-0.33 μg/l, country: Germany incidence: 4/60, conc. range: 1.7-6.5 μ g/l, \emptyset conc.: 3.6 µg/l, country: Germany incidence: 265/279, conc. range: 0.0003-0.68 μg/l, country: Germany incidence: 624/6445, conc. range: 0.01- $> 0.05 \ \mu g / l$, country: Germany incidence: 1507/1507, conc. range: 0-0.05 $\mu g/l$ (1504 samples), > 0.05 $\mu g/l$ (3 sa), country: Germany

incidence: 388/388, conc. range: 0-0.01 $\mu g/l$ (387 samples), > 0.01 $\mu g/l$ (1 sa) country: Germany incidence: 28/61, conc. range: 0.04-0.25 μg/l, country: Germany incidence: 4/36, conc. range: 1.7-6.5 µg/l, \emptyset conc.: 3.6 µg/l, country: Germany incidence: 4/99*, conc. range: 0.1-0.13 μg/l, country: Greece, *raw incidence: 72/81, conc. range: 0.0005-0.001 µg/l (31 samples), 0.0025-0.005 $\mu g/l$ (32 sa), > 0.005-0.177 $\mu g/l$ (9 sa)., country: Greece incidence: 89/504, conc. range: 0.1-3.5 μg/l, country: India incidence: 3/21, conc. range: \leq 13.3 µg/l, Ø conc.: 1159 µg/l, country: India incidence: 48/52, conc. range: \leq 23 µg/l, country: Iran incidence: 38*/95, conc. range: 8-500 μ g/l, country: Iran, *mainly AFM₁ and to a minor degree AFM₂ incidence: 12/18, conc. range: 0.005-0.03 μ g/l, country: Italy incidence: 76/106, conc. range: 0.004-0.28 μ g / l, country: Italy incidence: 34/82, conc. range: tr-0.569 μ g/l, country: Italy incidence: 46/59, conc. range: tr-0.378 μ g / l, country: Italy incidence: 24/27, conc. range: 0.005-0.065 μg/l, country: Italy incidence: 136/159, conc. range: < 0.001-0.1 μ g/l, Ø conc.: 0.01 μ g/l, country: Italy incidence: 5/31, conc. range: 0.03-0.07 μ g/l, country: Italy incidence: 9/52*, conc. range: 0.005-0.146 μg/l, country: Italy, *raw incidence: 8/31*, conc. range: 0.005-0.091 μg/l, country: Italy, *raw incidence: 24/57*, conc. range: 0.3-0.93 μg/l, country: Italy, *raw incidence: 3/60*, conc. range: 0.1-0.28 μg/l, country: Italy, *raw incidence: 5/107, conc. range: 0.024-0.094 μ g/l, country: Italy

incidence: 66/107, conc. range: 0.006-0.101 µg/l, country: Italy incidence: 56/107, conc. range: 0.003-0.06 μ g / l, country: Italy incidence: 1/50, conc.: $0.4 \mu g/l$, country: Italy incidence: 19/22*, conc. range: 0.18-0.434 µg/l, country: Italy, *commercial incidence: 30/276*, conc. range: 0.01-0.2 μg/l, country: Italy, *raw incidence: 4/4, conc. range: 1.3-6.8 μ g/l, country: Norway incidence: 11/22, conc. range: 0.01-0.25 μ g / l, country: Poland incidence: 5/21, conc. range: 0.02-0.2 μ g/l, country: South Africa incidence: 14/47, conc. range: 0.02-0.1 μg/l, country: Spain incidence: 61/61*, conc. range: < 0.01 μ g/l (49 samples), 0.01-0.02 μ g/l (10 sa), $0.02-0.04 \ \mu g/l$ (2 sa), country: Spain, *raw incidence: 1/84, conc. range: 0.05-0.1 μ g/l, country: Sweden incidence: 13/13, conc. range: 0.005-0.36 µg/l, country: Sweden incidence: 16/163, conc. range: 0.05-2 μ g / l, country: Switzerland incidence: 40/230, conc. range: 0.05-3 μg/l, country: Switzerland incidence: 8/91, conc. range: 0.001-0.609 μg/l, country: Switzerland incidence: 2/38, conc. range: 0.01-0.05 μg/l, country: Switzerland incidence: 84/105*, conc. range: 0.015-0.09 μ g/l, country: The Netherlands, *and UHT incidence: 74/95, conc. range: < 0.09-0.5 μ g/l, country: The Netherlands incidence: 85/278, conc. range: 0.03-0.52 μ g/l, country: UK incidence: 24/409, conc. range: 0.02-0.05 μ g/l (10 samples), 0.05-0.1 μ g/l (6 sa), $> 0.1 \ \mu g/l$ (8 sa), country: UK incidence: 7/22, conc. range: 0.2-0.5 $\mu g\,/\,l$ (6 samples), > 0.5 μ g/l (1 sa), country: Uruguay

Milk, cow

incidence: 192/302, conc. range: < 0.1 μ g/l (15 samples), 0.1-0.4 μ g/l (158 sa), 0.5-3.9 $\mu g/l$ (19 sa), country: USA incidence: 554/816, conc. range: 0.1-2 μ g/l, country: USA incidence: 116/912, conc. range: 0.1-2 μg/l, country: USA incidence: 144/624, conc. range: 0.1-2 μg/l, country: USA incidence: 107/847, conc. range: 0.1-2 μg/l, country: USA incidence: 235/786, conc. range: 0.1-2 μg/l, country: USA incidence: 99/168, conc. range: 0.1-2 μg/l, country: USA \rightarrow fumonisin B₁ incidence: 1/165, conc. 1.3 μ g/l, country: USA ochratoxin A incidence: 4/36, conc. range: 0.007-0.030 μg/l, country: Germany incidence: 9/50, conc. range: 1.7-6.6 µg/l, country: Italy incidence: 6/40*, conc. range: 0.011-0.058 μg/l, country: Norway, *conventional incidence: 5/47*, conc. range: 0.015-0.028 μg/l, country: Norway, *organic incidence: 5/36, conc. range: 0.01-0.04 μg/l, country: Sweden \rightarrow cheese, \rightarrow human breast milk

Milk (raw or dried, for infant formulae)

may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 46/376, conc. range: < 0.5 µg/ l, country: Czechoslovakia incidence: 2/376, conc. range: > 0.1 µg/l, country: Czechoslovakia incidence: 1/56, conc.: ca. 0.67 µg/kg, country: Germany incidence: 7/7, conc. range: 0.679-1.96 µg/l, country: Italy incidence: 58/233, conc. range: 0.2-0.8 µg/l, country: Italy

Milk (skim milk, dried) may contain the following \rightarrow mycotoxins:

→ aflatoxin M_1 incidence: 16/28, conc. range: 0.08-1.5 µg/l, country: Germany incidence: 8/93, conc. range: 0.05-0.4 µg/l, country: USA incidence: 17/27, conc. range: tr- > 2 µg/l, country: USA

Milk, pasteurized Pasteurization seems to have only a minor effect on inactivating \rightarrow aflatoxin M₁ in milk whereas sterilization will cause some losses of AFM₁. Pasteurized milk may contain the following \rightarrow mycotoxins: aflatoxin M₁ incidence: 4/204*, conc. range: 0.073-0.37 μ g/l, Ø conc.: 0.155 μ g/l, country: Brazil *includes pasteurized \rightarrow milk, \rightarrow milk powder and \rightarrow milk products incidence: 16/314, conc. range: $< 0.5 \ \mu g/$ l, country: Czechoslovakia incidence: 9/9, conc. range: $\leq 20.1 \, \mu g / l$, country: Iran incidence: 59/66, conc. range: 0.004-0.15 μ g / l, country: Italy incidence: 61/68, conc. range: 0.005-0.05 μ g/l, country: Italy incidence: 27/30, con. range: 0.003-0.022 μ g/l, country: Italy incidence: 7/143, conc. range: 0.1-0.4 μ g / l, country: Portugal incidence: 2/24, conc. range: 0.02-0.04 μg/l, country: Spain

Milk, sterilized Sterilization of milk will cause some losses in \rightarrow aflatoxin M₁ levels. Sterilized milk may contain the following \rightarrow mycotoxins: aflatoxin M₁ incidence: 5/33, conc. range: 0.01-0.04 µg/l, country: Spain

Milk, UHT may contain the following \rightarrow mycotoxins:

 \rightarrow aflatoxin M₁ incidence: 10/60, conc. range: 0.1-0.5 μ g / l, country: Italy incidence: 28/32, conc. range: 0.007-0.050 μ g / l, country: Italy incidence: 14/47, conc. range: 0.02-0.1 μ g / l, country: Italy incidence: 33/33, conc. range: < 0.010 μ g/l (28 samples), 0.01-0.025 μ g/l (5 sa), country: Spain incidence: 12/76*, conc. range: 0.02-0.04 µg/kg, country: Spain, *includes 24 semiskimmed samples, all not contaminated **Milk powder** The production of dry milk may lead to a decrease in \rightarrow aflatoxin M₁ concentration of about 85% compared to the raw milk. Milk powder may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 4/4*, conc. range: 320-5400 $\mu g^{**} / kg$, Ø conc.: 3193 $\mu g / kg$, country: Czechoslovakia, *leftover, ** in surface layers incidence: 5/5*, conc. range: 42-550 µg/kg, country: Czechoslovakia, *leftover incidence: 1/18, conc.: 6.4 µg/kg, country: Germany aflatoxin M₁ incidence: 8/210, conc. range: 0.2-nc μg/kg, country: Austria incidence: 468/837, conc. range: 0.03-0.69 μg/kg, country: Austria incidence: 33/300. conc. range: 0.1-1 μ g / kg, Ø conc.: 0.27 μ g / kg, country: Brazil incidence: 21/28, conc. range: 0.015-0.464 μ g/kg, Ø conc.: 0.1 μ g/kg, country: China incidence: 1/15, conc.: $15 \mu g/kg$, country: Denmark incidence: 129/222, conc. range: 0.050-5.2 μg/kg, country: France

incidence: nc/183, conc. range: ≤ 15.4 μ g / kg, Ø conc.: 1.79 μ g / kg, country: France incidence: nc/55, conc. range: ≤ 1.36 μ g / kg, Ø conc.: 0.225 μ g / kg, country: France incidence: 47/95, conc. range: 0.100-2.55 µg/kg, country: Germany incidence: 7/80, conc. range: 0.67-2 µg/kg, country: Germany incidence: 30/41, conc. range: 0.2-2 μ g/kg, Ø conc.: 0.5 μ g/kg, country: Germany incidence: 74/120, conc. range: 0.02-0.4 μg/kg, country: Germany incidence: 8/166, conc. range: 0.67-2.0 μg/kg, country: Germany incidence: 7/120, conc. range: 0.05-0.13 μg/kg, country: Germany incidence: 36/55*, conc. range: tr-4 μ g / kg, country: Germany, *27 samples of skim milk and 28 samples of whole milk powder incidence: 58/233, conc. range: 0.002-0.008 µg/kg, country: Italy incidence: 81/97, conc. range: < 0.001-0.1013 μg / kg, Ø conc.: 0.0218 μg / kg, country: Italy incidence: 4/21, conc. range: 0.030-0.25 μ g / kg, country: Italy incidence: 6/13, conc. range: 0.050-0.1 μg/kg, country: Italy incidence: 9/9, conc. range: 0.01-0.28 μ g / kg, country: Italy incidence: 10/10, conc. range: 0.015-0.1 μ g / kg, country: Italy incidence: 2/6, conc. range: 0.015-0.035 μ g / kg, country: Italy incidence: 3/18, conc. range: 0.040-0.090 μ g / kg, Ø conc.: 0.066 μ g / kg, country: Italy incidence: 3/12, conc. range: traces, country: Italy incidence: 3/3, conc. range: 0.015-0.085 μ g / kg, country: Poland incidence: 35/277, conc. range: < 0.03 μ g / kg (24 samples), 0.01-0.02 μ g / kg (6 sa), 0.02-0.04 µg/kg (5 sa), country: UK

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Milk powder

incidence: 213/213, conc. range: < 0.1-0.8 μ g/kg, country: UK incidence: 5/10, conc. range: 0.015-0.243 μ g/kg, country: USA incidence: ?/5, conc. range: 3.83-5.74 μ g/kg, Ø conc.: 4.91 μ g/kg, country: USA incidence: 24/320, conc. range: 0.1-0.4 μ g/kg, country: USA incidence: 192/302, conc. range: tr-3.9 μ g/kg, country: USA \rightarrow milk

Milk products → Dairy products

Millet may contain the following \rightarrow mycotoxins: \rightarrow 15-acetylscirpentriol incidence: nc, Ø conc.: 400 μg/kg, country: USA \rightarrow aflatoxins (AFB₁, AFB₂) incidence: 9/9*, conc. range: 1-27 µg/kg, \emptyset conc.: 9.8 μ g / kg, country: Gambia, *millet, Sanyo (Pennisetum typhoideum) aflatoxins (no specification) incidence: 9*/55, conc. range: 1-100 µg/kg, country: Uganda, *6 samples contained AFB₁, 4 AFB₂, 2 AFG₁, 1 AFG₂ incidence: nc, Ø conc.: 0.3 µg/kg, country: USA \rightarrow cyclopiazonic acid incidence: 2/2*, conc. range: nc, country: India, *kodo millet → deoxynivalenol incidence: 1/4*, conc.: 229 µg/kg, country: Korea, *Indian millet incidence: nc, \emptyset conc.: 300 µg/kg, country: USA \rightarrow nivalenol incidence: 1/4*, conc.: 340 µg/kg, country: Korea, *Indian millet incidence: nc, Ø conc.: 1200 µg/kg, country: USA \rightarrow ochratoxin A incidence: 1/2, conc.: \leq 0.3 µg/kg, country: The Netherlands

→ zearalenone incidence: nc, Ø conc.: 300 µg/kg, country: USA → cereals, → sorghum

Millet meal may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 1/1*, conc.: 720 µg/kg, country: Papua New Guinea, *imported \rightarrow nivalenol incidence: 1/1*, conc.: 1540 µg/kg, country: Papua New Guinea, *imported \rightarrow zearalenone incidence: 1/1*, conc.: 440 µg/kg, country: Papua New Guinea, *imported

The influence of milling on the Milling mycotoxin contamination of the \rightarrow flour fractions compared to the whole \rightarrow grains differs mainly due to the relative distribution of the \rightarrow mycotoxins throughout the kernel. In some cases the degree of kernel contamination is also decisive. E.g. grains showing an overall high \rightarrow deoxynivalenol contamination but predominantly in the surface layer will yield a flour low in DON concentration compared to grains having a lower overall DON contamination but predominantly colonized / contaminated by the mycotoxigenic fungus/ mycotoxins in the endosperm. The behavior of \rightarrow trichothecenes during milling is shown in the table Influence of baking (see Table Milling). During wetmilling of maize the pattern of distribution for deoxynivalenol, \rightarrow nivalenol, and \rightarrow zearalenone follows the physical solubility. As they are highly water-soluble, DON and NIV accumulated in the steep liquor whereas low levels were found in the solid fractions (germ, fibre and gluten). The distribution of the relatively insoluble zearalenone was quite the opposite. Compared to the original concentration in the whole grains levels of three important \rightarrow Fusarium mycotoxins,

→ nivalenol, deoxynivalenol and → zearalenone, in the flour fraction is lowered between 15-100% during milling. If fumonisin (→ fumonisins) contaminated → maize is milled the mycotoxins persist in the wet-milled products. Between 10-40% of the original fumonisin concentration is found in the fiber, gluten and germ fraction.

In a milling study a major portion (60-80%) of \rightarrow aflatoxins (B₁, B₂, G₁, G₂) occurred in the combined bran and polish fraction. These fractions contained 10 times more of the aflatoxins than the milled kernels.

No essential reduction of \rightarrow ochratoxin A concentration was observed in the wholemeal compared to the cleaned \rightarrow wheat kernels. Similar observations were made when white or wholemeal flour were baked into \rightarrow bread. White flour from hard and soft wheat contained only 30 and 60% respectively of the ochratoxin of the uncleaned wheat which was mainly found in the \rightarrow bran and offal fractions. Due to scouring which removes a proportion of the pericarp (bran coat) prior to milling, OTA levels were significantly removed (three-fold) for both hard and soft wheat.

Compared to milled \rightarrow rice an increase of the \rightarrow citrinin concentration in the bran and polish fraction has been observed, more pronounced in highly contaminated samples. However, it seems that citrinin will survive the milling process at least to some extent because this mycotoxin has been found in \rightarrow maize flour from e.g. Thailand. \rightarrow cereals

Miso may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 3/20, conc. range: 1400 µg/kg, country: Brazil \rightarrow fermented products, \rightarrow oriental fermentations

Mitosporic fungi (Syn.: Deuteromycetes, Deuteromycotina, Fungi Imperfecti, conidial fungi, asexual fungi) Artificial assemblage of mitosporic fungi with unknown meiotic states: e.g. \rightarrow Alternaria, \rightarrow Arthrinium, \rightarrow Aspergillus, \rightarrow Cladosporium, \rightarrow Fusarium, \rightarrow Penicillium

Modified atmosphere Compared to fungal growth mycotoxin production $(\rightarrow \text{ mycotoxins})$ is more susceptible to low O_2 and high CO_2 atmospheres $(\rightarrow \text{ atmosphere})$. A significant reduction in mycotoxin formation of some \rightarrow Fusarium spp., \rightarrow Aspergillus spp. and \rightarrow Penicillium spp. could be achieved attaining CO_2 concentrations between 20 and 60%. High CO_2 levels are more effective in preventing mycotoxin formation than reduction in O_2 content.

Product	Mycotoxin	Mycotoxin level
Bread	Deoxynivalenol	No losses, except some iso-DON formed
Cookies	Deoxynivalenol	No losses
Doughnuts (yeast)	Deoxynivalenol	Increase
Popcorn	Deoxynivalenol	Minor losses
Bread	Nivalenol	Minor losses
Bread	Other trichothecenes	Comparable to DON

Milling. Influence of baking and other heat processing on stability of trichothecenes (Scott 1990, modified)

Mold ripened cheese manufactured with \rightarrow Penicillium roguefortii Thom and / or \rightarrow Penicillium camembertii Thom may contain \rightarrow cyclopiazonic acid, \rightarrow mycophenolic acid, and roquefortines $(\rightarrow \text{ roquefortine A \& B, } \rightarrow \text{ roquefortine})$ C). However, adequate ripening and storage conditions (6-10 °C Roquefort, 14-19 °C Camembert) and those prevailing in the cheese during ripening greatly inhibit the enrichment of these \rightarrow mycotoxins. Contamination with \rightarrow mycophenolic acid is prevented by using strains unable to produce this mycotoxin. Only small amounts of these only weakly toxic and non carcinogenic mycotoxins can be found in \rightarrow cheese. It is most unlikely that the consumption of mold ripened cheese is associated with hazards to human health.

 \rightarrow cheese, Blue, \rightarrow cheese, Camembert,

 \rightarrow cheese, Roquefort

Moldy corn poisoning may be due to \rightarrow maize infected with \rightarrow Aspergillus flavus Link and \rightarrow Penicillium *rubrum* and contaminated with \rightarrow aflatoxin B₁ fed to pigs and \rightarrow cattle. In these domestic animals hepatic lesions occurred. \rightarrow Turkey "X" disease

Moldy corn toxicosis (Syn.: \rightarrow Hemorrhagic syndrome) The problem of moldy corn toxicosis occurred in the early 1960s in the midwestern states of the U.S. However, other countries also reported from this irregularly occurring, long-standing disease.

Symptoms in farm animals (\rightarrow cattle, pigs, \rightarrow poultry) mainly occurred in the digestive tract and included a general loss of appetite, dysentery often accompanied by bloody feces, reduction in milk yield, unthriftiness, and loss of weight. Massive hemorrhages (\rightarrow hemorrhage) throughout the body (e.g. bladder, heart, intestines, kidneys, lungs) were noted. Death only occurred in some cases. \rightarrow Trichothecenes such as \rightarrow diacetoxyscirpenol and \rightarrow T-2 toxin produced at low temperatures by a highly toxic strain of \rightarrow Fusarium *tricinctum* isolated from moldy sweet corn (\rightarrow maize) were suspected as causatives of hemorrhagic symptoms of farm animals in the USA. Moldy grain which induced moldy corn toxicosis in pigs was fed to dogs. The symptoms were almost the same as to those observed in pigs and resembled a disease called "hepatitis X". Based on the closely related if not identical syndromes, it was concluded that \rightarrow ATA and moldy corn toxicosis have the same origin, viz. T-2 toxin and diacetoxyscirpenol primarily produced by \rightarrow Fusarium sporotrichioides Sherb.

Moldy sweet potato toxicosis is due to a host parasite interaction (sweet potato / \rightarrow Fusarium *solani*) leading to the production of phytoalexins such as 4- and 1-ipomeanol, ipomeanine, 1,4-ipomeadiol. They are catabolized by the fungus to lung-toxic metabolites wich interfere with the respiration of \rightarrow cattle. Cases of death occured.

A chronic respiratory disease has also been reported from New Guinea where humans consume large quantities of sweet \rightarrow potatoes. Since 4-ipomeanol (as well as ipomeamarone) occurred in slightly blemished sweet potatoes destined for sale in US supermarkets, it is possible that these phenolic compounds are also responsible for the etiology of this human disease.

Monascidin A (Syn.: \rightarrow citrinin)

Moniliformin is a naturally occurring sodium or potassium salt of 1-hydroxycyclobut-1-ene-3,4-dione (see Figure Moniliformin). This mycotoxin (\rightarrow mycotoxins) was first isolated from \rightarrow maize in 1973 contaminated by \rightarrow Fusarium moniliforme Sheldon. During a study to determine the molecular structure of the toxin the corresponding strain losts its ability to produce the metabolite in culture. Isolation and structure elucidation eventually succeeded from a high-producing strain of *F. moniliforme* as a contaminant of \rightarrow millet in Nigeria. Since this strain produced chlamydospores it was recently identified as *F. nygamai*. In contrast to other \rightarrow Fusarium mycotoxins moniliformin occurs only in a very few crops.

CHEMICAL DATA

Empirical formula: C₄HO₃ Na/K, molecular weight: 120/136

FUNGAL SOURCES

At least 15 Fusarium species including Fusarium anthophilum, \rightarrow Fusarium avenaceum (Fr.) Sacc., F. chlamydosporum, \rightarrow Fusarium culmorum (Wm. G. Smith) Sacc., \rightarrow Fusarium moniliforme Sheldon (most of the strains either produce only small amounts or none moniliformin), F. nyagamai, \rightarrow Fusarium oxysporum Schlecht. emend. Snyd. & Hansen, \rightarrow Fusarium proliferatum (Matsushima) Nirenberg, F. sporotrichioides, F. subglutinans are moniliformin producers.

NATURAL OCCURRENCE

 \rightarrow maize, \rightarrow maize flour, \rightarrow maize meal, \rightarrow oats, \rightarrow rye, \rightarrow triticale, \rightarrow wheat There are not many data about the occurrence of moniliformin in \rightarrow food.

Тохісіту

rapid death (ducklings 1 h, rats 3 h) of experimental animals occurred (mycocardial degeneration / \rightarrow edema, respiratory distress, and necrosis (liver, kidney)). Action similar to that of arsenite. LD₅₀ (po): 41.57 mg and 50.00 mg/kg bw female and male rats, respectively.

Detection

GC, HPLC, spectroscopy, TLC

Possible Mycotoxicosis Keshan disease/China



Moniliformin

FURTHER COMMENTS

Although moniliformin occurred ten times more abundantly in foodstuff from areas with a high incidence of human \rightarrow esophageal cancer in the Transkei compared to low-incidence areas, the \rightarrow fumonisins are most probably involved in the etiology of this disease. **Stability:** A moderate stability of moniliformin has been found at room temperature, with 68-77% remaining after 6 days. However, heating at 100 °C for 0,5 h caused a 45% destruction in maize.

Monoacetoxyscirpenol is a 15-acetoxy- $3\alpha,4\beta$ -dihydroxy-12,13-epoxytrichothec-9-ene which belongs to the \rightarrow trichothecenes (\rightarrow mycotoxins) (see Figure Monoacetoxyscirpenol).

CHEMICAL DATA Empirical formula: $C_{17}H_{24}O_6$, molecular weight: 324

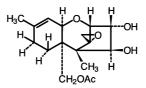
FUNGAL SOURCES \rightarrow Fusarium sambucinum Fuckel, *F. semi*tectum

NATURAL OCCURRENCE

 \rightarrow oats

Тохісіту

bilateral inflammation of the beak area, gastrointestinal hemorrhaging (birds) (\rightarrow hemorrhage), dermatotoxic (rat) LD₅₀ (sc): 0.752 mg/kg bw rat (20-dayold, white, female, weanling)



Monoacetoxyscirpenol

DETECTION GC-MS

Monodeacetylanguidin \rightarrow 15-acetylscirpentriol

Mselenin joint disease This progressive, crippling osteoarthropathic disease is common among blacks in the Mseleni area in Kwazulu (southern Africa). In general, the disease progresses further in women than in men. Life expectancy is not affected but the mobility of the individuals is limited to various degrees. It is estimated that in the endemic region about three thousand people from the Zulu and Toga tribes are affected (38.9% women, 11.1% men).

Since the diet in the endemic region mainly consists of products of plant origin such as \rightarrow cowpeas, \rightarrow peanuts, \rightarrow sweet potatoes, \rightarrow maize, \rightarrow melons, pumpkins, and \rightarrow vegetables and wild \rightarrow fruits lack of calcium, magnesium, and manganese have been proposed as possible causal agents in the etiology of the disease.

However, fungi and their \rightarrow mycotoxins may also be involved. \rightarrow Fusarium moniliforme Sheldon was predominant on maize from the endemic region, 96.3% of the samples were infected. Several other Fusaria, e.g. \rightarrow Fusarium equiseti (Corda) Sacc. sensu Gordon, \rightarrow Fusarium oxysporum Schlecht. emend. Snyd. & Hansen, and \rightarrow Fusarium poae (Wollenw.) Peck which might be implicated in the etiology of bone growth diseases in man and animals could be isolated. In addition, different species of the genera Acremonium, Lasiodiplodia, Macrophomina, Nigros*pora*, and \rightarrow Penicillium frequently occurred on maize and peanuts. Foodstuffs (maize, groundnuts) of affected households showed a higher contamination with these fungi than nonaffected ones.

Muesli may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 4*/14, conc. range: 20-120 μg/kg, country: Germany, *moldy → aflatoxins** incidence: 3/7, conc. range : nc, country: UK → ochratoxin A*** incidence: 2/26, conc. range: 0.4-0.5 µg/ kg, Ø conc.: 0.45 μ g/kg, country: Germany incidence: 6/50, conc. range: \leq 3.9 µg/ kg, country: UK incidence: 3/7, conc. range: nc, country: UK \rightarrow trichothecenes**** incidence: 1/1, conc.: nc, country: UK ** max. level: $\leq 25 \ \mu g / kg$, *** max. level: \leq 50 μg / kg, **** max. level: \leq 5 μg / kg \rightarrow cereals

Muesli ingredients may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins incidence: 2/7, conc. range: 1-5 µg/kg, country: UK \rightarrow deoxynivalenol incidence: 1/4, conc.: traces, country: UK \rightarrow ochratoxin A incidence: 2/7, conc. range: 0.2-1.49 µg/kg, country: Germany incidence: 3/7, conc. range: < 10 µg/kg, country: UK \rightarrow trichothecenes incidence: 1*/1, conc.: nc, country: UK *max. level: ≤ 5 µg/kg

Muffin \rightarrow Maize muffin

Muffin mix may contain the following \rightarrow mycotoxins: \rightarrow fumonisin B₁ incidence: 1/2*, conc.: 80 µg/kg, country: USA, * \rightarrow maize based \rightarrow fumonisin B₂ incidence: 1/2*, conc.: 10 µg/kg, country: USA, *maize based → fumonisins incidence: 1/3*, conc.: nc, country: USA incidence: 6/6*, conc. range: 450-1450 μ g / kg, country: USA, *maize based → zearalenone incidence: 1/5, conc.: 3.1 μ g / kg, country: USA

Mung beans → Beans

mutagenic is a biological, chemical or physical agent which increases the degree of mutation

Mycophenolic acid is a 6-(4-hydroxy-6methoxy-7-methyl-3-oxo-5-phthalanyl)-4methyl-4-hexenoic acid (\rightarrow mycotoxins) which was first isolated in 1896 from \rightarrow Penicillium brevicompactum Dierckx (see Figure Mycophenolic acid).

CHEMICAL DATA

Empirical formula: $C_{17}H_{20}O_6$, molecular weight: 320

FUNGAL SOURCES

P. brevicompactum, P. raciborskii, \rightarrow Penicillium roquefortii Thom chemotype I and II.

NATURAL OCCURRENCE

 \rightarrow cheese, \rightarrow cheese, Bleu des Causses,

 \rightarrow cheese, Blue, \rightarrow cheese, Gorgonzola, \rightarrow cheese, Roquefort

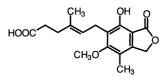
Generally, blue veined cheeses are very good substrates for mycophenolic acid production and may contain relatively high concentrations but Roquefort cheese is particularly suitable for the formation of mycophenolic acid.

TOXICITY

relatively less toxic, showing antibiotic, antitumor, and \rightarrow mutagenic activity, chromosome aberrations occurred in mice

LD₅₀ (po): 2500 mg/kg bw mice

DETECTION mainyl TLC



Mycophenolic acid

FURTHER COMMENTS

Although some adverse clinical reactions such as diarrhea, cramps and nausea occurred after a daily application of 2.4 -7.2 g for 52-104 weeks, this substance seems to be a promising drug for the treatment of psoriasis.

Mycosphaerellaceae → Dothideales

Mycotoxic nephropathy \rightarrow Mycotoxic porcine nephropathy

Mycotoxic porcine nephropathy (Syn.: Mycotoxic nephropathy) was first discovered in 1928 in Denmark while epidemics occurred in 1963 and 1971, associated with high-moisture grain (\rightarrow grains) due to unusual climatic conditions. The major causal agent is \rightarrow ochratoxin A but other substances like \rightarrow citrinin and \rightarrow viomellein (quinone) may also be involved. These nephrotoxic \rightarrow mycotoxins have been isolated from \rightarrow barley associated with mycotoxin porcine nephropathy. They mainly act on the \rightarrow renal tuber system, especially on the proximal tubules.

 \rightarrow Polydypsia and / or \rightarrow polyuria are the most characteristic renal alterations in domestic animals like pigs and horses due to feeding of moldy grain or hay. In pigs and \rightarrow poultry chronic ochratoxicosis may be manifestated by retarded growth rates. Since renal damages are easily overlooked they are usually detected only during inspection in slaughterhouses.

Experimental studies showed that in pigs, the corresponding symptoms occurred

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after feeding (i) grain infected with a citrinin / ochratoxin producer, (ii) this mold directly, (iii) application of citrinin and/or OTA as pure substances. 0.01-0.08% of slaughtered pigs from slaughterhouses in Denmark showed prevalence rates of porcine nephropathy. In most cases the affected kidneys (10 µg OTA / kg < 25 μ g OTA / kg) are condemned but the remaining carcass is accepted for consumption. In different European countries 25-39% of affected kidneys contained 2-100 µg OTA / kg. These kidneys are swollen and pale with a mottled surface. Histopathological abnormalities may include periglomerular \rightarrow fibrosis, degeneration of the proximal tubules, followed by atrophy of the tubular epithelium, hyalinization of some glomeruli, and interstitial fibriosis in the \rightarrow cortex. In a later stage, enlargement of the kidneys to several times the normal size may occur, associated with pronounced gross changes in texture and color, \rightarrow jaundice, and advanced cellular damage. Renal functions are deeply impaired. Depressed weight gains and decreased performance may also occur after feeding higher toxin levels. If uremia is developed, the whole carcass is condemned at \rightarrow meat inspection in Denmark ($\geq 25 \ \mu g$ OTA in the kidneys/ kg).

OTA contaminated \rightarrow meat and organs of pigs (kidneys, liver) may be a source for human OTA intake mainly due to the consumption of contaminated \rightarrow sausages.

Mycotoxicosis Toxic syndromes resulting from the ingestion of \rightarrow foods or feeds contaminated with fungal toxins by man and animals are known as mycotoxicosis. These, often seasonally occurring, disorders are primarily found in climatic regions with high rainfall, high relative humidity, and high temperatures. In some cases drought, insect damage and/ or cracked kernels during harvesting enhance fungal growth (e.g \rightarrow Aspergillus flavus Link, \rightarrow Aspergillus parasiticus Speare / \rightarrow peanuts) and subsequent mycotoxin contamination. The development of the fungi is further promoted by the presence of excessive chaff in the harvested \rightarrow grains and seeds although the molds may even no longer be present in contaminated grain.

It is suggested that about 50 fungal metabolites are involved in man and animal mycotoxicosis while ten of these \rightarrow mycotoxins such as \rightarrow aflatoxin B₁, \rightarrow ochratoxin A, \rightarrow sterigmatocystin and \rightarrow fumonisin B₁ are carcinogenic (\rightarrow carcinoma) (see Table Mycotoxicosis). Substantial difficulties arise when making the right diagnose of a mycotoxicosis because (i) mycotoxins, especially at low

dosis, or unknown toxins are difficult to detect in food and feed, (ii) contaminated food or feed are often disposed before a mycotoxicosis is suspected, (iii) clinical signs and / or symptoms are often not precise and of an acute nature, (iv) physicians and veterinarians are often not familar with the symptomatology of mycotoxicosis, (v) the "dose-response" principle is difficult to apply to the diagnosis of a mycotoxin-related disease, (vi) the interaction between individual mycotoxins and their effects on man and animals is yet not well elucidated. Because of animals due to lower feed quality and the way in which they are fed, animals are more likely to be exposed to mycotoxins than humans and a higher incidence of mycotoxicosis in animals has been reported. Certain common features for a mycotoxicosis are presented below:

- true cause is not immediately identified
- outbreaks often are seasonal
- disease is food or feed-related (e.g. \rightarrow peanuts, \rightarrow maize, \rightarrow rice)*
- no significant pathogenic microorganisms are present*

- fungal activity is obvious in the suspected food / feed
- treatment with drugs or antibiotics is not effective
- disease is not contagious, neither transmissible nor infectious*
- age, sex, and nutritional status are often decisive for severity of the disease
- withdrawal of suspected food / feed leads to signs of improvement*
- isolation of sufficient amounts of the mycotoxin(s) from the food / feed or man / animals to cause the disease*
- mycotoxin(s) isolated are known to produce the typical symptoms of the disease*

 feeding trials with the suspect ration reproduce the disease *criteria for a true mycotoxicosis

Possible Mycotoxicosis

→ acute cardiac beri-beri, → aflatoxicosis, AIDS, → akakabi byo disease, → alimentary toxic aleukia, → Balkan endemic nephropathy, → equine leukoencephalomalacia, → ergotism, → indian childhood cirrhosis, → Kashin-Beck disease, → kodua poisoning, → Kwashiorkor, → onyalai, → Pellagra, → porcine pulmonary edema

 \rightarrow premature thelarche, \rightarrow Reye's syndrom

Mycotoxicosis	Involved mycotoxin(s)	Involved foodstuff
Acute cardiac beriberi	Citreoviridin	rice
Aflatoxicosis (acute)	Aflatoxins	maize, peanuts
Akakabi byo disease	Trichothecenes	maize, wheat
	(e.g. deoxynivalenol,	
	fusarenon X)	
Alimentary toxic aleukia	Trichothecenes (e.g. diace-	cereals, mainly proso millet & wheat
	toxyscirpenol, HT-2 toxin,	but also barley, rye, oats, buckwheat
	T-2 toxin, nivalenol)	
Arthrinium sugarcane poisoning	β-Nitropropionic acid,	sugarcane
	fumonisins?	
Balkan endemic nephropathy	Ochratoxin A, citrinin	cereals, cereal products, beans, pig
		products
Deoxynivalenol toxicosis	deoxynivalenol, nivalenol,	grains, flours
	acetyldeoxynivalenol, T-2	
	toxin	
Ergotism	Ergot alkaloids	cereals, mainyl rye
Esophageal cancer	Fumonisins	maize, maize products
Fusariotoxicosis	Fusarium toxins, e.g. deoxy- nivalenol, zearalenoene	maize, wheat
Indian childhood cirrhosis	Aflatoxins	rice, peanut oil, human breast milk
Kashin-Beck disease	fusarochromanone, T-2 toxin	cereals, mainly maize, wheat
Kodua poisoning	Cyclopiazonic acid	grains (e.g. rice), bread
Kwashiorkor	Aflatoxins	starchy and low in protein (e.g. rice, maize, plantains
Mseleni Joint disease		maize, peanuts
Reye's syndrome	Fusarium toxins and others	different kinds of foodstuff e.g. milk,
		peanuts, rice
Onyalai	Aflatoxins	millet, sorghum
Pellagra	Tenuazonic acid, monili-	maize
~	formin	
	fumonisins, kojic acid, tri-	
	chothecenes, zearalenone	
Sago hemolysis	?	sago

Mycotoxicosis. Possible involvement of food relevant mycotoxins in human mycotoxicosis

Mycotoxin control Prevention (\rightarrow mycotoxin prevention) of mycotoxin contamination by "good farm management practice" is the most effective measure in the production of mycotoxin free or low contaminated \rightarrow foods and feeds. This includes moisture and temperature control which have a crucial effect on fungal growth and mycotoxin formation.

Mycotoxin degradation

physical: adsorption, heat, irradation chemical: acids, bases, bisulfite, oxidizing agents, vitamin C; however, many of these compounds are not in practical use since they may render the products unsafe

biological: various fungi (e.g. \rightarrow Aspergillus *niger*, *Rhizopus stolonifer*), bacteria (e.g. \rightarrow Flavobacterium aurantiacum, lacto-bacteria)

Mycotoxin detection is carried out by different techniques e.g.

physicochemical: gas chromatography (GC), high performance liquid chromatography (HPLC), thin layer chromatography (TLC)

immunoassays: enzyme-linked immunosorbent assay (ELISA), radio-immunoassay (RAI)

bioassays: animals, cells and tissue cultures, microorganisms

In seeds (e.g. \rightarrow peanuts) and fruits (e.g. \rightarrow figs) detection of \rightarrow mycotoxins is difficult since there is an uneven distribution in these kinds of substrates. However, in processed \rightarrow foods and drinks mycotoxins seem to be distributed in a more homogenous manner.

Mycotoxin legislation Currently, 77 countries are known for their mycotoxin regulations (see Table Mycotoxin legislation in the Appendix).

Mycotoxin prevention is achieved by the following factors:

control of the **moisture** content of stored products: cereal \rightarrow grains: < 13%, \rightarrow soybeans: < 12%, seeds with a high lipid content: 7% control of the **temperature** in storage: in

general, temperatures below 4 °C will prevent mycotoxin production control of the **atmosphere** in storage: mold growth / mycotoxin production is depressed by low oxygen and / or high concentration of other gases. Inhibition of aflatoxin (\rightarrow aflatoxins) formation occurred at 1% O₂ while the production of \rightarrow sterigmatocystin and \rightarrow patulin was completely depressed at 0.2% O₂. Only small amounts of sterigmatocystin were produced at 90% CO₂, patulin could not be detected.

microbial competition: different microorganisms such as \rightarrow Aspergillus *niger*, *Rhizopus stolonifer* or lactic bacteria, decreased / inhibited aflatoxin production. Little to no aflatoxin contamination occurred in grain invaded by a mixture of fungi, including \rightarrow Aspergillus flavus Link.

antimycotic agents: growth of mycotoxigenic fungi is inhibited by sufficient amounts of e.g. acetic acid, benzoic acid, propionic acid, sorbic acid or natamycin. In addition, production and manufacture of low fungal contaminated raw material, pasteurisation and sterilization of intermediate- and endproducts, suitable packaging, use of preservatives, suitable cooling, freezing and drying techniques, feeding of mycotoxin-free feed, and the use of non-toxic starter cultures in the fermentation industry enables the production of non-contaminated foods.

Mycotoxin producers Almost 350 mold species, mainly mitosporic fungi and only a few ascomycota (e.g. \rightarrow Claviceps *purpurea*), are known for their mycotoxi-

genic potential (see Table Mycotoxin producers). The most common and important mycotoxin producers which cause mycotoxin contamination of plants (e.g seeds, \rightarrow fruits) in the field belong to the genera \rightarrow Alternaria (to a minor degree), \rightarrow Aspergillus and \rightarrow Fusarium. The main mycotoxigenic species which attack plant products after harvest, during transport or when in storage are *Aspergillus* and \rightarrow Penicillium (for further information see also the listed species belonging to each single genus).

Correct fungal identification based on internationally agreed criteria is necessary since there is a close relationship between fungal species and the secondary metabolites they produce. Mycotoxin production is not uniform throughout all strains of a species. Even in the case of a producing strain, mycotoxin formation depends on environmental and nutritional conditions. These phenomena enable strains of potentially mycotoxigenic fungi to be used in food manufacture i.e. Aspergillus flavus group (e.g. \rightarrow aflatoxins) / koji, \rightarrow Fusarium graminearum Schwabe (e.g. \rightarrow zearalenone)/ microbial protein, \rightarrow Penicillium roquefortii Thom (e.g. \rightarrow PR toxin) / Blue cheese $(\rightarrow \text{ cheese, Blue}).$ → mycotoxins

Mycotoxin production Since \rightarrow mycotoxins are secondary metabolites, they are usually produced in the late exponential or early stationary phase. Production is mainly influenced by the following factors:

Moisture: High humidity and a high $\rightarrow a_w$ favor mycotoxin production. Synthesis of \rightarrow aflatoxins starts at $a_w > 0.83$, \rightarrow citrinin: $a_w 0.83$, \rightarrow ochratoxin A: a_w 0.83, \rightarrow patulin: $a_w 0.85$, \rightarrow penicillic acid: $a_w 0.80$. In \rightarrow grains maximum amounts of \rightarrow mycotoxins are produced at moisture contents between 20-25%.

Temperature: \rightarrow Aspergillus spp. aflatoxins: 9-42 °C, \rightarrow sterigmatocystin: refrigeration temperature

→ Penicillium spp. → cyclopiazonic acid: 4 °C, ochratoxin A, penicillic acid: 4-31 °C, patulin: 0-24 °C, pentirem A (→ penitrems): 6 °C

→ Fusarium spp. → trichothecenes: at and below 10 °C (→ Fusarium sporotrichioides Sherb.: 1.5 to 4 °C optimal production).

Temperatures well below 0 °C will prevent mycotoxin formation. There is a close link between moisture and temperature in mycotoxin production.

Oxygen levels: In general fungi need adequate oxygen concentrations to grow but some species of the genera, e.g. *Mucor*,

Mycotoxin producers.	Mycotoxigenic	fungal genera	
			_

Acremonium	Dichotomomyces**	Myrothecium	Rosellinia**
Alternaria*	Diplodia	Microdochium	Sclerotinia**
Aspergillus*	Drechslera	Monographella**	Spacelia
Bipolaris	Epichloe**	Nigrosabulum**	Stachybotrys
Botryodiplodia	Epicoccum	Nigrospora	Talaromyces**
Byssochlamys* **	Fusarium*	Paecilomyces	Thielavia*
Ceratocystis**	Gibberella* **	Penicillium*	Trichoderma
Chaetomium**	Gliocladium	Periconia	Trichothecium
Cladosporium*	Gloeotinia**	Phoma*	Verticillium
Claviceps* **	Khuskia**	Phomopsis	Verticimonosporium
Colletotrichum	Metarhizium	Pithomyces	Zygosporium
Curvularia			,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,

* important in food

** teleomorphic state

Rhizopus and *Fusarium* are able to develop under anaerobic conditions $(\rightarrow \text{ atmosphere})$

Substrate: Mycotoxin formation is enhanced by carbohydrates (e.g. glucose, saccharose), certain amino acids (e.g. asparagin, glycin), fatty acids and zinc (aflatoxins). Generally, plant-derived-products characterized by a high carbohydrate content are more likely to be prone to mycotoxin contamination than animal products (high protein content). The only important exception is \rightarrow milk. **Damage, plant stress:** Damage of plants (mechanical and/or insects) and/or

drought stress in e.g. \rightarrow peanuts or \rightarrow maize facilitate invasion of aflatoxigenic fungi and subsequent aflatoxin formation.

In addition, mycotoxin formation is influenced by the availability of trace elements, genetic strain variation and and/or competition with other organisms.

Mycotoxin stability In general, \rightarrow mycotoxins are quite (heat) stable in most \rightarrow food products but there are some exceptions; see e.g. \rightarrow fusarin C, \rightarrow patulin, and \rightarrow penicillic acid. (For further information see each single mycotoxin and the contaminated food items.)

Mycotoxins are structurally diverse complex organic compounds of low molecular weight (MW generally lower than 700; \rightarrow fumonisins, e.g. FB₁ = 721) which belong to the large and diverse group of secondary fungal metabolites. They are not all necessarily \rightarrow mycotoxins such as the antibiotic penicillin. Based on the inherent toxic effects in higher organisms, a chemical might be called a mycotoxin.

Mycotoxins are found in different chemical groups e.g. pyrones, anthrachinones, coumarins, macrolides, steroids and cyclic polypeptides. Formation usually occurs during the late exponential or early stationary phase of fungal development. These non-antigenic organic compounds are produced by a wide range of fungi. At least 15 different mycotoxins are synthesized by some species of these genera. They are further characterized by their frequent specificity with regard to the taxonomy of the producing fungi. It is estimated that approximately 400 toxic fungal compounds do exist. Almost all plant products may serve for mold growth and mycotoxin production. To a minor degree animal products such as \rightarrow milk and \rightarrow meat may be contaminated. Humans are exposed to mycotoxins mainly through the consumption of \rightarrow foods directly contaminated by mycotoxin-producers and their mycotoxins (e.g. \rightarrow aflatoxins, \rightarrow trichothecenes, \rightarrow patulin) or by ingestion of residue containing \rightarrow meat (e.g. \rightarrow ochratoxin A) or \rightarrow milk (e.g. \rightarrow aflatoxin M₁).

FUNGAL SOURCES

Although ca. 350 different fungal species are known to be mycotoxin producers, fungi of the genera \rightarrow Aspergillus, \rightarrow Fusarium, \rightarrow Penicillium and \rightarrow Alternaria (to a minor degree) are the most important. Worldwide at least 100 mycotoxigenic fungal species are associated with naturally occurring diseases in animals and humans. \rightarrow mycotoxin producers

NATURAL OCCURRENCE

→ Cereals and → oil seeds (→ nuts) and products derived from them are most likely to be contaminated by mycotoxigenic fungi / → mycotoxins. Several factors like area of crop growth, climate, conditions during growth, harvesting and storage are decisive for mycotoxin contamination of the crop. The warm and moist weather in tropical and subtropical countries favors the rapid growth of (aflatoxigenic) fungi and subsequent mycotoxin contamination (especially → aflatoxins) in such → seeds. Crops

grown in temperate regions are less prone to mycotoxin contamination. Here, \rightarrow trichothecenes and \rightarrow ochratoxin A predominate. Worldwide 25% of the annually produced food crops are contaminated with detectable amounts of mycotoxins (FAO 1985) resulting in economic losses of billions of dollars/year. The mycotoxins most commonly found in \rightarrow food and feedstuff are aflatoxins, \rightarrow fumonisins, ochratoxin A, patulin, trichothecenes and \rightarrow zearalenone. To minimize mycotoxin exposure to man almost 80 countries possess legal or recommended limits for mycotoxins such as aflatoxins, chaetomin, \rightarrow deoxynivalenol, \rightarrow diacetoxyscirpenol, \rightarrow fumonisin B_1 , \rightarrow fumonisin B_2 , ochratoxin A, patulin, phomopsin, stachybotryotoxin, \rightarrow T-2 toxin, and zearalenone.

TOXICITY

Often a substrate is contaminated by different mycotoxins which may act synergistically or additively. This fact limits the value of the administration of a pure crystalline mycotoxin, e.g. turkey "X" disease / aflatoxins and \rightarrow cyclopiazonic acid. Several factors such as molecular structure, dosage, duration of intake, species, age, sex, condition and nutrient status of the affected organism are decisive for the

detrimental effect of a mycotoxin. Its toxicity may be limited to only one or a few species, but another mycotoxin may affect a wide range of organisms. Mycotoxins are carcinogenic (e.g. \rightarrow aflatoxin B_1 , fumonisin B_1 , \rightarrow fusarenon X, griseofulvin, \rightarrow sterigmatocystin), cardiotoxic (e.g. \rightarrow ergot alkaloids, \rightarrow penicillic acid), dermatotoxic (e.g. trichothecenes such as \rightarrow HT-2 toxin), emetic (e.g. deoxynivalenol, T2- toxin), hemorrhagic (e.g. \rightarrow byssochlamic acid, patulin), hepatotoxic (e.g. \rightarrow islanditoxin, \rightarrow luteoskyrin, \rightarrow rubratoxins, \rightarrow rugulosin), \rightarrow immunosuppressive (e.g. ochratoxin A, trichothecenes), mutagenic (e.g. aflatoxins, \rightarrow alternariol methyl ether, \rightarrow altertoxin I-III), nephrotoxic (e.g. citrinin, ochratoxin A, penicillic acid, \rightarrow viomellein, \rightarrow xanthomegnin), estrogenic (zearalenone), neurotoxic (e.g. \rightarrow citreoviridin, cyclopiazonic acid, ergot alkaloids, \rightarrow penitrems), teratogenic (e.g. aflatoxins, \rightarrow alternariol) and / or tremorgenic (e.g. \rightarrow tremorgenic mycotoxins) (for further information see also each single mycotoxin). Chronic effects are merely the inhibition of protein synthesis and / or growth. At least some mycotoxins probably have synergistic effects in vivo (see Table Mycotoxins 1).

Mycotoxin	Mutagenic	Teratogenic	Carcinogenic
Aflatoxin	+++	+++	+++
Citrinin	-+	+	+
Cyclochlorotine			+
Fumonisin B ₁			+
Fusarenon X		+	+
Luteoskyrin	-		+
Ochratoxin A	+	+	+
Patulin	+	+	+
Penicillic acid	+		+
Rugulosin	-+		+
Sterigmactocystin	+	+	+++
T-2 toxin	-	+	+
Zearalenone	+	+	+

Mycotoxins 1. Toxicological effects of mycotoxins (Pohland 1993, modified)

Mycotoxins

1.	Mold damaged foodstuffs of plant origin a) Agricultural products	e.g. cereals, fruits, oilseeds (mainyl nuts), pulses, spices
	b) Consumer foods	1 • 1
2.	Residues in tissues and products of animal origin due to mold contaminated feedstuff	e.g. meat (mainly kidneys, liver) & meat products (mainly sausages), milk, dairy products (mainly cheese)
3.	Mold-ripened foods	e.g. cheeses (mainly Roquefort & Camem- bert cheese), meat products
4.	Fermentation products	e.g. enzymes, microbial proteins, organic acids, other food additives

Mycotoxins 2. Possible routes for mycotoxin contamination of human foods (Jarvis 1976, modified)

Testing different mycotoxins from Aspergillus spp. and Penicillium spp. the toxicity decreased between 0.001-100 μ g/ embryo as follows: aflatoxin B₁, ochratoxin A, \rightarrow PR toxin, \rightarrow aflatoxin B₂, aflatoxin M₁, sterigmatocystin, \rightarrow aflatoxin G₂, patulin, rubratoxin B (\rightarrow rubratoxins), secalonic acid D (\rightarrow secalonic acids), \rightarrow mycophenolic acid, α -cyclopiazonic acid, penicillic acid, citrinin, brevianamide A and griseofulvin. However, it is very difficult to assess the present-day risk to human health because quantifying exposure of mycotoxins in the diet is problematic (see Table Mycotoxins 2).

 \rightarrow extracellular mycotoxins, \rightarrow intracellular mycotoxins

Myocin (Syn.: \rightarrow patulin)

Neosartorya \rightarrow Trichocomaceae, anamorph \rightarrow Aspergillus fumigatus group *N. fischeri* possesses heat-resistant ascospores which cause spoilage of \rightarrow fruit juices and other heated (pasteurized) fruitbased products. *N. fischeri* may produce \rightarrow mycotoxins such as avenaciolide, fumitremorgins, terrein, verruculogen.

Neosolaniol (Syn.: solaniol, 8 α -hydroxydiacetoxyscirpenol) belongs to the group of naturally-occurring \rightarrow trichothecenes (4 β ,15-diacetoxy-3 α ,8 α -dihydroxy-12,13epoxytrichothec-9-ene), which was first isolated from \rightarrow Fusarium sporotrichioides Sherb. in 1971 (see Figure Neosolaniol). The previous name solaniol was changed by Ueno in 1972 to neosolaniol.

CHEMICAL DATA

Empirical formula: $C_{19}H_{26}O_8$, molecular weight: 382

FUNGAL SOURCES

F. acuminatum, \rightarrow Fusarium avenaceum (Fr.) Sacc.?, \rightarrow Fusarium culmorum (W. G. Smith) Sacc.?, \rightarrow Fusarium equiseti (Corda) Sacc. sensu Gordon, \rightarrow Fusarium graminearum Schwabe, \rightarrow Fusarium oxysporum Schlecht. emend. Snyd. & Hansen, \rightarrow Fusarium poae (Peck) Wollenw.,

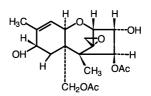
 \rightarrow Fusarium sambucinum Fuckel, \rightarrow Fusarium semitectum Berk. & Rav.?, *F. sporotrichioides*

NATURAL OCCURRENCE \rightarrow barley, \rightarrow curry, \rightarrow ginger, \rightarrow maize, \rightarrow oats, \rightarrow wheat

Тохісіту

cellular degeneration, karyorrhexis in actively dividing cells of thymus, lymph nodes, spleen, bone marrow, intestine, and testes, dermatotoxic LD_{50} (ip): 14.5 mg/kg bw mice

Detection GC, MS, spectroscopy, TLC



Neosolaniol

Further Comments

Neosolaniol produced by *F. sporotri-chioides* may be associated with outbreaks of \rightarrow ATA and \rightarrow bean hull poisoning.

Nephritis inflammation of the kidney

Nephrotoxin e.g. a mycotoxin which damages the kidney tissue

β-Nitropropionic acid (Syn.: bovinocidin, hiptagenic acid, 3-nitropropionic acid) was first isolated from the root bark of Hiptage in 1920 but later it was reported as being a metabolite of \rightarrow Penicillium spp. and *Streptomyces* spp. (see Figure ß-Nitropropionic acid). As a toxic metabolite of different \rightarrow mitosporic fungi it is probably involved in a Chinese \rightarrow mycotoxicosis (\rightarrow mycotoxins).

CHEMICAL DATA Empirical formula: $C_3H_5NO_4$, molecular weight: 119

FUNGAL SOURCES

Arthrinium ssp. (A. sacchari, A. saccharicola), \rightarrow Aspergillus spp. (e.g. possibly \rightarrow Aspergillus flavus Link, \rightarrow Aspergillus oryzae (Ahlburg) Cohn, \rightarrow Aspergillus parasiticus Speare), \rightarrow Penicillium spp. (e.g. *P. atrovenetum*).

NATURAL OCCURRENCE \rightarrow cheese, sugarcane ?,

N

β-Nitropropionic acid

Toxicity

clinical signs: rapid respiration with subsequent apnea, incoordination, marked dilation (subcutaneous and visceral blood vessels), mottled liver

 LD_{50} (po): 110 and 68.1 mg/kg bw male and female mice, respectively

DETECTION TLC

Possible Mycotoxicosis \rightarrow Arthrinium sugarcane poisoning

Nivalenol belongs to the group of naturally-occurring \rightarrow trichothecenes ($3\alpha,4\beta,7\alpha,15$ -tetrahydroxy-12,13-epoxytrichothec-9-en-8-one) which was first isolated from \rightarrow Fusarium sporotrichioides Sherb. in 1967 (see Figure Nivalenol). The first report on natural occurrence (Japanese scabby \rightarrow barley) dates from 1972 (together with \rightarrow deoxynivalenol).

CHEMICAL DATA

Empirical formula: $C_{15}H_{20}O_7$, molecular weight: 312

FUNGAL SOURCES

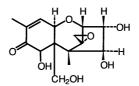
→ Fusarium equiseti (Corda) Sacc. sensu Gordon, → Fusarium graminearum Schwabe, → Fusarium sambucinum Fuckel (?), → Fusarium semitectum Berk. & Rav. (?), F. sporotrichioides

NATURAL OCCURRENCE

barley, \rightarrow barley flour, \rightarrow barley malt, \rightarrow beer, \rightarrow bread, \rightarrow chapatti, \rightarrow chilli sauce, \rightarrow curry, \rightarrow curry paste, \rightarrow flour, \rightarrow foods, \rightarrow garlic, \rightarrow ginger, \rightarrow grains, \rightarrow job's tears, \rightarrow maize, \rightarrow millet, \rightarrow millet meal, \rightarrow noodles, \rightarrow oats, \rightarrow rice, \rightarrow rye, \rightarrow rye flour, \rightarrow sesame seeds, \rightarrow sorghum, \rightarrow soybeans \rightarrow tandoori, \rightarrow wheat, \rightarrow wheat bran In contrast to deoxynivalenol, nivalenol is a less common contaminant of grains.

TOXICITY Similar to deoxynivalenol although DON has a greater acute toxicity. dermatotoxic, emetic, inhibition of DNA synthesis follows inhibition of protein synthesis LD_{50} (ip): 4.1 mg/kg bw mice DETECTION GC, HPLC, spectroscopy, TLC **FURTHER COMMENTS** Nivalenol is often co-occurring with \rightarrow deoxynivalenol. It may be produced from \rightarrow fusarenon X by a chemical or enzymatic deacetylation reaction (\rightarrow Fusarium nivale (Fr.) Ces.) because nivalenol lacks one acetyl group which is characteristic for fusarenon X.

Noodles During Chinese noodle making losses of \rightarrow deoxynivalenol and \rightarrow nivalenol amounted to \approx 30-40%. In these noodles no \rightarrow diacetoxyscirpenol, \rightarrow neosolaniol, \rightarrow T-2 toxin and \rightarrow fusarenon X could be detected after manufacturing (artifical contamination). Losses of the afore mentioned \rightarrow mycotoxins during processing of Japanese noodles were in the range of \approx 40-70%. Noodles may contain the following mycotoxins: \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 1/4*, conc.: 0.4 µg/kg, country: UK, $* \rightarrow$ wheat → deoxynivalenol incidence: 2/2*, conc. range: 2500-2720 μ g/kg, Ø conc.: 2610 μ g/kg, country: Canada, *Japanese noodles



Nivalenol

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incidence: 2/2*, conc. range: 2120-2310 μ g / kg, Ø conc.: 2215 μ g / kg, country: Canada, *Chinese noodles incidence: nc/4*, conc. range: 11-92 μ g / kg, country: UK, *wheat \rightarrow fumonisins (FB₁, FB₂) incidence: $1/4^*$, conc.: 26 µg/kg, country: UK, *wheat \rightarrow nivalenol incidence: nc/4*, conc. range: 14-26 μ g / kg, country: UK, *wheat \rightarrow ochratoxin A incidence: 97/106, conc. range: \leq 4.9-5.3 μg/kg, country: Germany incidence: 1/4*, conc.: 0.2 µg/kg, country: UK, *wheat may contain the following Nutmeg

 \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 3/5*, conc. range: 2.5-5.5 μg/kg, country: Canada, *imported incidence: 4/13, conc. range: 5-37.5 μg/kg, country: West India incidence: 11/28, conc. range: tr-7.7 μg/kg, country: Germany incidence: 3*/13, conc. range: 5-15 μg/kg, country: Germany, *moldy incidence: 2/3, conc. range: 0.4-0.6 μ g/kg, Ø conc.: 0.5 μ g/kg, country: Japan incidence: 29/67, conc. range: 0.2-16 μg/kg, country: Japan incidence: 25/56, conc. range: 0.2-60.3 μg/kg, country: Japan incidence: 30/32, conc. range: 1-23.2 µg/kg, country: The Netherlands \rightarrow aflatoxin B₂ incidence: 3/5*, conc. range: 0.75-1.1 µg/kg, country: Canada, *imported incidence: 1/3, conc.: $0.2 \mu g/kg$, country: Japan incidence: 8/67, conc. range: tr-0.6 μg/kg, country: Japan incidence: 25/56, conc. range: 0.1-0.2 μg/kg, country: Japan \rightarrow aflatoxin G₁

incidence: 1/3, conc.: 0.2 μg/kg, country: Japan incidence: 1/67, conc.: 0.3 µg/kg, country: Japan incidence: 25/56, conc. range: 0.2-1.4 μg/kg, country: Japan \rightarrow aflatoxin G₂ incidence: 25/56, conc. range: 0.3 µg/kg, country: Japan \rightarrow aflatoxins (no specification) incidence: 30*/32, conc. range: 2.7-36.5 μ g / kg, country: The Netherlands, *AFB₁, AFB₂, AFG₁, AFG₂ incidence: 5/5*, conc. range: $\leq 20 \ \mu g / kg$, \emptyset conc.: 13 µg/kg, country: USA, *imported \rightarrow spices

Nuts (no specification)

Almost all types of nuts grown in different geographic regions are prone to mycotoxin contamination, mainly \rightarrow aflatoxins. The degree of contamination as well as the observed levels are subject to significant variation. Although some kind of mycotoxin contamination has been established, nuts like \rightarrow almonds, \rightarrow cashew nuts, \rightarrow hazelnuts, and \rightarrow walnuts generally show a lower degree of contamination due to shell texture and harvesting methods.

 \rightarrow Peanuts and their derived products are most frequently and heavily contaminated. Individual \rightarrow Brazil nuts and \rightarrow pistachio nuts may contain levels of aflatoxins as high as several micrograms per gram. A blue fluorescence under ultraviolet light in e.g. Brazil nuts, peanuts, \rightarrow pecans, and pistachio nuts may indicate an aflatoxin contamination. Mechanical and electronic sorting leads to a significant reduction in aflatoxin contamination. Pneumatic separation is also used to remove contaminated nuts because fungal infected nuts are often lighter than healthy ones. However, it is not possible to detect fungal and mycotoxin contamination of inshell nuts during manufacture. Suspected individual nuts have to be handsorted and removed by the consumer. Compared with retail marketed whole nuts such as peanuts and almonds, processed nuts (chopped, sliced, grind etc.) usually show a higher degree of mycotoxin contamination. It seems that the aflatoxins are more evenly distributed in these comminuted and mixed samples. These brands give a better reflection of the true toxin concentrations compared to whole nuts due to inadequate sampling techniques.

According to Frisvad (1988) nuts may be contaminated with the following myco-toxins: aflatoxins, \rightarrow citrinin, \rightarrow cyclopia-zonic acid, emodin, roquefortine A

 $(\rightarrow \text{ roquefortine A \& B}), \text{ pentirem A}$

 $(\rightarrow \text{ penitrems})$, rugulovasine A, \rightarrow secalonic acid D, \rightarrow sterigmatocystin, wentilacton.

Nuts may contain the following \rightarrow mycotoxins:

aflatoxins

incidence: 3/5, conc. range: 1-8900 µg/kg, country: UK

 \rightarrow ochratoxin A

incidence: 1/5, conc.: traces, country: UK incidence: 3/5, conc.: ca. 1 μ g/kg, country: UK

almonds, Brazil nuts, cashew nuts, \rightarrow coconut, hazelnuts, \rightarrow marzipan, peanuts, pecans, pistachio nuts, walnuts

Nuts (mixed)

may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 2/10*, conc. range: 10-93 μ g / kg, Ø conc.: 51.5 μ g / kg, country: Finland, *imported \rightarrow aflatoxin B₂ incidence: 1/10*, conc. range: 29 µg/kg, country: Finland, *imported incidence: 1/16*, conc.: traces, country: Norway, *imported \rightarrow aflatoxin G₁ incidence: 1/16*, conc.: traces, country: Norway, *imported \rightarrow aflatoxin G₂ incidence: 1/16*, conc.: traces, country: Norway, *imported \rightarrow aflatoxins incidence: 1/3, conc.: 7 μ g/kg, country: USA

0

Oat bran may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A incidence: 1/14, conc.: 0.1 µg/kg, country: Germany incidence: 5/13, conc.: \leq 4.9 µg/kg, country: Sweden \rightarrow bran

Oat flakes may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 7/65, conc.: $\leq 1.5 \ \mu\text{g}/\text{kg}$, country: Germany \rightarrow ochratoxin A incidence: 4/92, conc. range: 1.2-2.0 $\mu\text{g}/\text{kg}$, country: Germany \rightarrow corn flakes, \rightarrow maize flakes, \rightarrow cereal flakes

Oats may contain the following \rightarrow mycotoxins: 3-acetoxynivalenol incidence: 2/6, conc. range: $< 200 \mu g / kg$, country: Finland → 3-acetyldeoxynivalenol incidence: 12/21, conc. range: 6-219 μ g / kg, Ø conc.: 67 μ g / kg, country: Finland \rightarrow aflatoxin B₁ incidence: 3/304, Ø conc.: 6 µg/kg, country: USA \rightarrow alternariol incidence: 8/339, conc.: nc, country: Austria incidence: 24/139, conc. range: 3-64 μ g/kg, Ø conc.: 15.6 μ g/kg, country: Germany incidence: 2/10, conc. range: 270-900 μ g / kg, Ø conc.: 596 μ g / kg, country: Poland \rightarrow alternariol methyl ether incidence: 105/339, conc.: nc, country: Austria

incidence: 33/156, conc. range: 5-95 μ g / kg, Ø conc.: 28.9 μ g / kg, country: Germany incidence: 5/10, conc. range: 450-750 μ g / kg, Ø conc.: 437 μ g / kg, country: Poland \rightarrow deoxynivalenol incidence: 6/6, conc. range: 1-6300 μg/kg, country: Finland incidence: 21/21, conc. range: 7-861 μ g / kg, Ø conc.: 168 μ g / kg, country: Finland incidence: 3/8, \emptyset conc.: 60 µg/kg, country: Germany incidence: 1/2, conc.: 365 µg/kg, country: Germany incidence: 3/37, conc. range: 200-700 μ g / kg, Ø conc.: 500 μ g / kg, country: Germany incidence: 7/7*, conc. range: 70-90 μ g / kg, Ø conc.: 270 μ g / kg, country: Germany, *moldy incidence: 4/10, Ø conc.: 200 µg/kg, country: Germany incidence: 11/72, conc. range: 20-500 μg/kg, country: Germany incidence: 2/3, conc. range: \leq 80 µg/kg, country: New Zealand incidence: 3/6, conc. range: 420-520 μ g / kg, Ø conc.: 470 μ g / kg, country: Sweden incidence: 11/32, conc. range: 40-260 μ g/kg, Ø conc.: 140 μ g/kg, country: Sweden incidence: 14/45, conc. range: 40-500 μ g/kg, Ø conc.: 200 μ g/kg, country: Sweden incidence: 1/6, conc.: $< 100 \mu g / kg$, country: UK incidence: 1/1, Ø conc.: 31 µg/kg, country: USSR → diacetoxyscirpenol incidence: 3/6, conc. range: 10-1700 μg/kg, country: Finland \rightarrow fusarenon X incidence: 1/6, conc.: $< 40 \ \mu g / kg$, country: Finland

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Oats

 \rightarrow HT-2 toxin incidence: 1/6, conc.: $< 80 \ \mu g / kg$, country: Finland incidence: 2/21, conc. range: 33-44 μ g / kg, Ø conc.: 39 μ g / kg, country: Finland incidence: 18/68, conc. range: \leq 700 μg/kg, country: Germany incidence: 10/107, conc. range: 300-900 μg/kg, country: Germany incidence: 18/59, conc. range: 100-700 μg/kg, country: Germany \rightarrow nivalenol incidence: 2/6, conc. range: < 1000 μg/kg, country: Finland incidence: 3/21, conc. range: 48-83 μ g / kg, Ø conc.: 70 μ g / kg, country: Finland incidence: 1/8, conc.: 1464 µg/kg, country: Germany incidence: 4/7, conc. range: 16 µg/kg, country: Nepal incidence: 2/3, conc. range: \leq 610 µg/kg, country: New Zealand incidence: 1/1, conc.: 1100 µg/kg, country: USSR \rightarrow moniliformin incidence: 3*/3, conc. range: 15,700-38,300 μg/kg, Ø conc.: 24,060 μg/kg, country: Poland, * hand-selected, visible fungal damage → monoacetoxyscirpenol incidence: 1/107, conc.: 50 µg/kg, country: Germany → neosolaniol incidence: 2/107, conc. range: 300-400 μ g/kg, Ø conc.: 350 μ g/kg, country: Germany \rightarrow ochratoxin A incidence: 23/48, conc. range: 5-1000 μg/kg, country: Austria incidence: 1/1, conc.: ca. 1700 μ g/kg, country: Canada incidence: 1/19, conc.: 1-2 µg/kg, country: Czechoslovakia incidence: 21/50*, conc. range: 0.05-4.9 μ g / kg (20 samples), 5.6 μ g / kg (1 sa),

 \emptyset conc.: 0.5 μ g/kg, country: Denmark, *conventional incidence: 6/17*, conc. range: 0.05-4.2 μ g/kg, Ø conc.: 0.3 μ g/kg, country: Denmark, *ecological incidence: 13/25*, conc. range: 0.05-4.6 μ g / kg, Ø conc.: 0.5 μ g / kg, country: Denmark, *conventional, imported incidence: 12/93, conc. range: 1-58.8 μ g / kg, Ø conc.: 9.5 μ g / kg, country: Germany incidence: 2/34, conc. range: 1.4-56.6 μ g / kg, Ø conc.: 29 μ g / kg, country: Italy incidence: 2/14, conc.: \leq 2.4 µg/kg, country: The Netherlands incidence: 4/18, conc. range: 0.1-2.4 μ g / kg, country: The Netherlands incidence: 17/24, conc.: \leq 3.8 µg/kg, Ø conc.: 0.95 µg/kg, country: UK incidence: 1/46, conc.: 80 µg/kg, country: UK incidence: 2/28, conc.: 52-110 μg / kg, Ø conc.: 81 µg/kg, country: USA \rightarrow T-2 toxin incidence: 1/6, conc.: $< 24 \mu g / kg$, country: Finland incidence: 11/19, conc. range: 1-160 μ g / kg, Ø conc.: 27 μ g / kg, country: Finland incidence: 2/21, conc. range: 45-73 μ g / kg, Ø conc.: 59 μ g / kg, country: Finland incidence: ?, conc. range: 10-90 µg/kg, country: Germany incidence: 16/49, conc. range: 10-50 μ g / kg, Ø conc.: 300 μ g / kg, country: Germany incidence: 7/10, conc. range: 13-500 μ g / kg, Ø conc: 220 μ g / kg, country: Germany incidence: 15/82, conc. range: 70-300 μg/kg, country: Germany T-2 triol incidence: 3/66, conc. range: 100-300 µg/kg, country: Germany

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 \rightarrow zearalenone incidence: 3/21, conc. range: 30-86 μ g / kg, Ø conc.: 63 μ g / kg, country: Finland incidence: 3/8, \emptyset conc.: 49 µg/kg, country: Germany incidence: 1/2, conc.: 41 µg/kg, country: Germany incidence: 22/144, conc. range: 1-150 µg/kg, country: Germany incidence: 6/7*, conc. range: $\leq 8 \ \mu g / kg$, \emptyset conc.: 3 µg/kg, country: Germany, *damaged kernels incidence: 4/10, Ø conc.: 50 μg/kg, country: Germany incidence: 17/80, conc. range: 10-440 μg/kg, country: Germany incidence: 1/5, Ø conc.: 2 µg/kg, country: Italy incidence: 5/7, Ø conc.: 6 µg/kg, country: Nepal incidence: 10/29, conc. range: \leq 90 μg/kg, country: New Zealand \rightarrow cereals

Ochratoxicosis This worldwild-occurring \rightarrow mycotoxicosis, due to the intake of \rightarrow ochratoxin A, is primarily a problem in temperate climates of such countries as Canada, Denmark, Ireland, Norway, Sweden, and the US. Pigs and \rightarrow poultry are mainly affected but humans may also suffer from this disease (\rightarrow Balkan endemic nephropathy).

The occurrence of the \rightarrow mycotoxic porcine nephropathy in Denmark is linked with apparently "extreme climatic conditions" such as high moisture and relatively high temperatures (about 25 °C). These conditions favor the growth of ochratoxin producing fungi like \rightarrow Aspergillus ochraceus group and \rightarrow Penicillium verrucosum Dierckx.

 \rightarrow nephropathy

Ochratoxin A (Abbr.: OTA) is a N-[[(3R)-5-chloro-3,4-dihydro-8-hydroxy-3methyl-1-oxo-1H-2-benzopyran-7-yl]car-

bonyl]-L-phenylalanine which belongs to the isocumarins having an amide linkage to L-phenylalanine (\rightarrow mycotoxins). Similar to \rightarrow aflatoxin B₁ it contains a lactone group but the Cl-atom is striking for a natural substance (see Figure 1 Ochratoxin A). It was first isolated from \rightarrow Aspergillus ochraceus K. Wilh. in 1965 by African scientists during laboratory screening for toxigenic fungi. OTA was found to occur naturally for the first time in an US \rightarrow maize sample in 1969. In contrast to South Africa this mycotoxin caused economically important animal diseases and possibly also a human disease in other, northern parts of the world due to the contamination of \rightarrow food and feedstuff.

CHEMICAL DATA

Empirical formula: C₂₀H₁₈O₆NCl, molecular weight: 403

FUNGAL SOURCES

In tropical and semitropical regions OTA is mainly produced by members of the \rightarrow Aspergillus ochraceus group. \rightarrow Aspergillus alutaceus var. alutaceus Berkely & Curtis (formerly \rightarrow A. ochraceus K. Wilh.) is the best known ochratoxin producer of the genus \rightarrow Aspergillus, e.g. A. melleus, A. sclerotiorum and A. sulphureus are of minor importance. Their incidence in food is rare. In temperate regions \rightarrow Penicillium verrucosum Dierckx predominates. Further producers: A. niger group, \rightarrow Eurotium herbariorum, \rightarrow Penicillium spp. (e.g. P. purpurescens), \rightarrow Petromyces alliaceus

NATURAL OCCURRENCE

→ almonds, → baby food, → bacon, → bakery products, → barley, → barley malt, → beans, → beer, → beer, wheat, → biscuits, → bran, → bread, → breakfast cereals, → breakfast drinks, → buckwheat, → cardamom, greater, → cassava flour, → cereal flakes, → cereal food, → cereal products, → cereals, → chapatti, → cheese, → cheese, Bhutanese,

 \rightarrow cheese, Cheddar, \rightarrow cheese, Chesire, \rightarrow cheese, Double Gloucester, \rightarrow cheese, Edam, \rightarrow cheese, Emmental, \rightarrow cheese, Leicester, \rightarrow cheese, Wensleydale, \rightarrow cheese, cake, \rightarrow cheese trimmings, \rightarrow chicken, \rightarrow chicken, yolk, \rightarrow chilli pickles, \rightarrow chilli powder, \rightarrow chilli sauce, \rightarrow cocoa beans, \rightarrow cocoa nibs, \rightarrow cocoa presscake, \rightarrow cocoa products, \rightarrow coffee beans, \rightarrow coffee, \rightarrow confectionery, \rightarrow copra, \rightarrow coriander, \rightarrow corn flakes, \rightarrow cow peas, \rightarrow curry, \rightarrow curry paste, \rightarrow duck, \rightarrow fennel, \rightarrow figs, \rightarrow fish, \rightarrow flour, \rightarrow foods, \rightarrow garlic pickle, \rightarrow ginger, \rightarrow goose, \rightarrow grains, \rightarrow grape juice, \rightarrow ham, \rightarrow hazelnuts, \rightarrow human breast milk, \rightarrow kulen, \rightarrow lentils, \rightarrow maize, \rightarrow maize flour, \rightarrow maize grits, \rightarrow maize products, \rightarrow majoran, \rightarrow malt, \rightarrow meat, \rightarrow milk, \rightarrow millet, \rightarrow muesli, \rightarrow muesli ingredients, \rightarrow noodles, \rightarrow nuts, \rightarrow oats, \rightarrow oat bran, \rightarrow oat flakes, \rightarrow olive oil, \rightarrow olives, \rightarrow paprika, \rightarrow peanuts, \rightarrow peas, \rightarrow pepper, \rightarrow pig blood, \rightarrow pig kidneys, \rightarrow pig liver, \rightarrow pig serum, \rightarrow pop corn, \rightarrow pork, \rightarrow porridge, \rightarrow poultry, \rightarrow rice, \rightarrow rice bran, \rightarrow rice cake, \rightarrow rye, \rightarrow rye bran, \rightarrow rye flour, \rightarrow rye grits, \rightarrow sausages, \rightarrow sesame seeds, \rightarrow sesame oil, \rightarrow snack food, \rightarrow soybean, \rightarrow soybean concentrate, \rightarrow spelt, \rightarrow spices, \rightarrow sunflower seeds, \rightarrow tandoori, \rightarrow tapioca, \rightarrow triticale, \rightarrow turkey, \rightarrow vegetables, \rightarrow wheat, \rightarrow wheat grits, \rightarrow wheat products, \rightarrow wine, \rightarrow zwieback OTA occurs widely in plants and plant products but most frequently in cereal grains infected with P. verrucosum, particularly in north temperate growing areas. Compared to pre-harvest production, post-harvest OTA formation is regarded as the predominant factor in the contamination of insufficiently dried starch-rich foodstuffs (cereals and derived products). It seems that the distribution of OTA in food and / or crops resembles that of aflatoxin with respect to inhomogenicity.

OTA contamination of wines (up to 0.4 μ g/l) from southern parts of Europe may be responsible for increased OTA levels found in the blood of males in southern Switzerland . Grape juice samples may also be contaminated ($\approx 0.2 \,\mu g \, \text{OTA} \, / \, \text{l}$). Although infection of \rightarrow meat and \rightarrow fish with P. verrucosum (and possible mycotoxin formation) has been reported, contamination of meat products is more usually due to the \rightarrow carry over of OTA from contaminated animal feed into blood, kidneys and muscles. Since OTA is extensively metabolized in the forestomachs by protozoan and bacterial enzymes to nontoxic metabolites, tissues of ruminants are not contaminated to any significant extent. Even at higher concentrations the rapid hydrolysis of OTA greatly impedes absorption and may cause only a transient suppression of \rightarrow milk production in cattle. It was estimated that the application of at least 1.66 mg OTA/kg bw for four days is necessary to detect any residues of OTA in the milk. Therefore, OTA levels commonly found in P. verrucosum-contaminated feeds do not represent a substantial health risk to these animals. However, significant contamination in a number of tissues of single-stomach food animals (e.g. pigs and poultry), especially the kidneys, due to carry over from feed is possible. These animals belong to the group of susceptible monogastric livestock showing nephropathy. \rightarrow Pork and \rightarrow bacon as well as pork-derived meat products (e.g. \rightarrow sausages, black pudding) may contain higher amounts of ochratoxin. A high incidence of ochratoxin A in swine blood samples was related to a high moisture content in barley (main ingredient of swine feed). In poultry muscles 29 µg OTA / kg have been found at slaughter under natural conditions but in general liver and kidney typically contain the highest residues. However, OTA residues in tissues

decrease rapidly following removal of the contaminated diet.

It is suggested that for humans the bioavailability for OTA residues is higher in cereals than in meats, as in the latter OTA is bound to proteins.

In the blood, ochratoxin A is present bound to serum albumin and in its free form. Particularly in humans, \rightarrow cattle and pigs, OTA is strongly bound to serum albumin. \rightarrow Human breast milk may also be contaminated with OTA (see Figure 2 Ochratoxin A).

TOXICITY

Clinical symptoms: emetic, strong nephrotoxic, hepatotoxic, \rightarrow immunosuppressive, \rightarrow teratogenic, \rightarrow mutagenic, cancerogenic

LD₅₀ (po): 20-22 mg/kg bw rats In all the mammalian species tested, the kidney is the major target for the toxicity of OTA. Besides the \rightarrow Balkan endemic nephropathy elevated exposure to OTA should also be associated with human nephropathies in Algeria and Tunisia. Furthermore, in rural Scandinavian populations high kidney failure rates have been observed which may be due to the ingestion of pig meat contaminated with excessive amounts of OTA. The fact that the half-life of OTA in humans is 8-12 times longer than in rats is important for risk assessment. Since this mycotoxin is fat soluble and not readily excreted, accumulation in fatty tissues occurs. Decreased weight gains in swine and poultry as well as losses in egg production occur at levels higher than 2000 μ g/kg. Higher dosis are often fatal. However, cattle are resistant to the OTA levels found naturally in feed. The primary source of excretion is the urine (rats) but faecal excretion also occurs to some extent. In experimental animals a synergistic effect between OTA and citrinin as well

as \rightarrow penicillic acid has been observed.

Detection

ELISA, HPLC (fluorescence detection), LC-MS, RIA, TLC

Possible Mycotoxicosis

 \rightarrow Mycotoxic porcine nephropathy, Balkan endemic nephropathy

FURTHER COMMENTS

It seems that cereals and cereal products are the main contributors to OTA intake in northern climates (Europe). There are no large uncertainties about the consumption data of this kind of food since they are regularly eaten by most people. In addition, roasted coffee, beer, pig meat, blood products, wine, and pulses may contribute to the intake of OTA. Because of the particularly strong binding of ochratoxin A to serum albumin of pig blood, products like black pudding are most likely to contain ochratoxin A. Estimations revealed the following mean daily OA intakes for adults: Germany = 1.2 and 1.3 ng/kg bm for women and men, respectively, Sweden = 0.4 ng/kgbm, Swiss = 0.7 ng/kg bm men (residing) north of the Alpes), Canada = 1.1 ng/kgbm for males (12-19 years). Due to the fact that OTA occurs in blood at a much higher rate than the frequency that nephropathy has detected, the analysis of swine blood residue levels may be a more suitable indicator of low amounts, or early exposition to the toxin. The use of this analytic technique allows the essential elimination of OTA residues from the kidneys and / or carcasses that have been fed on an OTA-free diet for a period of four weeks before slaughter. In this way the entry of contaminated meat into the food chain may be prevented. OTA possesses a relatively long half-life in certain edible animal species as well as in humans (35 days in serum). This explains the high incidence of OTA in human blood sera (but generally at low levels). It further documents the potential widespread occurrence of OTA in European food, particularly in whole-grain breads, \rightarrow pork and pig-blood-based products.

OTA is often accompanied by \rightarrow citrinin and the naphthoquinones viomellein and xanthomegnin (all nephrotoxic) which are products of \rightarrow Penicillium aurantiogriseum Dierckx.

Production: Minimum $\rightarrow a_w$ for ochratoxin production is a_w 0.85 A. ochraceus and aw 0.83 P. verrucosum with an optimum at > a_w 0.97. At the optimum a_w OTA is produced in a temperature range of 12-37 °C (optimum 25 °C) for A. ochraceus and 4-31 °C (optimum 24 °C) for P. verrucosum. On bread (pH 5.6) the minimum aw for OTA prodcution amounted to a_w 0.80, the optimum was a_w 0.92 $(\rightarrow$ Penicillium viridicatum Westling ? = P. verrucosum). The optimum pH for ochratoxin A production under in vitro conditions is \approx pH 5.6 which is the same for both species (A. sulphureus = pH 6.0-6.3).

In general the medium composition, especially the presence or absence of micronutrients such as metal ions, more strongly influenced biosynthesis of OTA than growth. Groundnuts and soybeans were the optimal substrata for OTA production of *A. alutaceus* whereas *P. verrucosum* produced highest yields on maize and wheat.

Levels of nitrogen applied to growing barley increased the protein content as well as the ochratoxin production of *A. ochraceus* and *P. verrucosum* on barley postharvest.

Subinhibitory concentrations of phosphine may increase the levels of ochratoxin produced whereas ochratoxin biosynthesis is inhibited by dichlorvos at concentrations which have relatively little effect on fungal growth. Growth and ochratoxin production by *A. sulphureus* and *P. verrucosum* are inhibited due to antimicrobial food additives like methyl paraben, sodium propionate, and potassium sorbate.

Irradation (\leq 10 kGy) enhanced OTA production by A. ochraceus. **Stability:** Compared to \rightarrow patulin or penicillic acid OTA appears to be more stable in foods but it is probably somewhat less stable than \rightarrow aflatoxins. Once ochratoxin A has been formed in a food this moderatly stable mycotoxin survives most food processing stages (such as cooking, roasting, fermenting) to quite appreciable degree. Even temperatures as high as 250 °C are not sufficient for complete degradation of OTA. Losses of \approx 20% occurred during frying of bloodpudding, kidneys, and muscular tissue from pigs. No losses were observed in adipose tissue after frying.

Frying (150-160 °C) of certain pig products such as ground muscle, fat, diced kidneys, and sliced blood pudding caused losses in total toxin of about 20% but in frozen pig kidney a high stability of OTA was observed. Cooking of animal products was less effective in the destruction of OTA.

No destruction of OTA occurred during bread baking but OTA levels partially decreased after biscuit baking. Soaking, blanching, cooking or canning of beans generally resulted in only small losses of OTA (10-34%). Cooking of wheat was also ineffective.

Regarding the stability of OTA, the roasting of coffee gave the most variable results. Losses were reported in the range of 0-100%. This may be due to several factors such as roasting conditions, inhomogenicity of natural coffee bean contamination, OTA levels, natural contamination versus spiking and analytical method performance. There are different opinions among researchers whether OTA passes into brewed coffee. The overwhelming majority of more than 600 European coffee samples did not show any OTA contamination while only a few samples contained more than 30 μ g/kg.

OTA can slowly break down merely during storage of grains and grain products as is documented by a decrease of more than 60% in naturally contaminated barley over a storage period of 2 years.

Depending on the temperature and possibly other factors the moisture can increase or decrease the stability of ochratoxin A during the heating of cereals.

OTA was moderately stable during drying of sausages.

Storage of cheese at room temperature caused a significant decrease in OTA concentration.

It is suggested that the mechanical removal of OTA is probably the most feasible procedure.

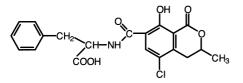
Cleaning / milling: Neither cleaning (dry or wet) nor milling did eliminate OTA from naturally-contaminated samples of barley and wheat. After milling similar levels of OTA were found in flour and \rightarrow bran. However, scouring (removal of the outer layers of the pericarp) as an additional cleaning procedure reduced the OTA concentration in wheat by as much as 50%.

During experimental wet-milling of maize the maize bits (starch, fibre and gluten) contained most of the OTA (51%) of all the maize fractions.

Malting / brewing: There are different results concerning the sources of OTA contamination in beer. OTA appears to be completely destroyed or lost during malting (mainly in the initial steeping stage) of moderately contaminated barley lots. Besides malting (malt mash but not the cooker mash) boiling of the wort with hops, and the final fermentation should also contribute to the destruction of OTA, possibly to ochratoxin α . A transmission rate of 2-28% of the ochratoxin in barley into beer was observed, if heavily contaminated lots are used for malting. However, such severely deteriorated barley brands would probably not be accepted for brewing. Contamination of beer is mainly due to the use of contaminated cereal adjuncts since OTA survives the fermentation step in beer-making. About 20-30% of the original OTA concentration may be found in the finished product.

Control: Proper storage of harvested grains (moisture content and temperature are most important factors) prevents growth of saprophytic storage fungi and subsequent OTA contamination. Since OTA is transmitted into animal tissue, particularly in pigs and poultry, no contaminated feeds should be fed to animals intended for human consumption. In general, only good practices at all stages of the food chain and approaches based on HACCP concepts will contribute to keep OTA contamination low in foodstuffs.

Regulations: The IARC has classified ochratoxin A as a possible human carcinogen (Group 2B) in 1993 based on sufficient evidence in humans. An acceptable safe level of the tolerable daily intake would fall in the range of 1.5-4.2 ng/kg bw/day. The Word Health Organization/ Food and Agricultural Organization Joint Expert Committee on Food Additives (JECFA) recently re-evaluated the toxicity of OTA. A PTWI of 100 ng/kg, bodyweight/week was determined. Among 77 countries with known mycotoxin regulations eight (Czech Republic, Denmark, France, Greece, Hungary, Sweden, Switzerland, Uruguay) also have specific regulations for ochratoxin A levels in one or more commodities whereas some countries have proposals for ochratoxin A regulations (Austria, Germany, Great Britain, Rumania, The Netherlands). Current (proposed) limits for OTA contamination are as follows: 1-5 μ g/kg children and infant foods, 2-50 μ g / kg foods, 5-300 μ g / kg animal feeds.



Ochratoxin A (Figure 1)

The proposed tolerance levels in the EU are 1 μ g/kg infant foods, 5 μ g/kg cereals.

Ochratoxin B is the dechloro-analogue of ochratoxin A (N-[[(3R)-3,4-dihydro-8-hydroxy-3-methyl-1-oxo-1H-2-benzo-pyran-7-yl]carbonyl]-L-phenylalanine) which was first isolated in 1965 from \rightarrow Aspergillus alutaceus var. alutaceus Berkely & Curtis (\rightarrow mycotoxins) (see Figure Ochratoxin B).

CHEMICAL DATA

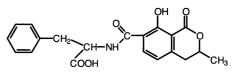
Empirical formula: $C_{20}H_{19}O_6N$; molecular weight: 369

FUNGAL SOURCES A. alutaceus var. alutaceus

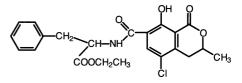
NATURAL OCCURRENCE \rightarrow bread, \rightarrow maize

TOXICITY

Ochratoxin B is approximately 16 times less toxic to chicks than \rightarrow ochratoxin A and also less toxic than ochratoxin C. However, similar pathological lesions occurred in chicks and rainbow trout as described for ochratoxin A.



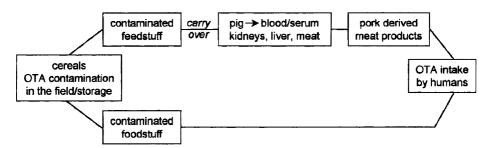
Ochratoxin B



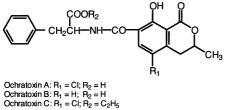
Ochratoxin C

Detection TLC

Ochratoxin C (Syn.: Ochratoxin A ethyl ester) Isolation methods which depend on the free carboxyl group are not successful and therefore the occurrence of this ochratoxin type may be underestimated (N-[[(3R)-5-chloro-3,4-dihydro-8-hydroxy-3-methyl-1-oxo-1H-2-benzo-pyran-7-yl]carbonyl]-L-phenylalanine; ethyl ester). It is produced by \rightarrow Aspergillus alutaceus var. alutaceus Berkely & Curtis and was first isolated in 1965. Natural occurrence of ochratoxin C ($C_{22}H_{22}O_6NCl$; mw 431) in \rightarrow wine has been reported (see Figure Ochratoxin C).



Ochratoxin A (Figure 2). Routes of OTA intake by humans



Ochratoxin C: $R_1 = CI; R_2 = C_2R_5$ Methylester of Ochratoxin A: $R_1 = CI; R_2 = CH_3$ Methyl or ethyl ester of Ochratoxin B: $R_1 = H; R_2 = CH_3$ or C_2H_5

Ochratoxins. Members of the ochratoxin group

Ochratoxins are isocoumarines composed of a 3,4-dihydroy-3-methylisocoumarin moiety linked via the 7-carboxy group to L- β -phenylalanine by an amide bond (\rightarrow mycotoxins). The isolation of a chlorine-containing metabolite designated \rightarrow ochratoxin A succeeded in 1965 when African scientists carried out a screening of toxigenic fungi. OTA as the major toxic principle in different kinds of food and feedstuff (mainly \rightarrow grains) is the most important toxic member of a group of nine or more \rightarrow ochratoxins produced with the highest yield. This group consists of ochratoxin A, its methyl and ethyl esters (all which are toxic), and 4-hydroxyochratoxin A (see Figure Ochratoxins). In contrast, the chlorine free derivative of OTA, ochratoxin B as well as ochratoxin C, is rarely found in \rightarrow foods and feeds. Contamination of grains with ochratoxins has been reported from e.g. most European countries and North America and is due to \rightarrow Aspergillus ochraceus group and \rightarrow Penicillium verrucosum Dierckx. OTA typically co-occurs with low amounts of \rightarrow citrinin, which is also a \rightarrow nephrotoxin.

Ogbono is a Nigerian type of foodstuff made from the plant *Irvingia gabunensis*. Ogbono may contain the following \rightarrow mycotoxins: aflatoxin B (\rightarrow aflatoxins) incidence: 1/1, conc.: 168 µg/kg, country: Nigeria **Ogili-ugba** is a Nigerian type of foodstuff made from the castor bean, *Riccinus communis*. Ogili-ugba may contain the following \rightarrow mycotoxins: aflatoxin B (\rightarrow aflatoxins)

incidence: 1/1, conc.: 362 μg/kg, country: Nigeria

Ogoro is a Nigerian indigenous beverage (palm juice) made from the sap from the stalk of the male inflorescence or the immature shoot of the oil palm (*Elais guinensis*). Ogoro may contain the following \rightarrow mycotoxins: aflatoxin B (\rightarrow aflatoxins) incidence: 2/2, conc. range: 116-118 µg/kg, Ø conc.: 117 µg/kg, country: Nigeria

Oil If the oil is removed from the \rightarrow oil seeds, \rightarrow aflatoxins are mainly found in the oil seed meal. The soap stock as a by-product from the alkali-refining step contains only the low levels that remained in the crude vegetable oil. In general, the refined oil is aflatoxin-free since aflatoxin residues are removed in the bleaching refining steps.

Oil may contain the following \rightarrow mycotoxins:

aflatoxin (no specification)

incidence: 10/25*, conc. range: ≤ 7

 μ g/kg, Ø conc.: 3 μ g/kg, country: Philippines, *cooking

aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂)

incidence: 1/4*, conc.: 0.1 µg/kg, country: UK, *chili, almond

 \rightarrow zearalenone

incidence: 1/4*, conc.: 5.4 μ g/kg, country: UK, *chili, almond \rightarrow coconut oil, \rightarrow olive oil, \rightarrow peanut oil,

 \rightarrow sunflower seed oil

Oil seed rape represents an important agricultural crop which is used as cook-

ing oil and for the production of margarine. Rapeseed meal is also used in cattle concentrates. Weather conditions during harvesting and threshing show extreme variations, enabling the development of different fungi especially if oilseed rape is stored under poor conditions. Subsequent mycotoxin contamination might occur. Oilseed rape may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/20, conc.: 0.25 µg/kg, country: Spain \rightarrow citrinin incidence: 1/1*, conc.: 4100 µg/kg, country: UK, *moldy \rightarrow sterigmatocystin incidence: 1/1*, conc.: 40 µg/kg, country: UK, *moldy \rightarrow viomellein incidence: 1/1*, conc.: traces, country: UK, *moldy vioxanthin incidence: 1/1*, conc.: 40 µg/kg, country: UK, *moldy \rightarrow xanthomegnin incidence: 1/1*, conc.: traces, country: UK, *moldy

Oil seeds (no specification) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 31/73*, conc. range: < 5-2000 μ g / kg, country: Natal (Union of South Africa), *includes \rightarrow peanuts, \rightarrow sunflower seeds, cottonseeds \rightarrow aflatoxin (no specification) incidence: 9/80, conc. range: 2-20 µg/kg (7 samples), > 20 μ g / kg (2 sa), country: Uruguay \rightarrow patulin incidence: 8/107*, conc.: nc, country: South Africa, *includes peanuts, sunflower seeds, cottonseeds → zearalenone incidence: 1/107, conc.: nc, country: South Africa

incidence: 6/64, conc. range: 100-200 μ g/kg (3 samples), > 200 μ g/kg (3 sa), country: Uruguay

Olive oil In some countries farmers sometimes store their \rightarrow olives for several weeks under conditions that contribute to the growth of molds such as \rightarrow Aspergillus flavus Link and the \rightarrow Aspergillus ochraceus group. This may result in aflatoxin and ochratoxin contamination of olives and olive \rightarrow oil. If the so called "virgin" olive oil is prepared from contaminated crude oil, the refining process which would remove the \rightarrow aflatoxins is omitted.

Nonchemically treated olive oil made from deteriorated olives may contain low levels of \rightarrow Alternaria mycotoxins. These low levels should not represent a concern for human health.

The transmission rate from olives into the oil amounted to only $4\% \rightarrow$ alternariol methyl ether (793.6 µg / kg) and 1.8% \rightarrow alternariol (285.7 µg / kg). No transmission has been reported for \rightarrow altenuene and \rightarrow tenuazonic acid, considering an oil yield of 15% from the processed olives (experimental study).

The results of a limited survey showed that olive oil samples collected from different oil mills did not show any mycotoxin contamination.

However, olive oil may contain the following \rightarrow mycotoxins:

 \rightarrow aflatoxin B₁

incidence: 3/46, conc. range: 1-13 μ g/kg, \emptyset conc.: 11 μ g/kg, country: Germany

incidence: 14/16, conc. range: 1-75

 μ g/kg, Ø conc.: 361 μ g/kg, country: Greece

 \rightarrow aflatoxin B₂

incidence: 10/16, conc. range: 1-55 μ g/kg, Ø conc.: 185 μ g/kg, country: Greece

→ aflatoxin G₁ incidence: 5/16, conc. range: 1-2.5 µg/kg, \emptyset conc.: 1.6 µg/kg, country: Greece → aflatoxin G₂ incidence: 5/16, conc. range: 1-5 µg/kg, \emptyset conc.: 2.2 µg/kg, country: Greece → ochratoxin A incidence: 3/60, conc. range: traces, country: Morocco → coconut oil, → oil, → peanut oil

Olives Physical damage of the surface seems to be a prerequisite for \rightarrow Alternaria mycotoxins to contaminate olives. Such olives are frequently infected with \rightarrow Alternaria spp., mainly \rightarrow Alternaria alternata (Fr.) Keissler. Under suitable conditions fungal attack starts with the penetration into the fruit pulp, followed by substantial mycelial growth and subsequent mycotoxin formation. Considerable amounts of Alternaria \rightarrow mycotoxins may be produced in physically damaged (heavily damaged, weathered or moldy) olives in the field before harvesting as well as during storage. Although Alternaria spp. could be isolated from sound, undamaged olives properly harvested from the ground in different areas, no mycotoxin contamination could be established.

Olives may contain the following myco-toxins:

 \rightarrow aflatoxin B₁

incidence: $12/103^{**}$, conc. range: 5-37 µg/kg, country: Morocco, **black, Greek-style → altenuene incidence: $1/4^*$, conc.: $1400 \mu g/kg$, country: Italy → alternariol incidence: $4/4^*$, conc. range: 109-2320

 μ g/kg, Ø conc.: 1120 μ g/kg, country: Italy

→ alternariol methyl ether incidence: 4/4*, conc. range: 30-2870 μ g / kg, Ø conc.: 818 μ g / kg, country: Italy → ochratoxin A incidence: 5/103**, conc. range: 40-80 µg/kg, country: Morocco, **black, Greek-style incidence: 4/7, conc. range: 0.3-46,830 µg/kg, country: Tunesia → tenuazonic acid incidence: 2/4*, conc.: 109-262 µg/kg, Ø conc. 1865 µg/kg, country: Italy, *samples visibly affected by *Alternaria* rot

Onions → garlic

Ontjom Processing of ontjom might result in detoxication of mycotoxin $(\rightarrow \text{ mycotoxins})$ contaminated $\rightarrow \text{ peanuts}$.

Onyalai This neither heritable nor infectious disease was first described in Angola (1904) and is widespread in the south of the Sahara. It most frequently occurs in summer. In general, individuals of all ages and both sexes of African races (e.g. Bantu) are almost exclusively affected although some cases in Europeans and Chinese have also been reported.

A distinct feature of this disorder is the sudden appearance of hemorrhagic $(\rightarrow \text{hemorrhage})$ bullae $(\rightarrow \text{bulla})$ in the mouth and sometimes on the skin. Further symptons are \rightarrow hematuria, profuse bleeding from the nose, mouth, and conjunctiva. Severe cases are characterized by hemorrhagic shock and cerebral hemorrhages. Death may occur within a few days of the onset of the disease. A mortality rate of 14% in one case study has been reported. However, first-time sufferers may recover spontaneously within two months of the onset. The severity and mortality of this disease depend on the area of its occurrence. Although the cause of onyalai is still unknown, \rightarrow mycotoxins such as

Onyalai

 \rightarrow tenuazonic acid (salts) and \rightarrow moniliformin should be involved. The mycotoxin hypothesis is due to the fact that the disease occurs only among eaters of \rightarrow millet (*Pennisetum typhoides*). This millet as well as \rightarrow sorghum was contaminated by \rightarrow Fusarium spp. and highly toxic \rightarrow Phoma spp. From *P. sorghina*inoculated \rightarrow maize culture calcium-, magnesium-, and sodium-tenuazonate were isolated. Acid treatment resulted in tenuazonic acid, which is also produced by certain \rightarrow Alternaria species. However, the suggested primary toxic actions of tenuazonic acid (emetic and cardiovascular action) are not consistent with the characteristic hemorrhagic bullae in the oral cavity due to onyalai. In addition, the ability to selectively complex with trace metals in vivo is not sufficient to explain the symptons of onyalai. Different environmental factors or even other mycotoxins may also be involved. The better nutritional status in the endemic regions is probably one reason for the decrease of onyalai in the last few years.

Oo-hen-mai Japanese: yellow rice (disease)

→ Acute cardiac beriberi

Oranges may contain the following \rightarrow mycotoxins:

→ aflatoxin B₁ incidence: 1/20*, conc.: 4600 μ g/kg, country: Germany incidence: 4/14*, conc. range: 5-50 μ g/ kg, country: Germany → aflatoxin G₁ incidence: 1/20*, conc.: 21.5 μ g/kg, country: Germany → aflatoxin G₂ incidence: 1/20*, conc.: 1200 μ g/kg, country: Germany *moldy → fruits

Oriental fermentations During the manufacturing of fermentation products like \rightarrow miso and \rightarrow shoyu none of the tested industrial used \rightarrow Aspergillus strains (in Japan) produced \rightarrow aflatoxins, \rightarrow ochratoxin A, \rightarrow patulin, \rightarrow penicillic acid or \rightarrow sterigmatocystin. \rightarrow Cyclopiazonic acid was produced by only a few isolates. Although the strains used for fermentation were able to synthesize aspergillic acid, \rightarrow kojic acid, nitropropionic acid and oxalic acid, the concentrations were to low to constitue any toxic hazard to humans.

OTA \rightarrow Ochratoxin A

Oxygen → atmosphere

P

191

Paecilomyces \rightarrow mitosporic fungi, teleomorph: \rightarrow Byssochlamys spp. Byssochlamys spp. and *P. variotii* are important producers of \rightarrow patulin.

Paprika may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A incidence: 1/4, conc.: 40 µg/kg, country: Germany \rightarrow spices

Paralysis Loss of feeling or power to move in any or every part of the body.

Parasiticol (Syn.: \rightarrow aflatoxin B₃)

paretic incomplete paralysis

Parmesan cheese → cheese, Parmesan

Pasta may contain the following \rightarrow mycotoxins: \rightarrow citrinin incidence: 1/2, conc.: 0.5 µg/kg, country: Switzerland \rightarrow ochratoxin A incidence: 11/21, conc. range: < 5 µg/kg, country: UK

Pasteurized foods According to Frisvad (1988) \rightarrow patulin may be excreted into \rightarrow fruit juices and vegetable juices (\rightarrow vegetables).

Pastries may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/86, conc.: < 5 µg/kg, country: Germany \rightarrow citrinin incidence: 1/2, conc.: 0.5 µg/kg, country: Switzerland **Patulin** (Syn.: clavacin, clavatin, claviformin, expansine, gigantic aicd, mycoin, penicidin, tercinin, leucopin) This 4-hydroxy-4-*H*-furo-[3,2-c]pyran-2(6H)-one was first isolated from *Penicillium patulum* (= \rightarrow Penicillium griseofulvum Dierckx) during the search for new antibiotics in 1941 (see Figure Patulin).

CHEMICAL DATA Empirical formula: C₇H₆O₄, molecular weight: 154

FUNGAL SOURCES

→ Aspergillus clavatus Desm., A. giganteus, → Aspergillus terreus Thom, → Byssochlamys nivea, B. fulva, → Eupenicillium spp., → Penicillium expansum Link (most important and the most commonly encountered patulin producer), → Penicillium spp. (e.g. P. claviforme, → Penicillium roquefortii Thom chemotype II, P. melinii)

NATURAL OCCURRENCE

 \rightarrow Apples, \rightarrow apple beverages, \rightarrow apple butter, \rightarrow apple flavor, \rightarrow apple jam, \rightarrow apple juice, \rightarrow apple juice concentrate, \rightarrow apple products, \rightarrow blueberries, \rightarrow cereals, \rightarrow cheese, \rightarrow cheese, goat, \rightarrow cider, \rightarrow cranberries, \rightarrow fruits, \rightarrow fruit juices, \rightarrow fruits products, \rightarrow grape juice, \rightarrow jam, \rightarrow lingonberries, \rightarrow meat, \rightarrow oil seeds, \rightarrow peaches, \rightarrow pear juice, \rightarrow pears, \rightarrow plums, \rightarrow scented supar, \rightarrow soft drinks Apples, apple products, and peaches are excellent substrates for patulin production. In nature patulin is found almost exclusively in apples and apple products but visual inspection will usually identify poor quality items. Patulin contamination of apple juice is an effective indicator of the use of unsound, substandard, P. expansum rotted apples in juice manufacture. Such juices may contain up to 1,000,000 µg/l of patulin. Although patulin commonly occurs in rotting apples and the incidence of patulin contamination of apple juice is fairly high, the level of contamination in general is relatively

M. Weidenbörner, *Encyclopedia of Food Mycotoxins* © Springer-Verlag Berlin Heidelberg 2001 low (< 100 μ g/l). Sporadically very high levels 45,000 μ g patulin/l apple juice from a roadside stand in the USA have been detected. Apples and especially apple products such as juice are the major human dietary sources of patulin. Alcoholic fermentation of fruit juices by *Saccharomyces cerevisia* and *S. ellipsoideus* caused almost total destruction (> 99%) of patulin.

Although potential patulin producers are present on foods such as \rightarrow oranges, oranges juice, wet \rightarrow maize, durum \rightarrow wheat, \rightarrow sorghum, \rightarrow flour, cheeses, meat and meat products (e.g. \rightarrow sausages) no or only decreased levels of patulin have been detected. The lack or decrease is mainly attributed to the reaction (binding) with sulfhydryl groups of compounds (cysteine, gluthatione) present in the \rightarrow foods, although not all contain sulfhydryl groups. This reaction makes patulin chemically undetectable and of lesser toxicity because the binding to functional groups is inhibited. Since contamination of livestock feeds has not been reported, patulin accumulation in meat and poultry products due to \rightarrow carry over seems to be unlikely.

TOXICITY

antibiotic (e.g. *Mycobacterium tuberculosis*), antifungal, \rightarrow immunosuppressive, neurotoxic, \rightarrow teratogenic (?), \rightarrow mutagenic, carcinogenic (?) Gastrointestinal \rightarrow hyperemia, distension, \rightarrow hemorrhage and ulceration LD₅₀ (po): 35 mg/kg bw mice Adducts formed with cysteine possessed a markedly lower toxicity values than patulin itself. The no-observed-effect level (NOEL) for patulin was considered to be 43 µg patulin / kg bw / day after a three times per week administration. In a guideline, the Joint Food and Agriculture Organization/

World Health Organization Committee on

Food Additives (JECFA) lowered the pro-

visional maximum tolerable daily intake (PMTDI) for patulin from a nominal 1 to 0.4 μ g/kg bw/day, based on the calculated NOEL and use of a 100-fold safety factor. Based on the fact that the patulin concentrations in apple juices are usually below 50 μ g/l, the JECFA estimated maximum intakes to be in the order of 0.2 and 0.1 μ g patulin/kg bw/day for children and adults, respectively (WHO 1995).

DETECTION

HPLC (reverse-phase), TLC

FURTHER COMMENTS

Natural patulin contamination is primarily found in apples and apple products. Two facts are responsible. Besides the inactivation of patulin by distinct compounds in certain foods patulin producing molds represent only a low percentage of the total fungal strains isolated from most of the food (1% of the penicillia from flour and bread, 1.42% of the total fungi from European-style dry sausages, 0.9% of the total fungi isolated from corn meal). However, almost 70% (P. expansum) of the isolated fungi from naturally rotted apples produce patulin. Storage of fruits under a controlled atmosphere reduced patulin formation significantly but after evacuation P. expansuminfected fruits show a rapid increase in patulin levels. Diffusion of the toxin into the surrounding tissue has been reported for peaches, pears, and tomatoes but not for apples (up to 1 cm).

Patulin contamination is mainly seen as an indicator of bad manufacturing practices (use of rotten raw materials) although it seems to be only a minor threat to human and animal health.

Although patulin exhibits strong antibacterial activity it was too toxic for all test animals (carcinogenic, mutagenic) to have been used therapeutically. **Production:** Patulin production of *Penicillium* spp. occurs in a temperature range from 0 °C to 31 °C. The limiting a_w amounted to a_w 0.95 (*P. griseofulvum*) and in excess of a_w 0.95 for *P. expansum* in a synthetic medium. The pH optimum for patulin production is between pH 3-6.5 whereas the optimum temperature ranges from 20-25 °C (*P. expansum*). Irradation (15 kGy) increased patulin formation of *P. griseofulvum*.

If the headspace O_2 levels in cans or jars of grape juice are below 0.5%, growth of *Byssochlamys* spp. is significantly reduced and no substantial patulin production can be expected.

Patulin production (up to 50 mg/kg) has been reported in soil under certain circumstances.

Reduction / elimination: An overall 24% decrease in patulin concentration has been observed in pressed apple juice during "down-line" technological production of concentrates. In addition, various chemicals like ascorbic acid, charcoal, sulfur dioxide, vitamin B_1 as well as irradation are suitable for reducing or destroying patulin during "down-line" processing. Besides the inactivation of patulin by sulfhydryl compounds this mycotoxin is also unstable in the presence of alkali. Patulin is more stable at acidic pH whereas temperatures up to 80 °C do not cause a significant reduction.

Peaches may contain the following

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\rightarrow mycotoxins:
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\rightarrow aflatoxin B<sub>1</sub>
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incidence: 9/20*, conc. range: 5-15 μ g / kg, country: Germany, *moldy \rightarrow patulin

incidence: 2/4*, conc. range: 200-400 μ g / kg, Ø conc.: 300 μ g / kg, country: Germany, *stewed, moldy





incidence: 1/8, conc.: 6 μ g / kg, country: Sweden \rightarrow fruits

Peach kernels may contain the following \rightarrow mycotoxins:

→ aflatoxins (AFB₁, AFB₂, AFG₁) incidence: nc, conc.: $\leq 10 \ \mu g / kg$, country: Germany

Peanut brittle may contain the following \rightarrow mycotoxins:

 \rightarrow aflatoxin B₁

incidence: 8/19, conc. range: $0.5-5 \ \mu g/kg$ (5 samples), 6-10 $\mu g/kg$ (1 sa), 11-30 $\mu g/kg$ (1 sa), 142 $\mu g/kg$ (1 sa), country: UK

→ aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 8/19, conc. range: 6-10 μ g/kg (6 samples), 31-100 μ g/kg (1 sa), 190 μ g/kg (1 sa), country: UK

Peanut butter The use of crushed \rightarrow peanuts which are lower in grade than whole peanuts contributes to the aflatoxin contamination of peanut butter. High peak exposure to \rightarrow aflatoxins is reduced by mixing and blending processes. However, average exposure to \rightarrow aflatoxin B₁ which is 60% of the total aflatoxins in peanuts remains the same. Highly effective automatic (electronic) peanut selectors are used in the peanut butter manufacturing process to remove poor-quality nuts. A sorting machine measuring near-infrared transmission spectra allows the dectection of molds in the inner part of shelled peanuts covered with inner skin. Peanut butter may contain the following \rightarrow mycotoxins: aflatoxin B_1 incidence: 64/111, conc. range: $< 5 \mu g / kg$ (36 samples), 10-662 µg/kg (28 sa), country: Germany

incidence: 44/182, Ø conc.: 46 μg/kg, country: Germany

Peanut butter

incidence: 4/4, conc. range: 147-208 μg/kg, country: Germany incidence: 1/1, conc.: 233 µg/kg, country: Germany incidence: 2/2, conc. range: 3.5-5.2 µg/ kg, Ø conc.: 4.4 μ g/kg, country: Germany incidence: 3/4, conc. range: 0.6-1.4 μ g/kg, Ø conc.: 1.3 μ g/kg, country: Japan incidence: 3/6, conc. range: 0.6-2.4 μg/kg, country: Japan incidence: 31/32, conc. range: < 10 μg/kg, country: UK incidence: 10/63, conc. range: 2-20 μ g / kg, Ø conc.: 7 μ g / kg, country: USA \rightarrow aflatoxin B₂ incidence: 2/2, conc. range: 0.5-0.6 μ g / kg, Ø conc.: 0.55 μ g / kg, country: Germany incidence: 3/4, conc. range: 0.1-0.3, Ø conc.: 0.2 µg/kg, country: Japan incidence: 3/6, conc. range: 0.4 µg/kg, country: Japan \rightarrow aflatoxin G₁ incidence: 2/2, conc. range: 3.5-5.2 μ g / kg, Ø conc.: 4.4 μ g / kg, country: Germany incidence: 2/4, conc. range: 0.3 μ g/kg, Ø conc.: 0.3 µg/kg, country: Japan incidence: 3/6, conc. range: 0.1-0.4 μg/kg, country: Japan \rightarrow aflatoxin G₂ incidence: 2/2, conc. range: 1.3-1.7 μ g / kg, Ø conc.: 1.5 μ g / kg, country: Germany aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 64/111, conc. range: $< 5 \mu g / kg$ (36 samples), 7-362 μ g/kg (28 sa), country: Germany incidence: 44/182, Ø conc.: 59 μg/kg, country: Germany incidence: 1/1, conc.: 278 µg/kg, country: Germany incidence: 98/2092, conc. range: 5-19.9 $\mu g / kg$ (95 samples), > 25 $\mu g / kg$ (3 sa), country: Canada aflatoxins (no specification)

incidence: 25/2477, conc. range: 15-30 $\mu g/kg$ (18 samples), 31-60 $\mu g/kg$ (3 sa), 61-90 $\mu g/kg$ (2 sa), 90 $\mu g/kg$ (2 sa), country: Canada incidence: 29/29, conc. range: 30-8600 $\mu g/kg$, country: Philippines incidence: 5/522, conc. range: $\leq 6600 \ \mu g/kg$, \emptyset conc.: 186 $\mu g/kg$, country: Philippines incidence: 17/104, conc. range: $\leq 27 \ \mu g/kg$, \emptyset conc.: 14 $\mu g/kg$, country: USA incidence: 1/3*, conc.: 43 $\mu g/kg$, country: USA, *imported

Peanut butter (crunchy) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 5/16*, conc. range: 2-5 µg/kg (3 samples), 6-10 $\mu g/kg$ (1 sa), 12 $\mu g/kg$ (1 sa), country: UK, *regular incidence: 10/14**, conc. range: 2-5 μ g/kg (2 samples), 6-10 μ g/kg (1 sa), 11-30 µg/kg (1 sa), 31-100 µg/kg (1 sa), $> 100 \le 318 \ \mu g/kg$ (5 sa), country: UK incidence: 7/9**, conc. range: 2-5 µg/kg (5 samples), 6-10 μ g/kg (1 sa), 58 μ g/kg (1 sa), country: UK incidence: 7/15**, conc. range: 6-10 µg/ kg (1 sa), 11-30 μ g/kg (3 sa), 31 \leq 73 μ g/kg (3 sa), country: UK **health food \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 47/59, conc. range: 1-50 µg/kg (32 samples), 51-100 μ g/kg (4 sa), > 100 μ g/kg (11 sa), country: UK incidence: 6/16*, conc. range: 2-5 μ g/kg (4 samples), 6-10 μ g/kg (1 sa), 14 μ g/kg (1 sa), country: UK, *regular incidence: 10/14**, conc. range: 2-5 µg/ kg (1 sample), 6-10 μg/kg (2 sa), 11-30 $\mu g / kg (1 sa), > 100 \le 345 \mu g / kg (5 sa),$ country: UK incidence: 7/9**, conc. range: 2-5 µg/kg (3 samples), 6-10 µg/kg (2 sa), 11-30 μ g/kg (1 sa), 211 μ g/kg (1 sa), country: UK

incidence: 7/15**, conc. range: 6-10 μ g/kg (1 sample), 11-30 μ g/kg (1 sa), 31-100 μ g/kg (2 sa), > 100 \leq 147 μ g/kg (3 sa), country: UK **health food

Peanut butter (smooth) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 7/16*, conc. range: 2-5 µg/kg (6 samples), 7 μ g/kg (1 sa), country: UK, *regular incidence: 4/11**, conc. range: 6-10 µg/ kg (2 samples), $31 \le 49 \ \mu g / kg$ (2 sa), country: UK incidence: 5/6**, conc. range: 11-30 µg/ kg (3 samples), $31 \le 76 \ \mu g / kg$ (2 sa), country: UK incidence: 1/4**, conc: 13 µg/kg, country: UK **health food \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 31/33, conc. range: 1-50 µg/kg (25 samples), 51-100 μ g/kg (4 sa), > 100 μ g / kg (2 sa), country: UK incidence: 7/16*, conc. range: 2-5 µg/kg (6 samples), 8 μ g / kg (1 sa), country: UK, *regular incidence: 6/11**, conc. range: 2-5 µg/kg (1 sample), 6-10 µg/kg (2 sa), 11-30 $\mu g / kg$ (1 sa), $31 \le 85 \mu g / kg$ (2 sa), country: UK incidence: 6/6**, conc. range: 6-10 µg/kg (1 sample), 11-30 µg/kg (1 sa), 31-100 μ g/kg (3 sa), 175 μ g/kg (1 sa), country: UK incidence: $1/4^{**}$, conc.: 27 µg/kg, country: UK **health food **Peanut candy** may contain the following \rightarrow mycotoxins:

 \rightarrow aflatoxins

incidence: 10/18, conc. range: ≤ 20

 μ g/kg, Ø conc: 10 μ g/kg, country: USA

Peanut mix may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins incidence: 1/1, conc.: 302 µg/kg, country: USA

Peanut oil In general, peanut \rightarrow oil does not contain any significant amounts of \rightarrow aflatoxins. These \rightarrow mycotoxins are removed during processing due to the use of solvents or they are destroyed by conventional alkali washing as a part of the refining process. Subsequent bleaching operation further contributes to the elimination of the aflatoxins. Low aflatoxin amounts have been found in crude oils which are not suitable for human consumption. They are obtained by solvent extraction or by hydraulic pressing of ground moldy peanuts. The corresponding meals contained the major portion of the aflatoxins. Peanut oil may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 4/6, conc. range: \leq 0.7 μg / kg, country: India \rightarrow aflatoxin B₂ incidence: 4/6, conc. range: $\leq 0.1 \, \mu g / kg$, country: Japan \rightarrow aflatoxin G₁ incidence: 4/6, conc. range: $\leq 0.1 \, \mu g \, / \, kg$, country: Japan aflatoxin (no specification) incidence: $5/8^*$, conc. range: ≤ 310 μ g / kg, Ø conc.: 246 μ g / kg, country: USA, *crude peanut oil aflatoxins (no specification) incidence: 544/1209, conc. range: 71-5000 μ g / kg, country: India \rightarrow coconut oil, \rightarrow oil, \rightarrow olive oil, \rightarrow peanuts, \rightarrow sunflower seed oil

Peanut paste may contain the following \rightarrow mycotoxins:

Peanut paste

\rightarrow aflatoxins

incidence: 3/4*, conc. range: 11 μ g/kg, Ø conc.: 9 μ g/kg, country: USA, *imported \rightarrow peanuts

Peanut products (no specification) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 13/20*, conc. range: 15-138 μ g / kg, Ø conc.: 64.9 μ g / kg, country: Germany \rightarrow aflatoxin B₂ incidence: 2/20*, conc. range: 3-24 μ g / kg, Ø conc.: 13.5 μ g / kg, country: Germany \rightarrow aflatoxin G₁ incidence: 8/20*, conc. range: 9-44 μ g / kg, Ø conc.: 28 μ g / kg, country: Germany \rightarrow aflatoxin G₂ incidence: 2/20*, conc. range: 4-18 μ g/kg, Ø conc.: 11 μ g/kg, country: Germany, *suspected aflatoxin (no specification) incidence: 1/6, conc.: $2 \mu g / kg$, country: USA \rightarrow aflatoxins (no specification) incidence: 11/32, conc. range: > 30- \leq 220 µg/kg, country: Philippines \rightarrow peanuts

Peanut sauce may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂)

incidence: 18/20, conc. range: 18-943 $\mu g / kg, \emptyset$ conc.: 162 $\mu g / kg$, country: Gambia \rightarrow peanuts

Peanuts (no specification)

From all types of \rightarrow nuts peanuts which grow in the soil are most susceptible to mycotoxin (\rightarrow aflatoxins) contamination. Contamination mainly occurs in the field during the harvest while the nuts are being dried. When harvesting is associated with wet weather conditions a higher contamination rate was established.

Pre-harvesting, harvesting, handling and storage conditions greatly influence the degree of aflatoxin contamination. A soil temperature of between 25.7-27 °C and drought stress during the latter part of the growing season should contribute to aflatoxin contamination in the field before harvest. Although the surface of the pods easily comes into contact with the soil borne \rightarrow Aspergillus flavus Link, pods of intact kernels are difficult to penetrate by the fungus. However, damaged kernels, especially mature ones which contain 30-60% water at the time of harvest, are very susceptible to A. flavus infection. Damages are due to various biotic and abiotic factors: insects (e.g. termites), fungi (e.g. Macrophomina phaseoli, Sclerotium rolfsii), nematodes (e.g. Meloidogyne arenaria), very rapid growth of the peanuts, over-advanced maturity and direct mechanical damage. In addition, pods harvested during the rains showed a high infestation rate while pods harvested during the dry season showed only little infection. Growth of A. flavus in infected peanuts immediately starts after lifting. The optimum moisture content for fungal growth in peanuts is between 10(15)-(25)30% but growth occurs in the range from 9-35%. The minimum $\rightarrow a_w$ for a flatoxin pro-

duction in immature broken peanuts is $a_w 0.83$. Contamination has been observed before digging, after digging and before combining, between combining and drying as well as in storage. The avoidance of preharvest stress in combination with effective drying techniques (moisture content

< 9-10%) and storage conditions (e.g. 32 °C/50% relative humidity, adequate ventilation) immediately after harvest lower or even prevent aflatoxin contamination. During improper storage the total amount of the produced aflatoxins and the ratio of different aflatoxin types is influenced by the temperature. The ratio AFB₁ : AFG₁ is smaller at higher temperatures (35 °C) than at lower temperatures (20 °C). Approximately 60% of the total aflatoxins found in peanuts is aflatoxin B₁. Stored in-shell peanuts having a moisture content > 11% in combination with a relative humidity of at least 84% allow the development of aflatoxin producing fungi.

However, according to the FDA aflatoxin contamination mainly occurs prior to harvest of the peanuts whereas very high kernel moistures may prevent aflatoxin production.

In Brazil the protein-enriched peanut meal and husks as by-products of peanut oil processing are fed to animals. Mycotoxin contamination of these products is harmful and may result in the contamination of \rightarrow meat and \rightarrow milk (\rightarrow carry over).

Wrinkled kernels seem to contain higher aflatoxin concentrations (up to 70 times) than the dark kernels while most of sound mature peanuts do not contain aflatoxin.

There are different ways for decontaminating contaminated peanuts. Roasting reduces (50-70%) but does not eliminate aflatoxin contamination whereas boiling and baking are less effective (20-30%). Microwave and oven roasting caused destructions of \approx 55% AFB₁ and \approx 36% AFG₁.

Peanuts may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/1, conc.: 625 µg/kg, country: Angola incidence: 1/88, conc.: 5 µg/kg, country: China incidence: 3/40, conc. range: 98-1056 µg/ kg, country: Egypt incidence: 2/6, conc. range: 3.6-5.4 µg/ kg, country: Egypt incidence: 9/104*, conc. range: $\leq 1-954$ µg/kg, country: Finland, *imported

incidence: 7/8, conc. range: 22-2222 µg/ kg, Ø conc.: 682 μ g/kg, country: Finland incidence: 3/6, conc. range: 15-240 µg/ kg, country: Gambia incidence: 42/1038, Ø conc.: 97 μg/kg, country: Germany incidence: 46/62*, conc. range: 20-28,000 μg/kg, country: Germany, *moldy incidence: 39/40, conc. range: 1.3-1600 μg/kg, country: Germany incidence: 9/19, conc. range: 5-15 µg/kg, country: India incidence: 926*/2062, conc. range: \leq 833 μ g / kg, contry: India, *exceeded 5 μ g / kg incidence: 1/2, conc.: 5 µg/kg, country: Nigeria incidence: 7/40*, conc. range: tr-400 µg/kg, country: Norway, *imported incidence: 48/1962, conc. range: 5-200 µg/kg, country: South Africa incidence: 10/553, conc. range: 5-20 µg/kg, country: South Africa incidence: 1/259, conc.: 20 µg/kg, country: South Africa incidence: 97/157, conc. range: 5-3000 μg/kg, country: Sudan incidence: 1/20, conc.: 40 μ g/kg, country: Sudan incidence: 2/14, conc. range: 5-20 µg/kg, country: Sudan incidence: 106*/216, Ø conc.: 1530 μ g / kg, country: Thailand, *total: Ø conc.: 12,256 µg / kg AFB₁, AFB₂, AFG₁, AFG₂ incidence: 4/65, conc. range: 6-46 µg/kg, country: Tunisia incidence: 59/605, conc. range: 5-625 μg/kg, country: USA incidence: 2/56, conc. range: 10-125 μg/kg, country: USA incidence: 10/63, conc. range: $\leq 5 - > 5$ μg/kg, country: USA \rightarrow aflatoxin B₂ incidence: 1/1, conc.: 180 μ g/kg, country: Angola

Peanuts

incidence: 2/6, conc. range: 1.8-2.6 μg/kg, country: Egypt incidence: 9/104*, conc. range: \leq 1-568 μg/kg, country: Finland, *imported incidence: 3/8, conc. range: 167-1111 μ g / kg, Ø conc.: 482 μ g / kg, country: Finland incidence: 39/40, conc. range: 1.5-744 μg/kg, country: Germany incidence: 7/40*, conc. range: 2-50 μg/kg, country: Norway, *imported \rightarrow aflatoxin G₁ incidence: 1/1, conc.: 315 µg/kg, country: Angola incidence: 2/109*, conc. range: 3-136 μg/kg, country: Finland, *imported incidence: 4/8, conc. range: 333-556 μ g/kg, Ø conc.: 500 μ g/kg, country: Finland incidence: 39/40, conc. range: 1-1540 µg/kg, country: Germany incidence: 7/40*, conc. range: tr-350 μg/kg, country: Norway, *imported incidence: 4/65, conc. range: $\leq 0.38 \ \mu g/$ kg, country: Tunisia \rightarrow aflatoxin G₂ incidence: 1/1, conc.: 40 µg/kg, country: Angola incidence: 1/109*, conc.: 34 µg/kg, country: Finland, *imported incidence: 2/8, conc.: 167 µg/kg, \emptyset conc.: 167 µg/kg, country: Finland incidence: 39/40, conc. range: 1-548 μg/kg, country: Germany incidence: 7/40*, conc. range: tr-30 μg/kg, country: Norway, *imported \rightarrow aflatoxins (no specification) incidence: 2/2, conc. range: 31-50 µg/kg, country: Brazil incidence: 284*/1679, conc. range: > 5-24.9 μ g/kg (186 samples), > 25 μ g/kg (98 sa), country: Canada, *AFB₁, AFB₂, AFG_1, AFG_2 incidence: 1/2, conc. range: 51-100 μg/kg, country: Egypt incidence: 5/5, conc. range: 1-440 µg/kg, country: Gambia

incidence: 42/1038, Ø conc.: 141 μg/kg, country: Germany incidence: 505/8081*, conc. range: nc, country: Germany, *peanuts and peanut products incidence: 17/35, conc. range: 1-410 μ g / kg, country: India incidence: 93/160, conc. range: tr-5850 μ g / kg, country: India incidence: 20/20*, conc. range: 126-1603 μg/kg, country: Indonesia, *from local farmers incidence: 80/80*, conc. range: 81-14,565 μ g / kg, country: Indonesia, *from the market incidence: 26/53, conc. range: 1-300 μg/kg, country: Malawi incidence: 5/67, conc. range: nc, country: Mocambique incidence: 5/71, conc.: > 30- \leq 100 µg/ kg, country: Philippines incidence: 27*/152, conc. range: 1-100 μ g/kg (11 samples), 100-1000 μ g/kg (8 sa), > 1000 μ g / kg (8 sa), country: Uganda, *24 samples contained AFB₁, 16 AFB₂, 17 AFG₁, 7 AFG₂ incidence: 13/56, conc. range: 1-200 μg/kg, country: USA incidence: 50/50, conc. range: 3-22,000 μ g/kg, Ø conc.: 1685 μ g/kg, country: USA \rightarrow citrinin incidence: 16/160, conc. range: tr-1200 μg/kg, country: India \rightarrow cyclopiazonic acid incidence: 1/6, conc. range: traces, country: USA incidence: 45/50, conc. range: < 50-2900 μ g / kg, Ø conc.: 460 μ g / kg, country: USA incidence: 21/27* conc. range: 32-6525 μ g / kg, country: USA, *loose-shell kernel fractions incidence: 4/21* conc. range: 32-130 μg/kg, country: USA, *sound mature kernel fractions

→ ochratoxin A incidence: 1/1*, conc.: 4900 μ g/kg, country: Canada, *visible moldy → nuts

Peanuts (boiled) may contain the following \rightarrow mycotoxins: aflatoxin (no specification) (\rightarrow aflatoxins) incidence: 8/8, conc. range: $\leq 103 \ \mu g / kg$, \emptyset conc.: 24 $\mu g / kg$, country: Philippines

Peanuts (chocolate-coated) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 3/17, conc. range: $0.5 \le 3$ μ g/kg, country: UK \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 3/17, conc. range: $0.5-5 \mu$ g/kg, country: UK

Peanuts (dry roasted) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 3/14, conc. range: 0.5 - \leq 5 μ g / kg, country: UK \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 3/14, conc. range: 0.5 - \leq 5 μ g / kg, country: UK

Peanuts (fresh, raw) may contain the following \rightarrow mycotoxins: aflatoxin (no specification) (\rightarrow aflatoxins) incidence: 110/169, conc. range: $\leq 885 \ \mu g / kg$, \emptyset conc.: 58 $\mu g / kg$, country: Philippines

Peanuts (in-shell) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins (no specification) incidence: 15/26, conc. range: 0.5-10 μ g/kg (10 samples), 11-50 μ g/kg (2 sa), > 50 μ g/kg (3 sa), country: UK

Peanuts (in-shell, raw) may contain the following \rightarrow mycotoxins:

→ aflatoxin B₁ incidence: 12/12, conc. range: 05-5 µg/kg (9 samples), 11-30 µg/kg (2 sa), 2520 µg/kg (1 sa), country: UK → aflatoxins incidence: 13*/24, conc. range: 0.5-5 µg/kg (8 samples), 6-30 µg/kg (2 sa), 31-100 µg/kg (2 sa), 4920 µg/kg (1 sa), country: UK, *AFB₁, AFB₂, AFG₁, AFG₂ incidence: 1/4, conc.: 273 µg/kg, country: USA

Peanuts (in-shell, roasted) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 5/13, conc. range: 0.5-5 µg/kg (4 samples), 9 µg/kg (1 sa), country: UK \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 5/10, conc. range: 0.5-10 µg/kg (4 samples), 11-50 µg/kg (1 sa), country: UK incidence: 7/13, conc. range: 0.5-5 µg/kg (3 samples), 6-10 µg/kg (2 sa), 11-28 µg/kg (2 sa), country: UK

Peanuts (processed) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 7/150, conc. range: 5-14.9 μ g/kg (5 samples), > 25 μ g/kg (2 sa), country: Canada

Peanuts (roasted) Since the roasting process destroys \rightarrow aflatoxins in contaminated peanuts to a varying degree some manufactures might use low-quality raw materials (see also sliced and crushed \rightarrow peanuts). The use of aflatoxin contaminated peanut oil in roasting further contributes to the contamination of the \rightarrow nuts (absorption). In Finland 6.6% and in Sweden 37% of imported roasted peanuts have been found to contain aflatoxins.

Roasted peanuts may contain the following \rightarrow mycotoxins:

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 \rightarrow aflatoxin B₁

incidence: 3/17, conc. range: $0.5-5 \ \mu g/kg$ (2 samples), $6 \ \mu g/kg$ (1 sa), country: UK \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 4/17, conc. range: $0.5-5 \ \mu g/kg$ (3 samples), $7 \ \mu g/kg$ (1 sa), country: UK

Peanuts (shelled) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 3/8, conc. range: 0.5-10 µg/kg, country: UK

Peanuts (shelled, raw) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/2, conc.: 2.7 µg/kg, country: Syria incidence: 5/8, conc. range: 0.5-5 µg/kg (4 sa), 88 µg/kg (1 sa), country: UK \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 5/8, conc. range: 0.5-5 µg/kg (4 sa), 182 µg/kg (1 sa), country: UK

Peanuts (shelled, roasted) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 8/121*, conc. range: 3-716 μ g/kg, Ø conc.: 160 μ g/kg, country: Finland, *imported incidence: 1/26, conc.: 0.1 µg/kg, country: Japan incidence: 2/3, conc. range: 0.4-1.9 µg/ kg, country: Syria incidence: 3/14, conc. range: 0.5-10 μg/kg, country: UK \rightarrow aflatoxin B₂ incidence: 8/121*, conc. range: 1-89 μ g / kg, Ø conc.: 21.6 μ g / kg, country: Finland, *imported incidence: 2/3, conc. range: 0.3-0.6 µg/ kg, country: Syria \rightarrow aflatoxin G₁ incidence: 2/108*, conc. range: 12-20 μ g/kg, Ø conc.: 16 μ g/kg, country: Finland, *imported

→ aflatoxins (no specification) incidence: 6/55, conc. range: \leq 329 µg/kg, Ø conc.: 68 µg/kg, country: USA incidence: 1/1, conc.: 4 µg/kg, country: USA

Peanuts (shelled, roasted, salted) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins incidence: 4*/17, conc. range: 0.5-10 μ g/kg, country: UK, *AFB₁, AFB₂, AFG₁, AFG₂ incidence: 5/3, conc. range: nc, country: UK

Peanuts (shelled, roasted, unsalted) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins (no specification) incidence: 5/12, conc. range: nc, country: UK

Peanuts (sliced) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 144/718, conc. range: 5-665 μ g/kg, Ø conc. 134 μ g/kg, country: Germany

Peanuts (sliced and crushed): In a Finnish study sliced and crushed peanut samples were frequently contaminated with \rightarrow aflatoxins. It was suggested that a more even distribution of aflatoxin in these lots and / or the use of low-quality material in the preparation of the corresponding lots are responsible. Sliced and crushed peanuts may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 20/68*, conc. range: \leq 1-716 μg/kg, country: Finland, *imported \rightarrow aflatoxin B₂ incidence: 18/68*, conc. range: \leq 1-76 μg/kg, country: Finland, *imported \rightarrow aflatoxin G₁ incidence: 6/68*, conc. range: \leq 1-91 μg/kg, country: Finland, *imported

 \rightarrow aflatoxin G₂ incidence: $4/68^*$, conc. range: $\leq 1-14$ μg/kg, country: Finland, *imported **Pear juice** may contain the following \rightarrow mycotoxins: \rightarrow patulin incidence: 1/4, conc.: 24 µg/kg, country: Germany **Pears** may contain the following \rightarrow mycotoxins: \rightarrow patulin incidence: 8/24, conc. range: 0.9-10 μg/kg, country: Spain \rightarrow apples **Peas** may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/35, conc.: 25 µg/kg, country: Tunisia \rightarrow aflatoxin G₁ incidence: 1/35, conc.: 42 µg/kg, country: Tunisia \rightarrow aflatoxins incidence: 3*/19, conc. range: 1-100 μg/kg, country: Uganda * 2 samples contained AFB₁, 1 AFB₂, 1 AFG₁ aflatoxin (no specification) incidence: 3/8, conc. range: \leq 40 µg/kg, \emptyset conc.: 13 µg/kg, country: Philippines \rightarrow ochratoxin A incidence: 2/72, conc.: 10 µg/kg, country: Sweden incidence: 6/71, conc. range: 10-442 μg/kg, country: Sweden \rightarrow beans, \rightarrow cabbage, \rightarrow cowpeas \rightarrow lentils, \rightarrow pigeon peas, \rightarrow soybeans, \rightarrow vegetables Pecans

Pecans Since \rightarrow aflatoxins have been detected in damaged as well as in nonvisibly damaged kernels the major cause of contamination is not clear. Nevertheless, weevil-damaged and late-harvested \rightarrow nuts (shell integrity) may be more

susceptible to mold invasion. The prevailing orchard temperatures during the latter part of the harvest season greatly influence the degree of contamination. In addition, nuts falling to the ground in pastures, especially on wet soil, are more likely to mold than those falling in nonpasture orchards. Besides other mycotoxins \rightarrow alternariol and \rightarrow alternariol methyl ether have been detected in pecans. These \rightarrow mycotoxins only occurred in discolored kernels which were removed from shelled pecans during processing. They would probably be rejected by consumers of in-shell pecans. In addition, per capita consumption of pecans is very low which further reduces the risk of intake of \rightarrow Alternaria mycotoxins. Pecans may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 3/48, conc. range: tr-25 µg/kg, country: USA \rightarrow aflatoxin G₂ incidence: 3/48, conc. range: traces, country: USA \rightarrow aflatoxins incidence: 1*/55, conc. range: 5-9.9 μ g / kg, country: Canada, *AFB₁, AFB₂, AFG₁, AFG₂ incidence: 39/575, conc. range: ≤ 172 μ g / kg, Ø conc.: 86 μ g / kg, country: USA incidence: 1/229, conc.: 40 µg/kg, country: USA incidence: 3/17, conc. range: ≤ 334 μ g/kg, Ø conc.: 135 μ g/kg, country: USA \rightarrow alternariol* incidence: nc/50, conc. range: nc, country: USA → alternariol methyl ether* incidence: nc/50, conc. range: nc, country: USA *in discolored pecans ("pickouts") → citreoviridin incidence: 1/1*, conc.: nc, country: USA, *moldy fragments

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Pecans

→ sterigmatocystin incidence: 1/20, conc.: 20,000 µg/kg, country: USA → nuts

Pellagra This human disease is characterized by the insufficient intake / failure to absorb the B complex vitamin niacin or its amide. People consuming deteriorated \rightarrow maize as a staple food are most frequently affected. Maize contains only low levels of niacin in an available form and the concentration of certain niacin precursors is also low. These compounds are essential for the activity of certain enzymes which are involved in detoxification processes of the \rightarrow mycotoxins ingested via contaminated maize. It is suggested, that the effects of this malnutrition are enhanced by certain mycotoxins such as \rightarrow trichothecenes, \rightarrow fumonisins, \rightarrow koji acid and \rightarrow zearalenone. Pellagra is more common in spring time and it is concluded that storing maize under cool humid conditions in winter promotes trichothecene (especially \rightarrow T-2 toxin) production and contamination. However, up to now the real cause of Pellagra remains unresolved.

Penicidin (Syn.: \rightarrow patulin)

Penicillic acid This 3-methoxy-5-methyl-4-oxo-2,5-hexadienoic acid or 2-keto- β methoxy- δ -methylene- $\Delta\alpha$ -hexenoic acid (\rightarrow mycotoxins) was one of the first metabolites isolated (\rightarrow Penicillium *puberulum*, 1913). It was recognized as a toxic fungal metabolite possibly as early as 1896 (see Figure Penicillic acid).

CHEMICAL DATA

Empirical formula $C_8H_{10}O_4$, molecular weight: 170

FUNGAL SOURCES important producers: \rightarrow Penicillium aurantiogriseum Dierckx and varieties, \rightarrow Penicillium roquefortii Thom chemotype II (only a few isolates), *P. janczewskii*, \rightarrow Eupenicillium spp., \rightarrow Petromyces alliaceus Malloch & Cain, \rightarrow Aspergillus alutaceus var. alutaceus Berkely & Curtis, \rightarrow Aspergillus quercinus (Bain.) Thom & Church, *A. sclerotiorum*.

NATURAL OCCURRENCE

→ apples, → barley, → beans, → cereals, → cheese, → cheese, Blue, → cheese, goat, → cheese, Swiss, → maize, → rye Certain strains of *P. roquefortii* which have been used in the cheese industry produced penicillic acid. Penicillic acid is not stable in foods containing reactive amino acids.

Тохісіту

nephrotoxic, \rightarrow mutangenic, carcinogenic,

 LD_{50} (po) : 35-600 mg/kg bw mice A potentiated effect in the nephrotoxic action of penicillic acid and \rightarrow ochratoxin A was observed. Furthermore, a synergistic effect between \rightarrow patulin and penicillic acid is evident.

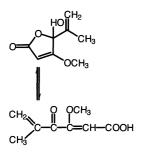
Although the adducts of penicillic acid with cysteine or glutathione should be biologically inactive they retained some toxicity to the chick embryo.

DETECTION GC, TLC

FURTHER COMMENTS

Stability: The inactivation of penicillic acid by SH-compounds is due to a reaction with the isolated rather than the conjugated double bond. In aqueous solution the reaction product formed with cysteine derived from the openchain form of penicillic acid or from the lactone form.

Compared to the aflatoxins, the penicillic acid like \rightarrow patulin seems to be less stable in certain foods. Thiol compounds should be responsible for the instability in orange juice and \rightarrow flour but stability was observed in \rightarrow grape juice and \rightarrow apple juice. No penicillic acid could be



Penicillic acid

detected in bologna 48 h, storage temperature 5 °C. Only low levels (< 10%) of this mycotoxin persisted in Swiss cheese after one week at 5 °C. A rapid loss of penicillic acid has been reported during the grinding of maize. The formation of penicillic acid during the aging of meats (\rightarrow meat) will result in non-toxic products since this mycotoxin reacts with amino acids.

Penicillic acid is not stable in stored \rightarrow wheat flour. After 2 weeks only 10% remained at 22 °C. Within 3 days a complete decomposition was observed after treament with 2% aqueous ammonia.

Penicillium anamorphic \rightarrow Trichocomaceae, teleomorphs \rightarrow Eupenicillium, *Talaromyces*

This ubiquitous distributed genus is more common in temperate climatic regions of the world such as Canada and northern Europe. Although generally accepted as storage fungi some species occur on cereal \rightarrow grains before harvest. *Penicillium* spp. are usually the dominant organisms of the blue and green molds associated with the spoilage of \rightarrow foods, especially \rightarrow fruits (citrusfruits) and \rightarrow vegetables. Cool storage (-2 to 5 °C) of damp grain (\rightarrow a_w > 0.90) causes the "blue eye" disease.

85 *Penicillium* species are known to be toxigenic but most \rightarrow mycotoxins in this genus are produced by a small, well defined range of species. Within each

species a wide range of mycotoxins with an extreme diverse molecular composition is synthesized. At least 27 Penicillium metabolites are known to be toxic to man and animals which are produced by 32 species. Although the toxicity of these mycotoxins is also very diverse, most toxins either affect liver and kidney function or they are neurotoxins. A water activity of a_w 0.80-0.82 is sufficient for the growth of P. aurantiogriseum and P. verrucosum whereas aw levels between 0.86-0.89 are necessary for mycotoxin production. Important mycotoxin producers are e.g. \rightarrow Penicillium aurantiogriseum Dierckx, \rightarrow Penicillium citroenigrum Dierckx, → Penicillium expansum Link, \rightarrow Penicillium islandicum Sopp, \rightarrow Penicillium roquefortii Thom, \rightarrow Penicillium verrucosum Dierckx. Important mycotoxins are e.g. \rightarrow citrinin, \rightarrow ochratoxin A, \rightarrow penicillic acid, and \rightarrow xanthomegnin. In general Penicillium spp. are capable of producing mycotoxins at lower temperatures than are \rightarrow Aspergillus spp.

Penicillium aurantiogriseum Dierckx (Syn.: *P. cyclopium*) is of ubiquitous distribution and found on different kinds of food such as \rightarrow bread, \rightarrow cereals, \rightarrow cheese, \rightarrow coffee beans, \rightarrow grains, frozen \rightarrow meat, \rightarrow nuts, \rightarrow sausages, \rightarrow shrimps. *P. aurantiogriseum* is the most important member of all Penicillia in stored \rightarrow cereals. There is a broad temperature range for \rightarrow ochratoxin A? and \rightarrow penicillic acid production (4-31 °C). The minimum a_w for penicillic acid production is a_w 0.97-0.99.

P. aurantiogriseum may produce \rightarrow mycotoxins such as penicillic acid, penitrem A (\rightarrow penitrems), terrestric acids, verrucosidin, \rightarrow viomellein, viridicatins, xanthomegnins (\rightarrow xanthomegnin).

Penicillium camembertii Thom (Syn.: e.g. *P. candidum, P. caseicola*) is a white grow-

ing mold used for the manufacturing of Camembert cheese (\rightarrow cheese, Camembert). Surface growth of this mold prevents (i) undesirable fungal infections and causes (ii) proteolytic degradation of casein and (iii) hydrolysis of triglycerides. However, it seems that *P. camembertii* is a consistent producer of \rightarrow cyclopiazonic acid whereas the minimum temperature for production is 4 °C.

Penicillium chrysogenum Thom (Syn.: *P.* notatum) is a penicillin producer and common on different types of food such as \rightarrow almonds, \rightarrow bread, \rightarrow cheese, \rightarrow fish, \rightarrow flour, \rightarrow ham, \rightarrow meat, \rightarrow nuts, \rightarrow sausages. In some countries (e.g. Canada) it is frequently isolated from \rightarrow cereals. \rightarrow Roquefortine C might occur naturally in cereals infected with *P.* chrysogenum. \rightarrow Ochratoxin A production of this fungus could not be confirmed (see Figure Penicillium chrysogenum Thom).

P. chrysogenum may produce \rightarrow mycotoxins such as \rightarrow PR toxin, roquefortine C and D, xanthocillins.

Penicillium citreonigrum Dierckx (Syn.: *Penicillium citreo-viride*) although widely distributed is not a commonly isolated species. \rightarrow Rice seems to be the best substrate whereas growth starts soon after the \rightarrow grains become wet under improper storage conditions. The lower tempera-

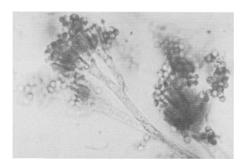
tures and shorter hours of daylight in the more temperate rice-growing areas favor its growth. At a moisture content of 14.6% *P. citreonigrum* starts growing in stored rice. It is overgrown by other fungi if the moisture content reaches 15.6%. This fungus is involved in the \rightarrow Yellow rice disease / \rightarrow acute cardiac beriberi.

P. citreonigrum may produce \rightarrow mycotoxins such as \rightarrow citreoviridin.

Penicillium citrinum Thom as an ubiquitous fungus is a contaminant of nearly every kind of foodstuff but is found predominately on subtropical and tropical \rightarrow cereals. Besides cereal \rightarrow grains and \rightarrow flour (the most common sources) this mold has been isolated from e.g. \rightarrow almonds, \rightarrow bread, \rightarrow cheese, \rightarrow coffee beans, \rightarrow fish, \rightarrow fruit juices, \rightarrow meat, \rightarrow nuts, \rightarrow spices (see Figure *Penicillium citrinum* Thom). *P. citrinum* is a consistent producer of \rightarrow citrinin although in the presence of \rightarrow Aspergillus niger and / or *Trichoderma viride* toxin production is inhibited.

P. citrinum may produce \rightarrow mycotoxins such as citrinins.

Penicillium commune Thom may produce the following \rightarrow mycotoxins: cyclopaldic acid, \rightarrow cyclopiazonic acid, \rightarrow roquefortine A & B (*P. commune* chemotype II), rugulovasines.



Penicillium chrysogenum Thom



Penicillium citrinum Thom

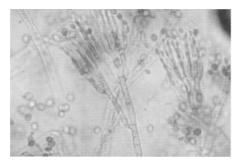
Penicillium crustosum Thom is an ubiquitous spoilage fungus, preferring products with a high content of lipids and proteins. \rightarrow Cereals are less often affected. Occurrence has also been reported for \rightarrow almonds, \rightarrow cheese, \rightarrow flour, \rightarrow fruit juices, \rightarrow maize (high lipid content), \rightarrow meat (processed), and \rightarrow nuts. Furthermore, it is a weak pathogen on pomaceous \rightarrow fruits and cucurbits. Nearly all isolates produce the tremorgenic penitrem A and therefore *P. crustosum* is by far the most important source of this mycotoxin.

P. crustosum may produce \rightarrow mycotoxins such as penitrem A (\rightarrow penitrems), \rightarrow roquefortine A, B, C, terrestric acid, viridicatin, \rightarrow xanthomegnin.

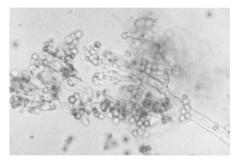
Penicillium expansum Link is a common storage mold in \rightarrow apples and \rightarrow pears. Strains of *P. expansum* tolerate low oxygen levels as well as high CO₂ tensions (see Figure *Penicillium expansum* Link). Since *P. expansum* is the most important \rightarrow patulin producer, infection is usually associated with patulin contamination of the \rightarrow fruits (Golden Delicious: 2-100 µg/g). Conventional CO₂ and O₂ tensions in CA storage inhibit the growth of this fungus. A minimum \rightarrow a_w of 0.99 is needed for patulin production (temperature 0-24 °C). *P. expansum* may produce \rightarrow mycotoxins such as chaetoglobosins, \rightarrow citrinin, \rightarrow patulin, \rightarrow roquefortine C.

Penicillium griseofulvum Dierckx as a ubiquitous species could be isolated from different kinds of \rightarrow foods such as \rightarrow cereals and \rightarrow meat. The minimum $\rightarrow a_w$ that allows \rightarrow patulin production is $\approx a_w$ 0.94 (temperature 30 °C). Temperatures that enabled patulin production were in the range of 4-31 °C (see Figure *Penicillium griseofulvum* Dierckx). *P. griseofulvum* may produce \rightarrow mycotoxins such as \rightarrow cyclopiazonic acid, griseofulvins, \rightarrow patulin, \rightarrow roquefortine C. Possible Mycotoxicosis Patulin (malt) and cyclopiazonic acid (cereals) are involved in mycotoxicoxsis.

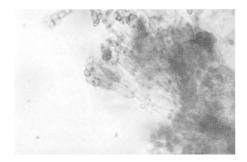
Penicillium islandicum Sopp is widely distributed but occurs infrequently in nature. Classified as a storage fungus or a member of the mycoflora of soils, this species is uncommon at least in the temperate zones. As a more or less frequent contaminant of \rightarrow rice, *P. islandicum* is involved in the \rightarrow yellow rice disease (see Figure *Penicillium islandicum* Sopp). It represents an important problem for rice consuming peoples in most Asiatic and African countries with high temperatures and a humid climate. In rice mycotoxin production is favored by moisture con-



Penicillium expansum Link



Penicillium griseofulvum Dierckx



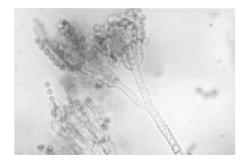
Penicillium islandicum Sopp

tents > 16% in combination with high temperatures (\approx 33 °C optimum). After the first isolation by Sopp (1912) on the Island Skyr (Norway) it became obvious that *P. islandicum* produces a series of very hepatotoxic substances. They cause acute liver atrophy, liver \rightarrow cirrhosis and liver tumors. *P. islandicum* may produce \rightarrow mycotoxins such as emodin, erythroskyrin, islandic acid, \rightarrow islanditoxin, \rightarrow luteoskyrin, \rightarrow rugulosin, skyrin.

Penicillium roquefortii Thom (Syn.: P. casei, P. biourgei, P. gorgonzolae, P. stilton, P. vesiculosum) is frequently found in \rightarrow cereals stored under controlled \rightarrow atmosphere and silage. Starter cultures of this fungus are used in the cheese industry for the preparation of blue veined cheese. P. roquefortii prevents (i) undesirable fungal infections and causes (ii) proteolytic degradation of casein as well as (iii) hydrolysis of triglycerides. However, this fungus produces a variety of toxic metabolites. Of these, \rightarrow mycophenolic acid, \rightarrow penicillic acid, \rightarrow roquefor tine C and \rightarrow roque for tine A & B have been detected in naturally contaminated \rightarrow cheeses. The natural contamination of blue veined cheese with these mycotoxins as well as their toxicological properties do not represent a risk for human health. *P. roquefortii* may produce \rightarrow mycotoxins such as \rightarrow cyclopiazonic acid, mycophenolic acid, \rightarrow patulin, penicillic acid, PRtoxins (PR-toxin *P. roquefortii* chemotype I only), roquefortine A, B, C (*P. roquefortii* Chemotype I and II) & D.

Penicillium verrucosum Dierckx is very common in temperate regions of the world (especially northern parts of Europe) where it almost exclusively occurs (see Figure Penicillium verrucosum Dierckx). Cereal \rightarrow grains (\rightarrow maize, \rightarrow wheat, \rightarrow barley) are most frequently affected resulting in a possible OTA contamination. Infestation of some kernels from anthesis and surface contamination is common at harvest. The absolute amount of pre-harvest infection is influenced by site and season. During combine harvesting, conidia of P. verrucosum are disseminated resulting in the contamination of other grains. In addition, refrigerated \rightarrow meat and \rightarrow cheese products in subtropical areas as well as \rightarrow fish may also contaminated.

P. verrucosum is the only known and confirmed producer of \rightarrow ochratoxin A within the genus \rightarrow Penicillium. Formation of this mycotoxin is enhanced by the amino acids proline and glutamic acid. A positive correlation between the protein concentration of \rightarrow barley and the production of OTA was established. P. verrusocum is the causal microorganism of \rightarrow Mycotoxic porcine nephropathy in pigs in Denmark, Sweden and Hungary. It is suggested that this disorder due to ochratoxin A may be enhanced by \rightarrow citrinin and oxalic acid. Particularly at lower temperatures P. verrucosum causes citrinin-contamination of cereals whereas no citrinin is produced on oilseeds crops. Similarly, wheat gave better OTA yields than corn or the oilseed crops. *P. verrucosum* may produce \rightarrow mycotoxins such as \rightarrow citrinin (*P. verrucosum* Chemotype II), \rightarrow ochratoxin A (*P. verruco*sum Chemotype I and II).

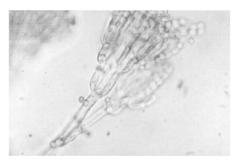


Penicillium verrucosum Dierckx

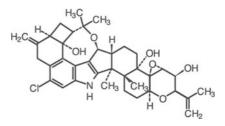
Penicillium viridicatum Westling should be involved in the \rightarrow Mycotoxic porcine nephropathy of Danish pigs but it could be shown, after correct identification, that \rightarrow Penicillium verrucosum Dierckx was the causal organism. *P. viridicatum* has been isolated from e.g. \rightarrow almonds, \rightarrow cereals, \rightarrow cereal products, \rightarrow fish, \rightarrow meat products, \rightarrow nuts, \rightarrow shrimps (see Figure *Penicillium viridicatum* Westling).

P. viridicatum may produce \rightarrow mycotoxins such as \rightarrow penicillic acid, \rightarrow viomellein, viridicatins, \rightarrow xanthomegnin.

Penitrems are indole derivatives $(\rightarrow \text{ mycotoxins})$ which contain only one nitrogen per molecule (see Figure Penitrems). Penitrem A, B, C are produced by \rightarrow Penicillium spp. even at low (refrigeration) temperature.



Penicillium viridicatum Westling



Penitrems. Penitrem A

Chemical Data

Empirical formula: C₃₇H₄₄O₆NCl, molecular weight: 633 (penitrem A)

FUNGAL SOURCES

 \rightarrow Penicillium crustosum Thom, *P. clavigenum*, *P. glandicola*

NATURAL OCCURRENCE \rightarrow cheese, cream, \rightarrow walnuts

Тохісіту

neurological (tremors, \rightarrow convulsions) and \rightarrow renal effects

 LD_{50} (ip): 1.05 mg/kg bw mice (penitrem A)

The mammalian toxicity of penitrem C is unknown.

In humans dizziness and vomiting may be caused by the intake of penitrems but patients recovered completely in all cases.

DETECTION HPLC, MS, spectroscopy, TLC

FURTHER COMMENTS

In nature compounds causing sustained trembling are rare, whereas most of them are synthesized by molds. The intoxication of dogs consuming moldy cream cheese was the first definitive natural occurrence of penitrem A toxicosis.

Pepper may contain the following

- \rightarrow mycotoxins:
- \rightarrow alternariol

incidence: 1/1*, conc.: 640 μg/kg, country: Italy

 \rightarrow alternariol methyl ether

incidence: 1/1*, conc.: 49 µg/kg, country: Italy

Pepper

→ ochratoxin A incidence: 11/11, conc. range: \leq 4.9-8 µg/kg, country: Austria incidence: 1/4, conc.: 40 µg/kg, country: Germany → tenuazonic acid incidence: 1/1*, conc.: 54 µg/kg, country: Italy *sample was visibly affected by → Alternaria rot → spices

Pepper (black) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 4/15, Ø conc.: 35 µg/kg, country: Egypt incidence: 5/8, conc. range: 17-190 μg/kg, country: India \rightarrow aflatoxin B₂ incidence: 5/8, conc. range: 12-150 μg/kg, country: India \rightarrow aflatoxin G₁ incidence: 2/20*, conc. range: 1.72-3.18 μ g / kg, Ø conc.: 2.45 μ g / kg, country: Egypt, *different \rightarrow spices incidence: 3/7*, conc. range: 1.8-3.7 μg/kg, country: Canada, *imported incidence: nc/137*, conc.: 1.1 µg/kg, country: Canada, *imported incidence: 5/8, conc. range: 15-75 µg/kg, country: India \rightarrow aflatoxin G₂ incidence: 5/8, conc. range: 12-76 µg/kg, country: India \rightarrow citrinin incidence: 1/8, conc.: 50 µg/kg, country: India \rightarrow sterigmatocystin incidence: 2/8, conc. range: 105-125 μ g/kg, Ø conc.: 115 μ g/kg, country: India \rightarrow zearalenone incidence: 1/8, conc.: nc, country: India

Pepper (red): It was suggested that aflatoxin production in red peppers starts

during eight to ten days they were spread out to dry after harvesting. Red pepper may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 11/22, conc. range: tr-24 μg/kg, country: Germany incidence: 6/6, conc. range: tr-6 µg/kg, country: India incidence: 4/9, conc. range: 15-146 μg/kg, country: India incidence: 1/2*, conc.: 0.8 µg/kg, country: Japan, *imported incidence: nc, conc. range: \leq 700 µg/kg, country: Nigeria incidence: 12*/106**, Ø conc.: 125 μg/kg, country: Thailand *total: \emptyset conc.: 966 µg/kg AFB₁, AFB₂, AFG₁, AFG₂, **chili peppers incidence: 7/15, conc. range: 0.2-32.9 μ g / kg, Ø conc.: 9.21 μ g / kg, country: USA \rightarrow aflatoxin B₂ incidence: 4/9, conc. range: 11-88 µg/kg, country: India incidence: 7/15, conc. range: 0.1-1.5 μ g/kg, Ø conc.: 0.43 μ g/kg, country: USA \rightarrow aflatoxin G₁ incidence: 4/9, conc. range: 8-58 µg/kg, country: India incidence: 4/15, conc. range: 0.7-28.4 μ g/kg, Ø conc.: 9.07 μ g/kg, country: USA \rightarrow aflatoxin G₂ incidence: 4/9, conc. range: 6-40 µg/kg, country: India incidence: 1/15, conc.: 1.1 µg/kg, country: USA \rightarrow aflatoxins incidence: 18/50*, conc. range: 1-3.9 $\mu g^{**} / kg$ (7 samples), 4-50 $\mu g^{**} / kg$ (11 sa), country: UK, *imported, **AFB₁, AFB₂, AFG₁, AFG₂ (total) incidence: 9/14*, conc. range: 1-3.9 $\mu g^{**} / kg$ (5 samples), 4- > 50 $\mu g^{**} / kg$ (4 sa), country: UK, *imported, port samples, **AFB₁, AFB₂, AFG₁, AFG₂ (total)

incidence: 9/12*, conc. range: \leq 30 µg/kg, Ø conc.: 10 µg/kg, country: USA, *imported \rightarrow ochratoxin A incidence: 13/18, conc. range: \leq 4.9-38 µg/kg, country: Austria incidence: 4/4, conc. range: \leq 4.9-50.4 µg/kg, country: UK \rightarrow zearalenone incidence: 1/9, conc.: nc, country: India \rightarrow spices

Pepper (white) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/7, conc.: 0.3 µg/kg, country: USA incidence: 4/15, \emptyset conc. range: $\leq 22 \ \mu g/$ kg, country: Egypt incidence: 1/13*, conc.: 0.6 µg/kg, country: Japan, *imported incidence: 7/24, conc. range: 0.6-2.3 μg/kg, country: Japan incidence: 1/7, conc.: 0.3 µg/kg, country: USA \rightarrow aflatoxin B₂ incidence: 7/24, conc. range: 0.1-0.2 μg/kg, country: Japan \rightarrow aflatoxin G₁ incidence: 7/24, conc. range: 0.2-1.4 μg/kg, country: Japan \rightarrow spices

Pepper cheese \rightarrow cheese, pepper \rightarrow

Persipan (apricot seed paste) Blanched peach and apricot seeds for persipan manufacture should be processed immediately after blanching because aflatoxin contamination may occur very rapidly. Persipan may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 6/16, conc. range: tr-5 µg/kg, country: Germany → aflatoxin B_2 incidence: 3/16, conc.: traces, country: Germany → aflatoxin G_1 incidence: 2/16, conc.: tr-3 µg/kg, country: Germany → marzipan

Petromyces \rightarrow Trichocomaceae, anamorph \rightarrow Aspergillus (*ochraceus* group) *P. alliaceus* belongs to the genus *Aspergillus*, subgenus Circumdati, section Circumdati. *P. alliaceus* is a known \rightarrow ochratoxin A producer.

Pheasants may contain the following → mycotoxins: → aflatoxin B₁ incidence: 56/94*, conc. range: 0.3-0.985 µg / kg, Ø conc.: 0.329 µg / kg, country: Czechoslovakia, *liver incidence: 79/94*, conc. range: 0.3-1.67 µg / kg, Ø conc.: 0.679 µg / kg, country: Czechoslovakia, *kidney → meat

Phoma anamorphic Pleosporaceae, teleomorph *Pleospora*

Pig blood may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A incidence: 910/1200, conc. range: 5-20 μ g/l (861 samples), 20-100 μ g/l (44 sa), 100-229 μ g/l (5 sa), country: Canada incidence: 574/1169, \emptyset conc.: ca. 1 μ g/l, country: Germany incidence: 178/216, conc. range: > 5 μ g/l, country: Norway incidence: 36/195, conc. range: 3-270 μ g / l, country: Poland incidence: 47/279, conc. range: 2-187 μ g/l, Ø conc.: 15.7 μ g/l, country: Sweden incidence: 6/76, Ø conc. range: 36-37 μg/l, country: Yugoslavia \rightarrow meat, \rightarrow pork

Pig kidneys (normal) may contain the following \rightarrow mycotoxins: \rightarrow citrinin incidence: 9/125, conc. range: 0.1- > 10 μ g / kg, country: UK \rightarrow ochratoxin A incidence: 9/95, conc. range: 0.2- > 80 μg/kg, country: Belgium incidence: 1/63, conc.: 1-5 µg/kg, country: Czechoslovakia incidence: 4403/7639, conc. range: > 25 μ g / kg (4293 samples), > 150 μ g / kg (110 sa), country: Denmark incidence: 137/686, conc. range: 2-67 μg/kg, country: Denmark incidence: 5/25, conc. range: > 25 μ g / kg, country: Denmark incidence: 20/20, conc. range: 0.5-1955 μg/kg, country: Denmark incidence: 10/193*, conc. range: < 0.1-5 μ g / kg, country: Finland, *normal and suspected incidence: 47/354, Ø conc.: 1.4 µg/kg, country: Germany incidence: 42/300, conc. range: 0.5-10.2 μg/kg, country: Germany incidence: 15/100, conc. range: 0.5-16.4 μg/kg, country: Germany incidence: 48/122, conc. range: 2-100 μg/kg, country: Hungary incidence: 2/131, conc. range: 7-10 μg/kg, country: Norway incidence: 32/129, conc. range: 2-104 μ g / kg, country: Sweden incidence: 24/90, conc. range: 2-88 μg/kg, country: Sweden incidence: 12/36, conc. range: 0.1-0.2 $\mu g \, / \, kg$ (11 samples), 0,3 $\mu g \, / \, kg$ (1 sa), country: Switzerland incidence: 7/12, conc.: \leq 1.0 μg / kg, country: The Netherlands incidence: 1/6, conc. range: 0.2-0.8 µg/kg, country: The Netherlands incidence: 15/104, conc. range: \leq 4.9-9.3 μ g / kg, Ø conc.: 0.84 μ g / kg, country: UK incidence: 242/378, conc. range: 0.5- > 10 μg/kg, country: UK

incidence: 43/278, conc. range: 1-10 μ g/kg (41 samples), 22-44 μ g/kg (2 sa), country: UK incidence: 4/76*, Ø conc.: 21 μ g/kg, country: Yugoslavia, *partly suspected \rightarrow meat, \rightarrow pork

Pig kidneys (suspected) may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A incidence: 69/104, Ø conc.: 0.75 µg/kg, country: Austria incidence: 28/95, conc. range: 0.2-9.99 μg/kg, country. Belgium incidence: 68/385, conc. range: 0.2-12 μg/kg, country: Belgium incidence: 76/96, conc. range: 1-20 μg/kg, country: Czechoslovakia incidence: 21/60, conc. range: 2-68 μg/kg, country: Denmark incidence: 20/20, conc. range: 0.2-1965 μ g / kg, Ø conc.: 34.2 μ g / kg, country: Denmark incidence: 3/38, conc. range: \leq 4.9 µg/ kg, Ø conc.: 0.7 μ g/kg, country: Finland incidence: 22/104, conc. range: 0.1-1.8 μ g/kg, Ø conc.: 0.45 μ g/kg, country: Germany incidence:48/122, conc. range: 10-7100 μg/kg, country: Hungary incidence: 77/197, conc. range: 5-100 μg/kg, country: Hungary incidence: 27/113, conc. range: tr-23 μg/kg, country: Poland incidence: 33/73, conc. range: 2-23 μ g / kg, country: Poland incidence: 32/129, conc. range: 2- < 5 μ g / kg (25 samples), 5- < 10 μ g / kg (2 sa), $10- \le 104 \ \mu g / kg$ (5 sa), country: Sweden incidence: 35/75, conc. range: $\leq 2.0 \ \mu g/$ kg, country: The Netherlands incidence: 33/46, conc. range: 0.2-2 μ g / kg, country: The Netherlands incidence: 6/6, conc. range: 0.2-1 µg/kg, country: The Netherlands

incidence: 2/29, conc. range: 0.2-0.4 μ g/kg, country: The Netherlands incidence: 17/24*, conc. range: 0.2-240 μ g/kg, country: The Netherlands, *originating from Denmark incidence: 112/303*, conc. range: 0.5- < 5 μ g/kg (104 samples), 5- < 10 μ g/kg (6 sa), 11.5-12.4 μ g/kg (2 sa), country: UK, *unsuitable for human consumption \rightarrow meat, \rightarrow pork

Pig liver may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 5/13, conc. range: < 5 µg/kg, country: Germany \rightarrow ochratoxin A incidence: 4/76*, Ø conc.: 21 µg/kg, country: Yugoslavia, *partly suspected \rightarrow meat, \rightarrow pork

Pig serum Certain \rightarrow sausages, e.g. frankfurter-type, are produced with pig serum (plasma) and may therefore be contaminated with ochratoxin A. Pig serum may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A incidence: 32/1445, Ø conc.: 12.6 µg/l, country: Canada incidence: 72/143, Ø conc.: 21 µg/l, country: Canada incidence: 146/283, Ø conc.: 1.2 μg/l, country: Germany incidence: 93/191, conc. range: 0.1-67.3 μ g/l, Ø conc.: 5.8 μ g/l, country: Germany \rightarrow meat, \rightarrow pork

Pigeon peas may contain the following \rightarrow mycotoxins: aflatoxin (no specification) (\rightarrow aflatoxins) incidence: 5/9, conc. range: $\leq 23 \ \mu\text{g}/\text{kg}$, \emptyset conc.: 7 $\mu\text{g}/\text{kg}$, country: Philippines \rightarrow beans, \rightarrow cabbage, \rightarrow cowpeas, \rightarrow lentils, \rightarrow peas, \rightarrow soybeans, \rightarrow vegetables **Pine nuts** Pudding prepared from contaminated \rightarrow nuts contained more than 83% of the original amount of aflatoxin. Pine nuts may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 26/50, conc. range: 25-2080 µg / kg, country: Tunisia \rightarrow aflatoxin G₁ incidence: 26/50, conc. range: 56-4570 µg / kg, country: Tunisia \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 26/50, conc. range: 95-7550 µg / kg, country: Tunisia

Piper betle (medicinal seeds) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: nc/nc, conc. range: 20-1000 µg/kg, country: India \rightarrow citrinin incidence: nc/nc, conc. range: 10-720 µg/kg, country: India

Pipian paste may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins (no specification) incidence: 3/3*, conc. range: \leq 78 µg/kg, \emptyset conc.: 53 µg/kg, country: USA, *imported

Pistachio candy may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins incidence: 1/1*, conc.: 78 µg/kg, country: USA, *imported

Pistachio nuts As in the case of \rightarrow peanuts an uneven distribution of \rightarrow aflatoxins has been established in pistachio nuts samples. Only a few nuts contained high aflatoxin concentrations (≤ 1.4 g/kg). The highest contamination occurred only in brown, brown spotted or fluorescent pistachio kernels. Using an automatic sorter, which removes \rightarrow nuts with fluorescent shells, the aflatoxin content could

be reduced by ca. 50%. However, nonfluorescent nuts (shells) which may also contain significant levels of aflatoxin, escape this control measure. Aflatoxin contamination of pistachio nuts occurs after soaking to remove the hulls from the shells and / or during improper storage. Aflatoxin producers gain entry to the nut along the vascular system. This tissue connects the kernel with the shell. Pistachios may contain the following \rightarrow mycotoxins: \rightarrow aflatoxicol incidence: 5/54, conc. range: 0.2-13.9 μ g / kg, Ø conc.: 3.62 μ g / kg, country: Thailand incidence: 51/247, Ø conc.: 27 μg/kg, country: Germany \rightarrow aflatoxin B₁ incidence: 6/54, conc. range: 7.9-1830 μ g / kg, Ø conc.: 585 μ g / kg, country: Japan incidence: 51/247, Ø conc.: 21 μg/kg, country: Germany incidence: 67/140, conc. range: $< 5 \mu g / kg$ (40 samples), 11-35 $\mu g\,/\,kg$ (27 sa), country: Germany incidence: 1/19, conc.: 22 µg/kg, country: Tunisia \rightarrow aflatoxin B₂ incidence: 6/54, conc. range: 1.5-235 μ g/kg, Ø conc.: 86 μ g/kg, country: Japan \rightarrow aflatoxin M₁ incidence: 5/54, conc. range: 0.9-51.8 μ g / kg, Ø conc.: 21.7 μ g / kg, country: Japan \rightarrow aflatoxins (no specification) incidence: 19*/175, conc. range: 5-24.9 $\mu g / kg (12 \text{ samples}), > 25 \mu g / kg (7 \text{ sa}),$ country: Canada, *AFB₁, AFB₂, AFG₁, AFG₂ incidence: 61/993, conc. range: nc, country: Germany incidence: 7/22, conc. range: $\leq 252 \ \mu g /$ kg, Ø conc.: 58 μ g/kg, country: USA

incidence: 10/21, conc. range: $\leq 133 \ \mu g / kg$, \emptyset conc.: 41 $\mu g / kg$, country: USA nuts

Pito \rightarrow beer, pito

Pleosporaceae → Pleosporales

Pleosporales → Dothideales

Plums may contain the following \rightarrow mycotoxins: \rightarrow patulin incidence: 1/6, conc.: 4 µg/kg, country: Sweden

Polenta \rightarrow maize grits

Polydypsia excessive thirst

Polyuria excessive urination

Popcorn may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 5/15, conc. range: 20-47 μ g / kg, Ø conc.: 35 μ g / kg, country: Brazil incidence: 3/28, conc. range: ≤ 1.5 μg/kg, country: Germany \rightarrow aflatoxin G₁ incidence: 1/15, conc.: 18 µg/kg, country: Brazil \rightarrow aflatoxin G₂ incidence: 1/15, conc.: 8 µg/kg, country: Brazil \rightarrow deoxynivalenol incidence: 2/12*, conc. range: 12-250 µg/kg, country: Japan, *import from USA incidence: 7/7, conc. range: 12-250 μg/kg, country: USA incidence: 1/1, conc.: 30 μ g/kg, country: USA

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 \rightarrow fumonisin B₁ incidence: 4/6, conc. range: 10-60 µg/kg, \emptyset conc.: 28.3 µg/kg, country: Italy incidence: 6/6, conc. range: ca. < 10-122 μ g / kg, Ø conc.: 70 μ g / kg, country: Germany incidence: 13/29, conc. range: < 10-160 μg/kg, country: Germany incidence: 7/22, conc. range: ≤ 1003 μ g / kg, Ø conc.: 347 μ g / kg, country: Thailand incidence: 5/5, conc. range: < 100-500 μ g / kg, Ø conc.: 100 μ g / kg, country: USA incidence: 2/2, conc. range: 10-60 µg/kg, \emptyset conc.: 35 µg/kg, country: USA \rightarrow fumonisin B₂ incidence: 1/6, conc.: 20 µg/kg, country: Italy incidence: 7/22, conc. range: \leq 273 μ g / kg, Ø conc.: 116 μ g / kg, country: Thailand \rightarrow fumonisins incidence: 5/5, conc. range: < 10-100 μ g / kg (HPLC), country: Germany incidence: 6/13*, conc. range: 14-784 μ g / kg, Ø conc.: 83 μ g / kg, country: UK, *popping and microwaveable corn incidence: 1/1, conc.: 250 µg/kg, country: USA → ochratoxin A incidence: 1/29, conc.: 1.4 µg/kg, country: Germany \rightarrow zearalenone incidence: 4/7, conc. range: 2.5-130 μ g / kg, Ø conc.: 38 μ g / kg, country: USA \rightarrow maize **Poppadoms** may contain the following

→ mycotoxins: → aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: nc/4, conc. range: 0.6-2 μ g/kg, country: UK

Porcine nephropathy \rightarrow Mycotoxic porcine nephropathy

Porcine pulmonary edema (Abbr.: PPE) This lethal disorder in swine due to the ingestion of fumonisin B_1 and FB_2 (\rightarrow fumonisins) causes severe lung \rightarrow edema and hydrothorax. Rapid death occurs after an acute onset of \rightarrow dyspnea, weakness, and cyanosis. Oral as well as intravenous administration induced the disease.

Pork Feed to tissue ratios of less than 100 (\rightarrow ochratoxin A/ \rightarrow pig kidneys) indicate an extensive \rightarrow carry over for this mycotoxin. The biological halflife of OTA in swine tissue is 4.5 days (oral administration). A several week withdrawal period would be necessary to eliminate all OTA residues after exposure to a contaminated dietary.

Since 1978 in Denmark the kidneys of all slaughtered pigs have been examined for macroscopic changes. Suspected kidneys are chemically analysed. The level for rejection of the entire carcass is 25 μ g OTA / kg pig kidney. This level ensures that the concentration in \rightarrow meat does not exceed 10 μ g OTA / kg because it could be shown that pig meat contains only ca. 40% of the OTA found in pig kidney.

→ Aflatoxin B₁ feeding studies show that the kidneys (followed by the liver) of pigs accumulate most aflatoxin residues, mainly aflatoxin M₁ and to a lesser extent AFB₁ and → aflatoxicol. Minor levels were found in muscle. In comparison to → cattle (ca. 18 days) pigs might require a shorter withdrawal period (ca. 7 days). Pork may contain the following → mycotoxins:

 \rightarrow ochratoxin A

incidence: 64/76*, conc. range: $\leq 1.3 \ \mu g / kg$, \emptyset conc.: 0.11 $\mu g / kg$, country: Denmark, *produced conventionally incidence: 4/7*, conc. range: $\leq 0.12 \ \mu g / kg$, \emptyset conc.: 0.05 $\mu g / kg$, country: Denmark, *produced ecologically

incidence: 1/12, conc.: 5 μg/kg, country: Yugoslavia

 \rightarrow pig blood, \rightarrow pig kidneys, \rightarrow pig liver, \rightarrow pig serum

Porridge may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A incidence: 3/6*, conc.: $\leq 0.3 \ \mu\text{g}/\text{kg}, \emptyset$ conc.: 0.10 $\ \mu\text{g}/\text{kg}$, country: Germany, *ready made incidence: 4/92*, conc.: $\leq 2 \ \mu\text{g}/\text{kg}, \emptyset$ conc. 0.10 $\ \mu\text{g}/\text{kg}$, country: Germany, * \rightarrow oats \rightarrow cereals

Port wine \rightarrow Wine

Potatoes Since artificial inoculation with *Fusarium sambucinum* or *F. sulphureum* resulted in the production of \rightarrow monoacetoxyscirpenol and \rightarrow diacetoxyscirpenol ($\leq 5 \ \mu g / g$ rot fresh weight) \rightarrow trichothecenes might be found in moldy potato \rightarrow tubers.

Potatoes may contain the following → mycotoxins: → deoxynivalenol

incidence: 4/17, conc.: nc, country: Canada

 \rightarrow sambutoxin

incidence: 9/21*, conc. range: 15.8-78.1 μ g/kg, Ø conc.: 49.2 μ g/kg, country: Korea, *rotten

Poultry Poultry tolerate relatively high levels of \rightarrow trichothecenes in their diet but only very small traces are transmitted into \rightarrow meat and eggs. Residues quickly decline to negligible levels if the contaminated diet is removed.

Poultry meat may contain the following \rightarrow mycotoxins:

 \rightarrow ochratoxin A

incidence: 62/113, conc. range: ≤ 0.18 $\mu g\,/\,kg,$ country: Denmark

PR toxin (Abbr.: PRT) is a 2-(acetyloxy)-2,3,3a,4,6,7b-hexahydro-3,3',3a-trimethyl-6-oxo-spiro[naphth[1,2-b]oxirene-5(1aH),2'-oxirane]-3'-carboxaldehyde which was first isolated from \rightarrow Penicillium roquefortii Thom chemotype I in 1973 (see Figure PR toxin).

CHEMICAL DATA Empirical formula: $C_{17}H_{20}O_6$, molecular weight: 320

Fungal Sources Penicillium roquefortii

NATURAL OCCURRENCE \rightarrow cheese, Blue

Тохісіту

causes degenerative changes in liver and kidney of rat

LD₅₀ (po): 58-100 mg kg/bw mice

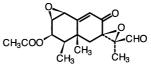
Detection HPLC, spectroscopy, TLC

Further Comments

If neutral and basic amino acids are present, PR-imines are formed. Compared to PR toxin, the toxicity of PR-imines is much lower. Further degradation products of PR toxin are PR-amide and the eremofortins A, B, C. The three latter ones are probably non-toxic.

Premature thelarche This \rightarrow mycotoxicosis may be induced by \rightarrow zearalenone.

Primary hepatocellular carcinoma (Abbr.: PHC) In various areas of central and southern Africa, Thailand, and Indonesia a high incidence of PHC in humans has been found which might be due to the ingestion of \rightarrow aflatoxins in the diet. A linear dose-response relationship between



PR toxin

the consumption of aflatoxins (0.003-0.222 μ g/kg bw) and human liver cancer has been demonstrated in several countries like Kenya, Mozambique, Uganda and Thailand.

Processed cheese \rightarrow cheese, processed

Proteinuria resulted in increased serum protein levels in the urine.

pulmonary pertaining the lung

Pulses Pulses may show a high contamination with \rightarrow ochratoxin A. They may be regarded as a possible contributor to OTA intake if they are regularly ingested. \rightarrow beans, \rightarrow cowpeas, \rightarrow lentils, \rightarrow peas, \rightarrow pigeon peas, \rightarrow soybeans

Pumpkin seeds may contain the following mycotoxins: \rightarrow aflatoxins incidence: 31/130, conc. range: nc, country: Germany

R

Ragi (*Eleusine coracan* (L.) Gaertn.) Fluctuation of temperature, change in relative humidity and excessive rainfall may contribute to \rightarrow Alternaria mycotoxin contamination under field conditions. Ragi may contain the following \rightarrow mycotoxins: \rightarrow altenuene incidence: 1/8, conc.: 30 µg/kg, country: India \rightarrow alternariol methyl ether incidence: 2/8, conc. range: 800-1400 μ g / kg, Ø conc.: 1100 μ g / kg, country: India \rightarrow tenuazonic acid incidence: 3/8, conc. range: 2030-5700 μ g / kg, Ø conc.: 3843 μ g / kg, country: India

Rape → oilseed rape

Rd-toxin (Syn.: \rightarrow deoxynivalenol)

Red mold toxicosis (Syn.: akakabi byo disease, red mold disease, red mold poisoning, scab disease) It takes its name from the reddish coloration of the predominately infected \rightarrow wheat and \rightarrow barley kernels. \rightarrow Fusarium graminearum Schwabe (Gibberella zeae) and other species like F. heterosporum, \rightarrow Fusarium nivale (Fr.) Ces., \rightarrow Fusarium poae (Peck) Wollenw., and \rightarrow Fusarium oxysporum Schlecht. emend. Snyd. & Hans. are mainly responsible for infection. Excessive rainfall and low temperatures throughout the ripening and harvest seasons favor the rate of invasion. In the severe development of this disease, more than 90% of the annual yield was damaged. Typically, people who ingested the discolored and shrivelled \rightarrow grains became ill from 5 to 30 min (2 h) after consumption. The following symptoms have been described: nausea, vomiting, diarrhea, feedrefusal, congestion or \rightarrow hemorrhage

in the lung, adrenals, intestine, uterus, vagina and brain, and destruction of the bone marrow. It was concluded that toxic metabolites of fusaria, especially \rightarrow deoxynivalenol, are responsible for this disease.

Red mold disease due to DON contamination has occurred sporadically during the years from 1946 to 1963 in several northern districts of rural Japan and in a southern area of Korea. The disease is akin to the wheat scab which has frequently been recorded in the USA. It became particularly widespread in Canada between 1980 and 1982. In India (Kashmir Valley) an outbreak of the disease for the last time in the 1980s was reported after the consumption of DONcontaminated wheat and \rightarrow wheat products (ca. 10,000 $\mu g \rightarrow$ trichothecenes/ kg). From 1961 to 1985 the disease affected at least 7818 victims and was attributed to consumption of scabby wheat and moldy \rightarrow maize; no deaths were reported. In China over a 20-year period ca. 10,000 cases of acute trichothecene toxicosis were reported but it is estimated that the real rate is significantly higher due to the difficulties of acquiring and evaluating such information from rural China and India.

renal pertaining to the kidney

Reye's syndrome A disease originally described in Australia by Reye but it is also known for children in Czechoslovakia, New Zealand, Thailand, and the U.S. In these cases \rightarrow aflatoxins have been implicated. Livers and blood serum from patients with Reye's syndrome contained aflatoxins. Children who suffer from an infection with influenza virus type A or B or with varicella were mainly (exclusively) affected. It seems that RS is a complex disease caused by a combination of factors including viral infection and

xenobiotic compounds possibly aflatoxins. The disease is characterized by clinical signs such as vomiting, convulsions, coma, and death within 24 h to 48 h after onset. Histopathological changes show acute encephalopathy, fatty degeneration of the liver (and kidney), pale, slightly widened \rightarrow renal cortexes (\rightarrow cortex), associated with elevated serum transaminase levels. A mortality rate of 81% of the originally diagnosed cases (21) has been reported. In Thailand the typical histopathological changes have been reproduced experimentally by feeding aflatoxin to Macaque monkeys. The involvement of aflatoxins in the Reye's syndrome was linked to the seasonal and geographic distribution of the disease (Thailand). Especially in rural areas there was a high incidence of death among children, which may be correlated with the aflatoxins detected in the food. There was also an increasing incidence of this disorder towards the latter part of the rainy season. This is typical for a \rightarrow mycotoxicosis (acute \rightarrow aflatoxicosis). From 1963 to 1974 more than 250 cases were reported in the US; 139 occurred in Thailand. In the following years (1973-1981) the RS incidence ranged from 0.37-0.88 per 100,000 per year, with a value of 2-4 during influenza epidemics (USA). Although a lower RS incidence was established in Britain during a five year study, a higher mortality rate (59%) occurred. According to the Center for Disease Control (USA) the following criteria are used to diagnose RS: (i) acute onset of encephalopathy, (ii) hepatic involvement (enlargement) of the liver or elevated serum transaminase levels (glutamic: pyruvic acid transaminase / oxalacetic acid transaminase), (iii) the disorder cannot be explained in any other way.

Rice Approximately one third of USstrains of \rightarrow Aspergillus flavus Link isola-

ted from rice produces significant amounts of \rightarrow aflatoxins. In addition, these aflatoxin producers make up a significant part of the normal mycoflora. Toxin formation probably results from rapid growth of the mold although limited penetration of the endosperm has been observed. Approximately 95% of the toxin is found in the \rightarrow bran layer. During ordinary \rightarrow milling procedures much of the aflatoxin in a contaminated kernel (rough rice) is removed. In consequence, the bran fractions contained ca. 10 times more aflatoxin than the milled rice fractions. Naturally contaminated rice may contain aflatoxins at levels of $< 4-50 \mu g / kg$. Since artificial drying is an efficient and effective operation, and since the toxin level in contaminated rice is greatly reduced by the milling process, contamination of rice with aflatoxins is not a serious problem at this time (USA). White rice, which is most widely used in human diets, did not contain any detectable levels of \rightarrow fumonisins. However, rough rice and rice hulls (feeding diets) contained fumonisin concentrations above 5 μ g/kg, indicating that fumonisins are localized primarily in the hulls and bran. However, since fumonisins are heat-stable they would probably not be destroyed by rice cooking methods and, to assure maximum safety, the rice should be monitored for fumonisin contamination (US). Studies documenting the fumonisin contamination of rice grown in other geographical areas are necessary.

Rice may contain the following \rightarrow mycotoxins:

 \rightarrow aflatoxin B₁

incidence: 2/52*, conc. range: 26-38 μ g / kg, Ø conc.: 32 μ g / kg, country: Brazil, *polished

incidence: 1/1, conc.: 8 µg/kg, country: Egypt

incidence: 1/50, conc.: 28 µg/kg, country: Italy incidence: 6/8, conc. range: < 2.5-15 μg/kg, country: Nepal incidence: 4/4*, conc. range: < 2.5-12.5 μg/kg, country: Nepal, *parboiled incidence: 7/364, \emptyset conc.: 20 µg/kg, country: Thailand incidence: 9/9*, conc. range: ≤ 600 μ g / kg, Ø conc.: < 1-2 μ g / kg, country: Thailand, *total: Ø conc.: 98 µg/kg AFB₁, AFB₂, AFG₁, AFG₂ incidence: 1/182, conc.: 5 μg/kg, country: USA \rightarrow aflatoxin B₂ incidence: 1/52*, conc.: 15 µg/kg, country: Brazil, *polished incidence: 1/1, conc.: 2 µg/kg, country: Egypt incidence: 1/4*, conc. range: 1.8 µg/kg, country: Nepal, *parboiled \rightarrow aflatoxin G₁ incidence: 1/52*, conc.: 20 µg/kg, country: Brazil, *polished incidence: 2/84, conc. range: 73.1-77.5 μ g / kg, Ø conc.: 75.3 μ g / kg, country: Malaysia \rightarrow aflatoxin G₂ incidence: 3/84, conc. range: 3.7-96.3 μ g / kg, Ø conc.: 45.6 μ g / kg, country: Malaysia aflatoxin (no specification) incidence: $3/15^*$, conc. range: ≤ 38 μ g / kg, Ø conc.: 16 μ g / kg, country: Philippines, *rice bran incidence: $17/82^*$, conc. range: ≤ 43 μ g / kg, Ø conc.: 12 μ g / kg, country: Philippines, *milled incidence:1/6*, conc.: \leq 3 µg/kg, \emptyset conc.: 3 µg/kg, country: Philippines, *pop incidence: $3/10^*$, conc. range: ≤ 18 μ g / kg, Ø conc.: 15 μ g / kg, country: Philippines, *rough \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: 13/30, conc. range: 22-317 µg AFB₁ / kg, 15-125 μg AFB₂ / kg, 14-107

 μ g AFG₁ / kg, 20-98 μ g AFG₂ / kg, country: India incidence: nc/4*, conc. range: 0.1-2.4 μg/kg, country: UK, *Basmati rice aflatoxins (no specification) incidence: 14/20, conc. range: 2-19 μ g / kg, Ø conc.: 7.9 μ g / kg, country: Gambia incidence: 12/80*, conc. range: tr-430 μg/kg, country: India, *cyclone-affected incidence: 23/81*, conc. range: 30-1130 μg / kg, country: India, *cyclone-affected incidence: 32/43*, conc. range: 30-130 μg/kg, country: India, *parboiled incidence: 1/23, conc.: 1000 µg/kg, country: Mozambique incidence: 16/72, conc. range: \leq 33 μ g / kg, Ø conc.: 16 μ g / kg, country: Philippines \rightarrow citrinin incidence: 4/30, conc. range: 49-92, country: India incidence: 2*/2, conc. range: 700-1130 μg/kg, country: Japan → deoxynivalenol incidence: $1/1^*$, conc.: 90 μ g/kg, country: Papua New Guinea, *imported, brown trukai incidence: nc/4*, conc. range: 4-6 µg/kg, country: UK, *Basmati rice incidence: $nc/4^*$, conc. range: 4-7 μ g/kg, country: UK, *Chinese rice \rightarrow fumonisin B₁ incidence: 8/20, conc. range: \leq 4300 μg/kg, country: USA \rightarrow fumonisin B₂ incidence: 6/20, conc. range: ≤ 1200 μg/kg, country: USA \rightarrow fumonisin B₃ incidence: 5/20, conc. range: ≤ 600 μg/kg, country: USA \rightarrow fumonisins (FB₁, FB₂) incidence: 1/4*, conc.: 28 μg/kg, country: UK, *Basmati rice \rightarrow nivalenol incidence: 2/9, \emptyset conc.: 22 µg/kg, country: Nepal

Rice

incidence: 1/1*, conc.: 63 µg/kg, country: Papua New Guinea, *imported incidence: nc/4*, conc. range: 4-11 μg/kg, country: UK, *Basmati rice \rightarrow ochratoxin A incidence: 1/3*, conc.: 533 µg/kg, country: Egypt, *rice germ incidence: 2/36, conc. range: $\leq 0.3 \ \mu g/$ kg, country: Germany incidence: 2/32, conc. range: 8-25 µg/kg, Ø conc.: 16.5 µg/kg, country: India incidence: 2/15, conc. range: 1.7-2.4 μg/kg, country: Indonesia incidence: 8/15, conc. range: $\leq 1.0 \ \mu g/$ kg, country: Italy incidence: 1/various \rightarrow food samples, conc.: 50 µg/kg, country: Japan incidence: 2*/2, conc. range: 230-430 µg/ kg, country: Japan, *deteriorated → sterigmatocystin incidence: 3/30, conc. range: 108-157 μg/kg, country: India incidence: 2/nc, conc. range: 50-450 μg/kg, country: Japan incidence: ?/?*, conc. range: 3800-4300 μg/kg, country: Japan, *moldy incidence: 12/37, conc. range: \leq 16,300 μg/kg, country: Japan T2-triol incidence: 1/4*, conc.: 49 µg/kg, country: UK, *Chinese rice \rightarrow zearalenone incidence: 1/9, conc.: 8 μ g/kg, country: Nepal incidence: 1/1*, conc.: 3060 µg/kg, country: Papua New Guinea, *imported incidence: 3/42*, conc. range: > 200 μg/kg, country: Uruguay, *and by-products Besides the \rightarrow mycotoxins listed under \rightarrow cereals rice may additionally be contaminated with \rightarrow citreoviridin, \rightarrow islanditoxin, \rightarrow luteoskyrin (Frisvad 1988). \rightarrow cereals

Rice bran may contain the following \rightarrow mycotoxins:

→ ochratoxin A
 incidence: 1/3, conc.: 9 µg/kg, country:
 Egypt
 → bran

Rice cake may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ & \rightarrow aflatoxin B₂ incidence: 1*/53**, conc.: \approx 10 µg/kg, country: Japan, *moldy, **different moldy samples \rightarrow ochratoxin A incidence: 1/3*, conc.: 4 µg/kg, country: Egypt, *rice germ cake

Roe deer may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 38/56*, conc. range: 0.3-2.17 μ g/kg, Ø conc.: 0.696 μ g/kg, country: Czechoslovakia, liver incidence: 39/56*, conc. range: 0.3-1.93 μ g/kg, Ø conc.: 0.795 μ g/kg, country: Czechoslovakia, kidney \rightarrow meat

Roquefort cheese \rightarrow cheese, blue; \rightarrow cheese, Roquefort

Roquefortine (Syn.: roquefortine C) is an indole alkaloid (10b-(1,1-dimethyl-2propenyl)-6,10b,11,11a-tetrahydro-3-(1Himidazol-4-ylmethylene)-2H-pyrazinol[1',2':1,5]pyrrolo[2,3-b]indole-1,4-(3H,5aH)-dione) which was originally named roquefortine C (\rightarrow mycotoxins). It was first isolated from \rightarrow Penicillium roquefortii Thom in 1975 by Japanese workers (see Figure Roquefortine).

CHEMICAL DATA Epmirical formula: $C_{22}H_{23}N_5O_2$, molecular weight: 389

FUNGAL SOURCES

 \rightarrow Penicillium spp. such as \rightarrow Penicillium chrysogenum Thom, \rightarrow Penicillium crustosum Thom, \rightarrow Penicillium expansum Link,

 \rightarrow Penicillium griseofulvum Dierckx, *P. roquefortii* chemotype I and II

NATURAL OCCURRENCE

→ cheese, Blue, → cheese, Blue Castello, → cheese, Danish Blue, → cheese dressing, blue, → cheese, Gorgonzola, → cheese, Roquefort, → cheese, Stilton

TOXICITY

 LD_{50} (ip): 15-189 mg/kg bw male mice DETECTION Electrochemical detectors, TLC

Further Comments

Roquefortine was detected in the stomach of several dogs. They showed a strychnine-like poisoning.

Roquefortine A & B (Syn.: isofumigaclavine A & B) roquefortine A (9-acetoxy-6,8-dimethylergolin) – roquefortine B (6,8-dimethylergolin-9-ol) is the hydrolysis product – and \rightarrow roquefortine have been isolated from the mycelium of \rightarrow Penicillium roquefortii Thom in 1975 (see Figure Roquefortine A & B).

CHEMICAL DATA

Empirical formula: $C_{18}H_{20}N_2O_2$, molecular weight: 296 (Roquefrotine A) Empirical formula: $C_{16}H_{20}N_2O$, molecular weight: 256 (Roquefrotine B)

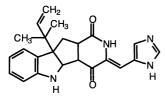
FUNGAL SOURCES

 \rightarrow Penicillium commune Thom chemotyope II, *P. clavigerum*, *P. roquefortii*,

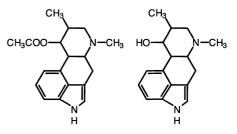
NATURAL OCCURRENCE \rightarrow cheese, Blue

TOXICITY

Roquefortine A is weakly toxic and possesses neurotoxic properties.



Roquefortine. Roquefortine C



Roquefortine A & B

 LD_{50} (ip): 340 mg roquefortine A and 1000 mg roquefortine B/kg bw mice. Weak pharmacological actions (e.g. muscle relactant, antidepressant, and local anaesthetic effects) have been observed.

Detection TLC

Roquefortine D (Syn.: dihydroroquefortine) a mycotoxin (12,13-dihydroroquefortine) which is a probable precursor of \rightarrow roquefortine C (\rightarrow mycotoxins). It is produced by *P. atramentosum*, \rightarrow Penicilium chrysogenum Thom, *P. glandicola*, and \rightarrow Penicillium roquefortii Thom.

Rubratoxins are complex nonarides with anhydride groups (relatively stable) and lactone rings (\rightarrow mycotoxins) which were first isolated as pure compounds in 1966 from a culture filtrate of \rightarrow Penicillium rubrum (see Figure Rubratoxins). Since difficulties in isolating the toxic fractions occur, the compounds were named rubratoxin A (10-[(R)-[(2R)-3,6-dihydro-6oxo-2H-pyran-2-yl]hydroxymethyl]-5,9,10,11-tetrahydro-4-hydroxy-5-[(1S)-1hydroxyheptyl]-1H-cyclonona[1,2-c:5,6c']difuran-1,3,6-trione (4S,5R,10R)) (more easily to isolate) and B. The more prevalent member was named rubratoxin B (10-[(R)-[(2R)-3,6-dihydro-6-oxo-2Hpyran-2-yl]hydroxymethyl]-5,9,10,11-tetrahydro-4-hydroxy-5-[(1S)-1-hydroxyheptyl]-1H-cyclonona[1,2-c:5,6-c´]difuran-1,3,6,8(4H)-tetrone (4S,5R,10R)) and is the more toxic.

Rubratoxins

CHEMICAL DATA

Empirical formula: $C_{26}H_{32}O_{11}$, molecular weight: 520 (Rubratoxin A) Empirical formula: $C_{26}H_{30}O_{11}$, molecular

weight: 518 (Rubratoxin B).

The fact that rubratoxin A is significantly more soluble in ethyl alcohol whereas rubratoxin B is significantly more soluble in ethyl acetate is important in fractionating mixtures of the two toxins.

FUNGAL SOURCES P. purpurogenum, P. rubrum

NATURAL OCCURRENCE

 \rightarrow tumeric, \rightarrow wheat

In addition, rubratoxins have been produced on \rightarrow maize by *P. purpurogenum* and *P. rubrum*.

Тохісіту

Although various effects on animals have been recorded, rubratoxin B is mainly hepatotoxic and nephrotoxic. Rubratoxin A possesses acute toxicity.

 LD_{50} (po): 120 mg/kg bw mice

DETECTION

HPLC, MS, RIA, spectroscopy, TLC

Possible Mycotoxicosis

Rubratoxin B was first implicated in "moldy corn toxicosis" in cattle, pigs, and poultry although their role in natural outbreaks of animals disease is, as yet, not clearly defined.

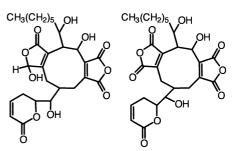
FURTHER COMMENTS

A synergistic action between \rightarrow aflatoxin B₁ and rubratoxin B, especially in the case of "hepatitis X", a toxicosis in dogs, is suggested.

Since rubratoxins are excreted into the medium and not retained by the mycelium (*P. rubrum*) they belong to the \rightarrow extracellular mycotoxins.

Rubratoxins are degraded by *P. puberu-lum*.

Rubratoxin (Syn.: rubratoxin B (\rightarrow rubratoxins))



Rubratoxins. Rubratoxin A & B

Rugulosin is a 2,2',4,4',5,5'-hexahydroxy-2,2',3,3'-tetrahydro-7,7'-dimethyl-1,1'-bianthraquinone (\rightarrow mycotoxins) (see Figure Rugulosin).

CHEMICAL DATA

Empirical formula: $C_{30}H_{22}O_{10}$, molecular weight: 542

FUNGAL SOURCES

→ Penicillium spp., e.g. → Penicillium islandicum Sopp (the (-) form), *P. rugulosum*, *P. variabile*, *Talaromyces wortmanii*

NATURAL OCCURRENCE It might be present in "yellow rice".

Тохісіту

antibiotic, hepatotoxic, carcinogenic LD_{50} (ip) : 83 mg/kg bw mice

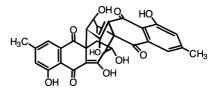
Detection TLC

Possible Mycotoxicosis Yellow rice disease

Further Comments

Thermal decomposition of rugulosin leads to the mycotoxins emodin and skyrin.

Long term feeding studies demonstrated the hepato-carcinogenicity of rugulosin



Rugulosin

and \rightarrow luteoskyrin and caused almost identical clinical signs.

Rye Rye seems to be more contaminated with \rightarrow ochratoxin A than \rightarrow wheat. Rye may contain the following \rightarrow mycotoxins: \rightarrow 3-acetyldeoxynivalenol incidence: 4/31, conc. range: 15-38 µg/ kg, \emptyset conc.: 24 µg/kg, country: Finland \rightarrow aflatoxin B₁ incidence: 1/2, conc.: 15 µg/kg, country: Germany incidence: 2/35, Ø conc. range: traces, country: USA \rightarrow alternariol incidence: 5/23, conc. range: 20-230 μ g / kg, Ø conc.: 85 μ g / kg, country: Poland \rightarrow alternariol methyl ether incidence: 1/8, conc.: 20 µg/kg, country: Germany incidence: 7/49, conc. range: 20-460 μ g / kg, Ø conc.: 117 μ g / kg, country: Poland \rightarrow citrinin incidence: 1/2, conc.: 960 μ g/kg, country: Canada \rightarrow deoxynivalenol incidence: 2/14, conc. range: 420-500 µg/ kg, \emptyset conc.: 460 µg/kg, country: Austria incidence: 1/1*, conc.: 56 µg/kg, country: Austria, *ecological incidence: 1/1, conc.: 204 µg/kg, country: Canada incidence: 8 products analysed, \emptyset conc.: 100 µg/kg, country: Canada incidence: 9/10*, conc. range: 10-47 μ g/kg, Ø conc.: 31 μ g/kg, country: Finland, *imported from Germany, Hungary, Soviet Union, Sweden, USA incidence: 20/50*, conc. range: ≤ 1250 μ g / kg, Ø conc.: 160 μ g / kg, country: Germany, *conventional incidence: 28/50, conc. range: \leq 500 μ g / kg, Ø conc.: 427 μ g / kg, country: Germany, *ecological

incidence: 4/22, Ø conc.: 406 µg/kg, country: Germany incidence: 1/2, conc.: 950 µg/kg, country: Germany incidence: 24*/31, conc. range: 30-2140 μ g / kg, Ø conc.: 330 μ g / kg, country: Germany, *moldy incidence: 1/23, conc.: 100 µg/kg, country: Germany incidence: 4/7*, conc. range: 31-86 μ g / kg, Ø conc.: 53.5 μ g / kg, country: Germany, *organic produce incidence: 24/31, conc. range: 9-93 μ g / kg, Ø conc.: 52 μ g / kg, country: Finland incidence: 5/5, Ø conc.: 1 μg/kg, country: Korea incidence: 4/4, conc. range: 8-384 µg/kg, \emptyset conc.: 106 μ g/kg, country: The Netherlands \rightarrow HT-2 toxin incidence: 1/31, conc.: 23 µg/kg, country: Finland incidence: 1/23, conc.: 100 µg/kg, country: Germany \rightarrow nivalenol incidence: 1/1, conc.: 8 μ g/kg, country: Canada incidence: 4/22, Ø conc.: 12 µg/kg, country: Germany incidence: 5/5, Ø conc.: 83 µg/kg, country: Korea incidence: 3/4, conc. range: 10-34 µg/kg, \emptyset conc.: 21 µg/kg, country: The Netherlands \rightarrow moniliformin incidence: 3/3*, conc. range: 6100-12,300 μ g / kg, Ø conc.: 9030 μ g / kg, country: Poland, *hand-selected, visible fungal damage \rightarrow nivalenol incidence: 1/31, conc.: $33 \mu g/kg$, country: Finland ochratoxin A incidence: 1/18, conc.: 2 μ g/kg, country: Austria

inicdence: 18/41, conc. range: 5-100 μg/kg, country: Austria incidence: 1/2, conc.: ca. 480 µg/kg, country: Canada incidence: 177/503*, conc. range: 0.05-4.9 μ g / kg (157 samples), 5-25 μ g / kg (16 sa), > 25-121 μg/kg (4 sa), Ø conc.: 1.2 μg/kg, country: Denmark, *conventional incidence: 71/91*, conc. range: 0.05-4.9 μ g / kg (55 samples), 5-25 μ g / kg (12 sa), > 25-120 μ g/kg (4 sa), Ø conc.: 5.4 μg/kg, country: Denmark, *ecological incidence: 8/22*, conc. range: 0.05-0.7 μ g/kg, Ø conc.: 0.1 μ g/kg, country: Denmark, *conventional, imported incidence: 149/682, conc. range: \leq 4.9 μg/kg, country: Germany incidence: 4/45, conc. range: 0.3-4.7 μg/kg, country. Germany incidence: 5/29, conc. range: 50-800 μ g / kg, Ø conc.: 354 μ g / kg, country: Poland incidence: 62/228, conc. range: 5-2400 μg/kg, country: Poland incidence: 44/94, conc. range: \leq 4.9-28 μ g / kg, country: Sweden incidence: 2/12, conc. range: \leq 16.7 μg / kg, country: The Netherlands incidence: 5/14*, conc. range: 0.1-16.8 μ g / kg, country: The Netherlands, *imported \rightarrow penicillic acid incidence: 1/29, conc. 2400 µg/kg, country: Poland \rightarrow T-2 toxin incidence: 1/31, conc.: $17 \mu g/kg$, country: Finland incidence: 10/25, conc. range: 200-700 μg/kg, country: Germany \rightarrow zearalenone incidence: 5/14, conc. range: 5-10 µg/kg, \emptyset conc.: 9 µg/kg, country: Austria incidence: 1/1, conc.: 2 µg/kg, country: Canada incidence: 9/50*, conc. range: \leq 7 µg/kg, \emptyset conc.: 4 µg/kg, country: Germany, *conventional

incidence: $5/50^*$, conc. range: ≤ 199 μ g / kg, Ø conc.: 51 μ g / kg, country: Germany, *ecological incidence: 3/22, \emptyset conc.: 5 µg/kg, country: Germany incidence: 15/31, conc. range: \leq 100 μ g / kg, Ø conc.: 17 μ g / kg, country: Germany incidence: 1/26, conc.: $< 70 \ \mu g / kg$, country: Germany incidence: 3/5*, conc. range: 3-4 µg/kg, country: Korea, *polished incidence: 1/29, conc.: 2000 µg/kg, country: Poland incidence: 1/4, conc.: 11 μ g/kg, country: The Netherlands → cereals

Rye bran may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 1/1, conc.: 150 µg/kg, country: Austria \rightarrow ochratoxin A incidence: 2/3, conc. range: $\leq 0.6 µg/kg$, country: The Netherlands \rightarrow zearalenone incidence: 1/1, conc.: 30 µg/kg, country: Austria \rightarrow bran

Rye flour may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 3/21, conc.: 150-335 µg/kg, \emptyset conc.: 272 µg/kg, country: Austria incidence: 3 products analysed, \emptyset conc.: 120 µg/kg, country: Canada incidence: 1/1, conc.: 174 µg/kg, country: Germany incidence: 2/2*, conc. range: 55-56 µg/kg, \emptyset conc.: 55.5 µg/kg, country: Germany, *organic incidence: 1/1, conc.: 33 µg/kg, country: Germany ergocornine (\rightarrow ergot alkaloids)

incidence: 4/4, conc. range: 1.8-6 µg/kg, country: Canada ergocristine incidence: 4/4, conc. range: 9.6-31 µg/kg, country: Canada ergometrine incidence: 4/4, conc. range: 1.2-9.3 μ g / kg, country: Canada ergosine incidence: 4/4, conc. range: 1.4-5.3 μg/kg, country: Canada ergotamine incidence: 4/4, conc. range: 5.5-23 µg/kg, country: Canada α-ergokryptine incidence: 4/4, conc. range: 2.4-7 µg/kg, country: Canada \rightarrow nivalenol incidence: 1/1, conc.: $3 \mu g / kg$, country: Germany \rightarrow ochratoxin A incidence: 4/35, conc. range: 0.1-17.7 µg/ kg, country: Germany

incidence: 2/15, conc. range: \leq 1.8 µg/ kg, country; Ø conc.: 0.28 μg/kg, Germany incidence: 11/11, conc. range: tr-20 μg/kg, country: Japan incidence: 8/14, conc. range: $\leq 1.2 \ \mu g$ / kg, Ø conc.: 0.3 μ g/kg, country: Sweden \rightarrow zearalenone incidence: 1/21, conc.: 10 µg/kg, country: Austria \rightarrow flour **Rye grits** may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A incidence: 2/15, conc. range: 1.7-1.8 μ g / kg, Ø conc.: 1.75 μ g / kg, country: Germany

 \rightarrow barley grits, \rightarrow maize grits, \rightarrow wheat grits

S

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Sago (\rightarrow cassava starch) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 2*/65, Ø conc. 150 µg/kg, country: Thailand, *total: Ø conc.: 294 µg/kg AFB₁, AFB₂, AFG₁, AFG₂

Sago hemolysis This disease which has been reported in Papua New Guinea since 1974 affects both males and females. Several clinical signs such as severe \rightarrow anemia, sudden onset of \rightarrow jaundice, and dark red urine are suggestive of hemolysis. In addition, fever and vomiting occurred in some cases, mental confusion and loss of consciousness in the worst cases. A mortality rate of almost 20% has been observed although blood transfusions were given.

"Stale" sago was suggested as being the cause of the disease because the patients consumed it the day before the onset of symptoms. Microscopical examination revealed bacterial and fungal cells but no fungal hyphae which would indicate excessive fungal growth. Unsuspicious, non-pathogenic microorganisms such as \rightarrow Paecilomyces *lilanicus* and *Tilletiopsis minor* (one colony of each), two yeast species and *Bacillus* spp. were isolated after plating the sago on agar. So far no toxin (\rightarrow mycotoxins) has been detected in the sago.

Saint Anthony's fire \rightarrow Ergotism

Salami may contain the following → mycotoxins: → aflatoxin B₁ incidence: 1/1, conc.: 5 µg/kg, country: Germany → sausages

Sambutoxin is a mycotoxin (\rightarrow mycotoxins) (4-hydroxy-5-(-4hydroxyphenyl)-1-

M. Weidenbörner, *Encyclopedia of Food Mycotoxins* © Springer-Verlag Berlin Heidelberg 2001

methyl-3-[(2R,5S,6S)-tetrahydro-5methyl-6-[(1E,3R,5S)-1,3,5-trimethyl-1heptenyl]-2H-pyran-2-yl]-2(1H)-pyridinone) which was first isolated in 1994 (see Figure Sambutoxin).

CHEMICAL DATA

Empirical formula: $C_{28}H_{40}NO_4$, molecular weight: 453

FUNGAL SOURCES

mainly \rightarrow Fusarium sambucinum Fuckel and \rightarrow Fusarium oxysporum Schlecht. emend. Snyd. & Hans.

NATURAL OCCURRENCE

 \rightarrow potatoes

This mycotoxin was not only found in rotten Korean potatoes but also in potatoes from parts of Iran where humans showed a high incidence of esophageal cancer.

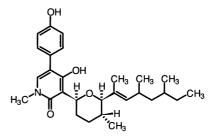
TOXICITY

hemorrhagic (\rightarrow hemorrhage) (stomach, intestines of rats), feed refusal, weight loss LD₅₀ : 29.6 µg/egg (chicken) DETECTION

HPLC

Sarcoma is a tumor composed of connective-like tissue.

Sausages \rightarrow Ochratoxin A seems to be the most important mycotoxin in sausages. The use of OTA-containing \rightarrow meat/ and/or organs is the main cause for the contamination of sausages.



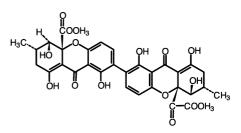
Sambutoxin

Sausages

Aflatoxin contamination of sausages is primarily due to the use of mycotoxin contaminated \rightarrow spices and / or the incorporation of aflatoxin producing fungi. Sausages may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/25, conc.: 7 µg/kg, country: Egypt \rightarrow aflatoxin B₂ incidence: 1/25, conc.: $3 \mu g / kg$, country: Egypt incidence: 1*/5, conc.: 7 µg/kg, country: Germany, *German Rohwurst \rightarrow aflatoxin G₂ incidence: 1/5, conc.: 30 µg/kg, country: Germany, *German Rohwurst ochratoxin A incidence: 20/125*, conc. range: 0.1-3.4 μ g / kg, Ø conc.: 0.9 μ g / kg, country: Germany, *cooked, black pudding incidence: 19/100*, conc. range: 0.1-1.7 μ g / kg, Ø conc.: 0.3 μ g / kg, country: Germany, *liver-type incidence: 19/100*, conc. range: 0.1-3.2 μ g / kg, Ø conc.: 0.8 μ g / kg, country: Germany, *Bologna-type incidence: 19/100*, Ø conc.: 3.4 μg/kg, country: Germany, *scalding incidence: 1/12*, conc.: 0.8 µg/kg, country: Switzerland, *scalding incidence: $4/32^*$, conc.: $\leq 1.8 \, \mu g / kg$, \emptyset conc.: 0.6 μ g/kg, country: UK, *black pudding incidence: 25/206*, conc. range: 10-920 μg/kg, country: Yugoslavia, *total of smoked meat products \rightarrow salami Scabby grain intoxication \rightarrow Red mold

disease \rightarrow Red

Scented supar may contain the following \rightarrow mycotoxins: \rightarrow patulin incidence: 1/147, conc.: nc, country: India



Secalonic acids. Secalonic acid D

Secalonic acids represent a group of six diastereoisomeric toxic fungal pigments (ergochromes, xanthone dimers) initially isolated in 1965 and 1966 from cultures of \rightarrow Claviceps *purpurea*. The most important member is secalonic acid D (see Figure Secalonic acids).

Chemical Data

Empirical formula: C₃₂H₃₀O₁₄, molecular weight: 638 (all secalonic acids)

FUNGAL SOURCES

Secalonic acids are produced by the five fungal genera \rightarrow Aspergillus, *Claviceps*, \rightarrow Penicillium, \rightarrow Phoma, and *Pyreno-chaeta*. Secalonic acid D is the major toxic fungal metabolite of *P. oxalicum*.

NATURAL OCCURRENCE

→ maize dust 300-4500 µg secalonic acid D/kg, grain dust (secalonic acid D); Secalonic acids are produced on a variety of substrates (→ grains) suitable for human consumption such as → barley, maize, → rice, → sorghum, → soybeans, and → wheat.

Τοχιςιτή

toxic to mice and rats, \rightarrow teratogenic, possibly \rightarrow mutagenic.

 LD_{50} (po) : 24.6 mg/kg bw new borne rats

DETECTION ELISA, HPLC, TLC

ELISA, III LC, ILC

Further Comments

The six secalonic acids A-G (B = E) are known.

Secalonic acids may be involved in \rightarrow dyspnea, grain fever and airway

obstruction of grain workers. Since secalonic acid D seem to be produced almost exclusively in stored grain (maize), proper grain storage should inhibit contamination. Even in fungal-contaminated maize little or no secalonic acid D could be detected prior to harvest.

Semi-hard cheese → cheese (semi-hard)

Semolina \rightarrow maize grits

Septic angina → Alimentary toxic aleukia

Sesame oil may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A incidence: 1/3, conc.: 0.4 µg/kg, country: UK \rightarrow oil

Sesame seeds may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 4*/19, conc. range: 4-10 $\mu g / kg$, country: Germany, *moldy incidence: 2*/75, Ø conc.: < 1 $\mu g / kg$, country: Thailand, *total Ø conc.: < 10 / kg AFB₁, AFB₂, AFG₁, AFG₂ \rightarrow nivalenol incidence: 2/7, Ø conc.: 10 $\mu g / kg$, country: Yemen \rightarrow ochratoxin A incidence: 3/3, conc.: \leq 0.4 $\mu g / kg$, country: UK

Sherbet may contain \rightarrow aflatoxin M_1 if it is made from naturally AFM₁ contaminated \rightarrow milk. The toxin remained stable during 8 months of frozen storage.

Sherry \rightarrow wine

Shoshin-kakke → Acute cardiac beriberi

Shoyu may contain \rightarrow aflatoxins if (i) \rightarrow Aspergillus flavus Link or \rightarrow Aspergillus parasiticus Speare are used to make koji for soy sauce (ii) the koji may be contaminated with an aflatoxin producer. The presence of Lactobacillus delbrueckii does not enable an aflatoxin free product to be made. Shoyu may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin G₂ incidence: 1/149, conc.: nc, country: Taiwan aflatoxin incidence: 1/nc, conc.: nc, country: Hong Kong \rightarrow citrinin incidence: nc, conc.: nc, country: China

 \rightarrow Oriental fermentations

Shrimps (fried with pork, garlic, & chilli peppers) may contain the following → mycotoxins: → aflatoxin B₁ incidence: 1/1, conc.: 207 µg / kg, country: Thailand, *total: 355 µg AFB₁, AFB₂, AFG₁, AFG₂ / kg → fish

Small grains Small grains (\rightarrow barley, \rightarrow millet, \rightarrow oats, \rightarrow rice, \rightarrow rye, \rightarrow sorghum, \rightarrow wheat) are only very rarely contaminated by \rightarrow aflatoxins. Small grains may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins incidence: 19/3489, \emptyset conc.: 5 µg/kg, country: USA

Snack foods may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 25/44, conc. range: \leq 450 μ g/kg, country: USA \rightarrow ochratoxin A incidence: 4/11, conc. range: 0.1-0.49 μ g/kg, country: Germany

Snack foods

 \rightarrow zearalenone incidence: 1/9, conc.: 2.9 µg/kg, country: USA

Soft drinks may contain the following \rightarrow mycotoxins: \rightarrow patulin incidence: 2/24, conc. range: 2-10 µg/kg, country: Germany \rightarrow apple juice, \rightarrow breakfast drinks, \rightarrow fruit juices, \rightarrow grape juice

Solaniol → neosolaniol

Sordariales → Ascomycota

Sorghum (*Sorghum* spp.)

Good quality sorghum does not seem to be contaminated by \rightarrow Alternaria mycotoxins. However, in weathered and discolored sorghum (U.S.) which was repeatedly wetted and then dried during rainy periods, the two \rightarrow Alternaria metabolites \rightarrow alternariol (AOH) and \rightarrow alternariol methyl ether (AME) were detected. A correlation between the degree of grain discoloration and rainy days during plant growth (September and October) and the level of alternariols was established. Fluctuation of temperature, change in relative humidity and excessive rainfall seem to promote Alternaria infection and subsequent toxin production in the seeds. As the number of rain-free days increased the AOH/AME level decreased. Contamination with \rightarrow alternariols might be due to several separate invasions of the maturing and mature seeds. \rightarrow Altenuene and \rightarrow tenuazonic acid were not detected, while altertoxin I (\rightarrow altertoxin I-III) occurred only in trace amounts in weathered sorghum. It was suggested, that the time for mycelial growth of Alternaria spp. was not sufficient for the synthesis of these late-produced metabolites. Wet conditions during or shortly after ripening of the grain contribute to

Alternaria mycotoxin contamination whereas the date of harvest was not decisive. Sorghum may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 3/6, conc. range: 70-120 μg/kg, country: Thailand incidence: 2/6, conc. range: 30-35 µg/kg, country: Tunisia incidence: 10/788, Ø conc.: 12 μg/kg, country: USA incidence: 6/533, conc. range: 3-19 μg/kg, country: USA \rightarrow aflatoxin B₂ incidence: 2/6, conc. range: nc, country: Thailand \rightarrow aflatoxin G₁ incidence: 1/6, conc.: nc, country: Thailand incidence: 3/533, conc. range: 3-19 μg/kg, country: USA \rightarrow aflatoxin (no specification) incidence: 2/2, conc. range: 29 µg/kg, country: Philippines \rightarrow aflatoxins incidence: 2/8, conc. range: 2-16 µg/kg, \emptyset conc.: 9 μ g / kg, country: Gambia incidence: 26*/69, conc. range: 1-100 μ g/kg (19 samples), 100-1000 μ g/kg (5 sa), > 1000 μ g/kg (1 sa), country: Uganda, * 16 samples contained AFB₁, 11 AFB₂, 13 AFG₁, 1 AFG₂ incidence: 4/786, conc. range: \leq 50 μ g / kg, Ø conc.: 17 μ g / kg, country: USA incidence: 6/533, conc. range: 3-19 µg/ kg, country: USA incidence: 103/200, conc. range: 1-100 μg/kg, country: USA incidence: 2/66, conc. range: 13-50 μ g / kg, Ø conc.: 61.5 μ g / kg, country: USA altenuene incidence: 3/12, conc. range: 120-1500 µg/ kg, Ø conc.: 670 μ g / kg, country: USA incidence: 5/20, conc. range: 20-700 μ g / kg, Ø conc.: 264 μ g / kg, country: India (Sorghum bicolor (L.) Moench)

alternariols* (alternariol and alternariol methyl ether) incidence: 21/63, conc. range: tr-7900 μg/kg country: USA, *weathered, discolored sorghum alternariol methyl ether incidence: 7/20*, conc.: 600-1800 µg/kg, \emptyset conc.: 1012 µg/kg, country: India, *Sorghum bicolor (L.) Moench altertoxin I incidence: 3/12, conc. range: traces, country: USA \rightarrow deoxynivalenol incidence: 31/32, conc. range: 1540 μ g / kg, Ø conc.: 190 μ g / kg, country: USA \rightarrow nivalenol incidence: 1/5, con.: 100 µg/kg, country: Yemen tenuazonic acid incidence: 5/20*, conc. range: 1300-5600 μ g / kg, Ø conc.: 3380 μ g / kg, country: India, *Sorghum bicolor (L.) Moench \rightarrow zearalenone incidence: 60/200, conc. range: 251-1500 μg/kg, country: USA incidence: 57/197, conc. range: 400 µg/kg (4 samples), 400-900 µg/kg (16 sa), 1000-5000 μ g/kg (35 sa), > 5000 μ g/kg (2 sa), country: USA \rightarrow cereals, \rightarrow millet

Sorghum meal may contain the following → mycotoxins: → fumonisin B₁ incidence: 2/2, conc. range: 20 µg/kg, Ø conc.: 20 µg/kg, country: Botswana incidence: 1/1, conc.: 28,200 µg/kg, country: Burundi → maize meal

Soy sauce → shoyu

Soybean concentrate may contain the following \rightarrow mycotoxins:

 \rightarrow ochratoxin A incidence: 1/2, conc. range: 50-500 $\mu g\,/\,kg,$ country: UK

Soybean flour may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins incidence: 1/4, conc.: nc, country: UK \rightarrow ochratoxin A incidence: 1/4, conc. range: 50-500 μ g/kg, country: UK incidence: 4/21*, conc. range: < 50-500 μ g/kg, country: UK, *defatted \rightarrow flour

Soybean milk powder may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin M₁ incidence: 1/8, conc.: 0.015-0.035 µg/kg, country: Italy

Soybeans Several factors such as high moisture content, number of splits and high total damage contribute to aflatoxin contamination of soybeans. However, generally soybeans are not a good substrate for aflatoxin production. The availability of zinc bound to phytic acid seems to be a decisive factor. With the breakdown of phytic acid due to heat or the addition of zinc increased aflatoxin production was observed in soybeans. Reddish and dark discolored soybean seeds indicate the potential presence of \rightarrow Fusarium mycotoxins, especially \rightarrow HT-2 toxin. Reddish seeds contained the highest mycotoxin concentrations with a maximum found in the seed coat. However, although in lower concentration HT-2 toxin was also present in the nonreddish seeds. The absence of reddish seeds therefore does not necessarily denote the absence of Fusarium mycotoxins.

Soybeans

Soybeans may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 2/866, conc. range: 7-10 μ g / kg, Ø conc.: 8.5 μ g / kg, country: USA incidence: 5/34, conc. range: < 5-20 μg/kg, country: USA \rightarrow aflatoxin G₁ incidence: 1/866, conc.: 4 µg/kg, country: USA \rightarrow aflatoxin (no specification) incidence: 12/25, conc. range: \leq 48 $\mu g / kg$, Ø conc.: 18 $\mu g / kg$, country: Philippines → deoxynivalenol incidence: 2?/30, conc. range: 490-1000 μg/kg, country: Canada incidence: $1/2^*$, conc.: 36 μ / kg, country: Papua New Guinea, *imported \rightarrow nivalenol incidence: $1/2^*$, conc.: 50 μ g/kg, country: Papua New Guinea, *imported \rightarrow HT-2 toxin incidence: 2?/30, conc. range: tr-330 µg/ kg, country: Canada → ochratoxin A incidence: 9/25, conc. range: < 50-500 μg/kg, country: UK \rightarrow zearalenone incidence: 2/17*, conc. range: > 200 μg/kg, country: Uruguay, *and by-products \rightarrow beans, \rightarrow cabbage, \rightarrow cowpeas, \rightarrow lentils, \rightarrow pigeon peas, \rightarrow peas, → vegetables

Spaghetti During cooking of spaghetti, average losses of \rightarrow deoxynivalenol amounted to 43-53% of the amount present before cooking. Spaghetti may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: nc, conc. range: $\leq 12.5 \ \mu g/kg$, country: Canada deoxynivalenol incidence: 7/7, conc. range: < 10-175 μ g / kg, Ø conc.: 89.3 μ g / kg, country: Austria incidence: 8/2*, conc. range: 2960-5020 μ g / kg, country: Canada * 2 wheat samples served for 8 different noodle preparation \rightarrow cereals

Spelt may contain the following mycotoxins: \rightarrow ochratoxin A incidence: 1/22, conc.: 0.9 µg/kg, country: Germany

Spices (no specification)

During growth in tropical climates, spices are often exposed to extremely wide ranges of rainfall, temperature and humidity. Although these factors in addition to their botanical origin may contribute to a pre-harvest mycotoxin contamination of the spices in a tropical environment, the sun drying process represents the greatest potential for contamination. The typical ground drying in the open air with high temperatures and humidities favor mold growth, especially \rightarrow Aspergillus spp., and subsequent mycotoxin (\rightarrow aflatoxins) production. During handling and storage mycotoxin contamination is also possible.

The largest amounts of spices are used in the \rightarrow meat industry where they represent a potential hazard for mold and mycotoxin contamination of the endproducts. However, due to their essential oils which reduce mold growth as well as aflatoxin production spices are not an ideal substrate for aflatoxin formation. In addition, spices generally are consumed in small amounts and therefore contribute little to the total health hazard posed by \rightarrow mycotoxins, especially aflatoxins. Spices (mixed) may contain the following mycotoxins:

 \rightarrow aflatoxin B₁ incidence: 5/37, conc. range: 0.2-0.8 μg/kg, country: Japan \rightarrow aflatoxin B₂ incidence: 5/37, conc. range: 0.2 µg/kg, country: Japan aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: $1/4^*$, conc.: 4 µg/kg, country: UK \rightarrow fumonisins (FB₁, FB₂) incidence: nc/4*, conc. range: 13-17 µg/ kg, country: UK \rightarrow ochratoxin A incidence: 5/108, conc. range: nc, country: India incidence: 1/4*, conc.: 2.6 µg/kg, country: UK \rightarrow zearalenone incidence: nc/4*, conc. range: 3.2-5.2 µg/ kg, country: UK *five spice powder According to Frisvad (1988) spices may be contaminated with the following mycotoxins: \rightarrow aflatoxins, \rightarrow citrinin, \rightarrow cyclopiazonic acid, \rightarrow luteoskyrin, \rightarrow ochratoxin A, \rightarrow penicillic acid, rubratoxin (\rightarrow rubratoxins), \rightarrow rugulosin, \rightarrow sterigmatocystin, \rightarrow viomellein, \rightarrow xanthomegnin. The following spices showed a contamination with mycotoxins: \rightarrow bay leaf, \rightarrow cardamom, \rightarrow cardamom, greater, \rightarrow cayenne pepper, \rightarrow chilli, \rightarrow chilli pickles, \rightarrow chilli powder, \rightarrow chilli sauce, \rightarrow coriander, \rightarrow cumin, \rightarrow curcuma, \rightarrow curry, \rightarrow curry paste, \rightarrow fennel, \rightarrow fenugreek, \rightarrow nutmeg, \rightarrow pepper, \rightarrow tandoori, \rightarrow turmeric

St. Nectaire cheese \rightarrow cheese, St. Nectaire

Starch may contain the following

 \rightarrow mycotoxins:

 \rightarrow fumonisin B₁

incidence: 1/1, conc.: 283 μg/kg, country: USA \rightarrow fumonisin B₂ incidence: 1/1, conc.: 70 $\mu g \,/\, kg$, country: USA

Sterigmatocystin as a furofuran (3*a*,12*c*dihydro-8-hydroxy-6-methoxyfuro[3',2',4,5]furo[3,2-*c*]xanthene-7-one) is a precursor in the biosynthesis of \rightarrow aflatoxin B₁ (see Figure Sterigmatocystin). It was originally isolated and named in 1954 (\rightarrow mycotoxins). In 1962 elucidation of its molecular structure followed. CHEMICAL DATA Empirical formula C₁₈H₁₂O₆, molecular weight: 324 FUNGAL SOURCES mainly \rightarrow Aspergillus versicolor (Vuill.) Tiraboshi and Emericella nidulans, further producers: e.g. \rightarrow Aspergillus spp. (ca. 20 different species), \rightarrow Emericella spp., \rightarrow Eurotium spp. Sterigmatocystin is an intermediate in the biosynthesis of \rightarrow aflatoxins by \rightarrow Aspergillus flavus Link and \rightarrow Aspergillus parasiticus Speare. NATURAL OCCURRENCE \rightarrow barley, \rightarrow breakfast cereals, \rightarrow cheese, \rightarrow cheese, Edam Cake, \rightarrow cheese, Gouda, \rightarrow cheese, Moravian Block, \rightarrow coffee beans, \rightarrow corn flakes, \rightarrow fennel, \rightarrow maize, \rightarrow oil seed rape, \rightarrow pecans, \rightarrow pepper, \rightarrow rice, \rightarrow wheat In general, isolation succeeded only from severely moldy substrates. Apart from that this mycotoxin is rarely found in nature. However, analytical methods for its detection are not as sensitive as for the \rightarrow aflatoxins. Therefore, low concentrations in \rightarrow food products may not always be detected. Residues in fresh meats are unlikely to be expected although in Canada sterigmatocystin has occasionally been detected in feeds. Here, a higher degree of sterigmatocystin contaminated \rightarrow grains in storage has also been reported.

Sterigmatocystin has also been reported to be a contaminant of marihuana.

Sterigmatocystin

TOXICITY

hepatotoxic, nephrotoxic, carcinogenic, \rightarrow mutagenic, \rightarrow teratogenic The toxic effects are much the same as those of aflatoxin B₁ but it is less acutely toxic.

 LD_{50} (po): 60-166 mg/kg bw rat In rat the metabolized sterigmatocystin is primarily secreted via the gastrointestinal tract and to a minor degree via the urine and feces within 12-24 hours.

DETECTION ELISA, GC-MS, HPLC, TLC

POSSIBLE MYCOTOXICOSIS Implication in the etiology of chronic liver disease in man in Africa is suggested.

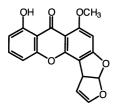
FURTHER COMMENTS

Sterigmatocystin was the first known substance of natural origin which contained the dihydrofurobenzofuran system.

At least eight derivatives are known. **Production:** A minimum $\rightarrow a_w$ of 0.80 is necessary for sterigmatocystin production on bread (*A. versicolor*), a_w 0.85 on agar media. The optimum a_w for production of this mycotoxin lays between 0.92-0.93.

Sterigmatocystin is only rarely found in \rightarrow foods which are usually visibly moldy. Despite its considerable carcinogenicity, it does not seem to be a significant hazard to human health.

Reduction / elimination: In milled brown rice sterigmatocystin concentration decreased gradually with a decrease in milling yield.



In contrast to the aflatoxins sterigmatocystin was stable in 2% ammonia solution.

Stilton cheese \rightarrow cheese, Stilton

Storage fungi The original source of these fungi is the field. They represent those microorganisms which are most tolerant to low water availability and therefore primarily grow on stored cereal \rightarrow grains. As xerophilic saprophytes they develop at relative humidities of 65-90% where free water is not available; e.g. a seed moistur of only $\approx 14\%$ (cereals) is sufficient for initial growth of \rightarrow Eurotium *halophilicum* and \rightarrow Aspergillus restrictus G. Sm. Eurotium spp. represent the most important genus within the group of storage fungi, growing at $\rightarrow a_w$ values from 0.62 to 0.75. Members of the genera \rightarrow Aspergillus and \rightarrow Penicillium are responsible for mycotoxin (\rightarrow mycotoxins) contamination. Mycotoxin production set in if the moisture content of cereal seeds rises to 13-16%. However, water activities of less than aw 0.70 which correspond to a moisture content of $\approx 15\%$ (most \rightarrow cereals) minimizes growth as well as mycotoxin production. Highest mycotoxin yields may occur at a water content of 20-25%. → field fungi

Sunflower seed oil may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 2/21, conc. range: 0.8-1 µg/kg, \emptyset conc.: 0.9 µg/kg, country: Germany \rightarrow aflatoxin G₁ incidence: 1/21, conc.: 0.3 µg/kg, country: Germany \rightarrow coconut oil, \rightarrow oil, \rightarrow olive oil, \rightarrow peanut oil

Sunflower seeds are a good substrate for aflatoxin production which may be due

Sterigmatocystin

to their high lipid content. However, the hard and thick seed coat impedes penetrability for aflatoxigenic fungi (\rightarrow Aspergillus flavus Link, \rightarrow Aspergillus parasiticus Speare) and should be responsible for low toxin production. Broken seeds gave substantially higher mycotoxin yields than whole seeds almost comparable to other \rightarrow oil seeds such as \rightarrow peanuts and \rightarrow soybeans. To prevent aflatoxin contamination sunflower seeds should be stored with the seed coat. Dehulling should be carried out just prior to extraction of \rightarrow oil. The phenomenon of a hard seed coat protecting seeds against fungal penetration is also known from e.g. certain varieties of peanuts and cotton seeds. Sunflower seeds may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/4*, conc.: 10.5 µg/kg, country: Germany \rightarrow aflatoxin B₂ incidence: 1/4*, conc.: 0.5 µg/kg, country: Germany \rightarrow aflatoxin G₁ incidence: 1/4*, conc.: 0.4 µg/kg, country: Germany \rightarrow aflatoxin G₂ incidence: $1/4^*$, conc.: 0.03 µg/kg, country: Germany *moldy \rightarrow aflatoxins (no specification) incidence: 7*/136, conc. range: 5-19.9 μ g / kg, country: Canada, *AFB₁, AFB₂, AFG_1, AFG_2 incidence: 9/135, conc. range: 25-230 μg/kg, country: Tunesia

 \rightarrow alternariol incidence: 37/50, conc. range: 35-792 μ g/kg, Ø conc.: 166 μ g/kg, country: Argentina incidence: 128/150, conc. range: 50-676 μ g/kg, Ø conc.: 189 μ g/kg, country: Argentina incidence: 2/2*, conc. range: 357-1840 μ g / kg, Ø conc.: 1090 μ g / kg, country: Italy, *samples visibly affected by \rightarrow Alternaria rot \rightarrow alternariol methyl ether incidence: 31/50, conc. range: 90-630 μ g/kg, Ø conc.: 114 μ g/kg, country: Argentina incidence: 70/150, conc. range: 30-836 μ g/kg, Ø conc.: 202 μ g/kg, country: Argentina incidence: 1/2*, conc.: 129 µg/kg, country: Italy, *samples visibly affected by \rightarrow Alternaria rot \rightarrow cyclopiazonic acid incidence: 1/1*, conc.: 10,000 µg/kg, country: USA, *moldy → ochratoxin A incidence: 4/25, conc. range: 0.2-0.49 μ g / kg (2 samples), 1.5-9.99 μ g / kg (2 sa), country: Germany → tenuazonic acid incidence: 98/150, conc. range: 2500-15,796 μg / kg, Ø conc.: 6459 μg / kg, country: Argentina

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Sweet potatoes \rightarrow tubers
Swine \rightarrow pork
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Swiss cheese \rightarrow cheese, Swiss

237

T

T-2 toxin belongs to the group of naturally occurring \rightarrow trichothecenes (3 α -hydroxy-4,15-diacetoxy-8 α -(3-methylbutyryloxy)-12,13-epoxytrichthec-9-ene) produced by different species of the genus \rightarrow Fusarium (see Figure T-2 toxin). During the search for causatives of the \rightarrow moldy corn toxicosis in 1966 besides \rightarrow diacetoxyscirpenol this new trichothecene, named T-2 toxin, could be isolated. The molecular structure was established in 1968.

CHEMICAL DATA

Empirical formula: $C_{24}H_{34}O_9$, molecular weight: 466

FUNGAL SOURCES

F. acuminatum, \rightarrow Fusarium avenaceum (Fr.) Sacc. (?), \rightarrow Fusarium culmorum (W. G. Smith) Sacc. (?), *Fusarium equiseti* (Corda) Sacc. sensu Gordon, \rightarrow Fusarium graminearum Schwabe, \rightarrow Fusarium oxysporum Schlecht. emend. Snyd. & Hans. (?), \rightarrow Fusarium poae (Peck) Wollenw., *F. semitectum*, \rightarrow Fusarium sporotrichioides Sherb.

NATURAL OCCURRENCE

 \rightarrow barley, \rightarrow beans, \rightarrow beer, \rightarrow curry,

→ ginger, → grains, → maize, → oats, → rye, → wheat

Although this mycotoxin is quite common in animal feed, T-2 toxin is a rare contaminant of \rightarrow foods.

Тохісіту

During metabolization of T-2 toxin into more hydrophilic compounds the trichothecane skeleton is not modified. dermatotoxic (like \rightarrow HT-2 toxin), emetic, \rightarrow immunosuppressive, cancerogenic (?) LD₅₀ (po): 4 mg/kg bw rat clinical symptoms: e.g. inflammation and hemorrhaging (\rightarrow hemorrhage) of the digestive tract, \rightarrow edema, \rightarrow leucopenia, degeneration of the bone marrow, and death (\rightarrow cattle, swine) inhibition fo the initiation step of protein synthesis on polyribosomes

Detection

ELISA, GC, HPLC, MS, RIA, TLC

Possible Mycotoxicosis

 \rightarrow alimentary toxic aleukie, \rightarrow moldy corn toxicosis

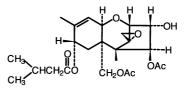
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FURTHER COMMENTS
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In general, T-2 toxin is an uncommon fungal secondary metabolite because most cereal \rightarrow grains are harvested appropriately.

Feeding studies reveal that T-2 toxin at levels typically encountered in contaminated feeds is extensively metabolized and rapidly eliminated from most of the host tissues (swine). The liver appears to be the primary site of residue accumulation. A higher transmission rate for T-2 toxin and T-2 metabolites into edible tissue occurred in the case of chick compared to pig. A hydroxy derivative and a deacetylated hydroxy derivative of the toxin are the major toxic metabolites found in tissue (*in vivo*). \rightarrow Carry over into the \rightarrow milk is much less than 1%. A synergistic effect with \rightarrow deoxynivalenol has been discussed.

Reduction / elimination: During the wet \rightarrow milling of maize the major portion (almost 70%) of T-2 toxin initially present was found in the steep and process water, 4% (8%) occurred in the starch whereas the rest was detected in the germ, gluten, and fiber.

No residues could be detected in \rightarrow oil prepared from the germ following the refining process.



T-2 toxin

M. Weidenbörner, *Encyclopedia of Food Mycotoxins* © Springer-Verlag Berlin Heidelberg 2001 Maize syrup will contain only low T-2 toxin levels because of the acidic processing conditions.

Table wine → wine

Tachycardia Excessive increase in heart rate.

Taco → Tortilla

Tandoori may contain the following \rightarrow mycotoxins: \rightarrow aflatoxins (AFB₁, AFB₂, AFG₁, AFG₂) incidence: nc/3, conc. range: 1.9-6.8 μg/kg, country: UK \rightarrow fumonisins (FB₁, FB₂) incidence: 1/3, conc.: 19 µg/kg, country: UK \rightarrow nivalenol incidence: nc/3, conc. range: 60-126 μ g / kg, country: UK \rightarrow ochratoxin A incidence: nc/3, conc.: $2.2-23.9 \mu g/kg$, country: UK T2-triol incidence: 1/3, conc.: 281 µg/kg, country: UK \rightarrow spices

Tapioca and products containing tapioca are starchy foods made from \rightarrow cassava. Tapioca may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A incidence: 3/17, conc. range: < 5 µg/kg, country: UK \rightarrow zearalenone incidence: 6/17, conc. range: < 5 µg/kg, country: UK

Taro may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁

incidence: $1^*/140$, conc.: $30 \ \mu\text{g}/\text{kg}$, country: Thailand, *total: $46 \ \mu\text{g}/\text{kg} \ AFB_1$, AFB₂, AFG₁, AFG₂

Temperature Environmental factors are decisive for mycotoxin production and one of the most important besides the \rightarrow a_w is temperature. In general, \rightarrow Penicillium spp. and \rightarrow Fusarium spp. need lower temperatures for the synthesis of \rightarrow mycotoxins, e.g. \rightarrow patulin: 0-24 °C \rightarrow Penicillium expansum Link, 4-31 °C P. patulum, \rightarrow Fusarium mycotoxins: 1.5 to 4 $^{\circ}C \rightarrow$ Fusarium sporotrichioides Sherb. than \rightarrow Aspergillus spp. (no patulin production below 12 °C). A similar pattern was also observed for ochratoxin production by \rightarrow Penicillium aurantiogriseum Dierckx?, \rightarrow Pencillium viridicatum Westling? (= Penicillium verrucosum Dierckx), and A. ochraceus. Penicillium species are able to produce mycotoxins over a broader range of temperature than Aspergillus spp. Since Penicillium spp. prefer temperate climatic regions (Northern Europe, Canada) their mycotoxins predominate in \rightarrow foods originating from these areas while Aspergillus species and their mycotoxins are more common in warmer climates (South-East Asia, Africa).

Tenuazonic acid is a 3-acetyl-5-*sec*-butyltetramic acid (3-acetyl-5-[(1S)-1-methylpropyl]-2H-pyrrolol-2-one (5S)-) produced by \rightarrow Alternaria spp. and other fungi (see Figure Tenuazonic acid). It was first isolated in 1957 (\rightarrow mycotoxins) and probably possesses the highest toxicity of all \rightarrow Alternaria mycotoxins.

Снемісаl Data Empirical formula: $C_{10}H_{15}NO_3$, molecular weight: 197

FUNGAL SOURCES

Alternaria spp. (most important \rightarrow Alternaria alternata (Fr.) Keissler), A. citri, A. japonica, A. kikuchiana, \rightarrow Aspergillus spp. (\rightarrow Aspergillus nomius Kurtzman et al.), Magnaporthe grisea (anamorph: Pyriculuria oryzae), \rightarrow Phoma sorghina.

NATURAL OCCURRENCE

→ apples, → mandarin fruits, → olives, → pepper, → ragi, → sorghum, → sunflower seeds, → tomatoes, → tomato paste, → wheat

Тохісіту

acutely very toxic, inhibition of protein synthesis, cardiovascular collapse, salivation, \rightarrow anorexia, erythema, \rightarrow convulsions, emesis, \rightarrow tachycardia, massive gastrointestinal hemorrhages (\rightarrow hemorrhage) etc. and death; antiviral, antibacterial, antifungal, phytotoxic and antitumor activity LD₅₀ (po): 81 / 168 mg / kg bw female /

male mice

DETECTION GC, HPLC, spectroscopy, TLC

Possible Mycotoxicosis

It is suggested that tenuazonic acid is involved in the etiology of a hematologic disorder named \rightarrow onyalai.

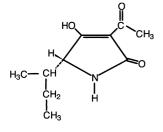
Further Comments

Tenuazonic acid seems to be one of the most important mycotoxins among the *Alternaria* mycotoxins.

It has been reported that tenuazonic acid occurred as magnesium, calcium, sodium and potassium salts in unacidified cultures.

teratogenic is a substance, causing malformations.

Tercinin (Syn.: \rightarrow patulin)





Tilsit cheese \rightarrow cheese, Tilsit

Time In general, mycotoxin production starts at the same time as the formation of conidia with an increase up to the period of intense sporulation. Subsequently a decrease in mycotoxin (\rightarrow mycotoxins) synthesis occurs sometimes associated with a metabolization of these secondary fungal metabolites.

Toast → bread

Tomato ketchup may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 2/18, conc. range: $\approx 1 \ \mu g/kg$, country: Germany

Tomato paste may contain the following \rightarrow mycotoxins: \rightarrow tenuazonic acid incidence: 6/8, conc. range: 10-100 μ g/kg, country: Canada incidence: 8/nc (several brands), conc. range: 0.01-0.1 μ g/kg, country: USA

Tomatoes Decay of the post-harvest tomato fruit (black rot lesion) is mainly due to \rightarrow Alternaria alternata (Fr.) Keissler. This \rightarrow black mold predominatly invades tomato tissue damaged by sun scald. Warm and rainy weather or dew formation on the fruit surface favors the disease. Tomatoes in the ripe stage are more susceptible than in the green stage. Substantial losses of tomatoes, especially those used for canning, have been reported. Fungal deterioration of the \rightarrow fruits is often associated with the contamination of \rightarrow Alternaria mycotoxins. In rotted tomatoes \rightarrow alternariol, \rightarrow aternariol methyl ether, and \rightarrow tenuazonic acid are the most common \rightarrow mycotoxins. Infections with \rightarrow Aspergillus flavus Link, A. niger and Rhizopus stolonifer are of minor importance.

Tomatoes

Tomatoes may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 1/8, conc.: 5 µg/kg, country: Germany \rightarrow altenuene incidence: 4/19, conc. range: < 100-1100 μ g / kg, Ø conc.: 100 μ g / kg, country: USA alternariol incidence: 1*/nc, conc.: 1274 µg/kg, country: Italy incidence: 6/19, conc. range: < 100-5.300 μ g / kg, Ø conc.: 300 μ g / kg, country: USA alternariol methyl ether incidence: 2*/nc, conc. range: 37-268 μg/kg, country: Italy incidence: 5/19, conc. range: < 100-800 μ g / kg, Ø conc.: 100 μ g / kg, country: USA tenuazonic acid incidence: 2*/nc, conc. range: 24-7210 μg/kg, country: Italy incidence: 73/142* (USA), conc. range: 400-1900 µg/kg (28 samples), 2000-70,000 µg/kg (45 sa), country: USA incidence: 11/19, conc. range: < 100-139,000 μg/kg, Ø conc.: 17,600 μg/kg, country: USA *samples visibly affected by Alternaria rot

Tortilla chips Experimental studies show that aflatoxin losses (\rightarrow aflatoxins) during cooking are associated primarily with the alkaline conditions. Tortilla chips may contain the following \rightarrow mycotoxins: \rightarrow fumonisin B₁ incidence: 8/12, conc. range: tr-216 µg/kg, country: Canada incidence: 1/2, conc.: 60 µg/kg, country:

Italy

incidence: 1/2, conc.: 30 μ g / kg, country: USA

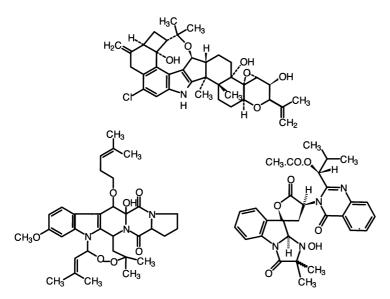
incidence: 2/2, conc. range: ca. 310-320 μg/kg, country: USA \rightarrow fumonisin B₂ incidence: 1/2, conc.: 10 µg/kg, country: Italy hydrolyzed fumonisin B₁ incidence: 2/2, conc.: present, country: USA \rightarrow fumonisins (no specification) incidence: 14/14*, conc. range: 200-1450 μ g / kg, country: USA, *white incidence: 1/1*, conc.: 400 µg/kg, country: USA, *yellow incidence: 2/2*, conc. range: 400-1000 μ g / kg, Ø conc.: 700 μ g / kg, country: USA, *blue incidence: 2/2*, conc. range: 300-400 μ g / kg, Ø conc.: 350 μ g / kg, country: USA, *organic blue \rightarrow maize

Tortillas Tortillas as a staple food in Mexico and Central America are traditionally made from \rightarrow maize. During cooking under alkaline conditions the high pH of the alkaline system seems to promote ionization of starch hydroxyl groups, producing Ca-starch crosslinks. This nixtamalization may hydrolyze \rightarrow fumonisin B₁ to HFB₁. The alkali processing during tortilla and tortilla-type food preparation causes an effective reduction in the amounts of \rightarrow aflatoxins in contaminated maize. This might be due to the initial soaking of the maize in lime water and a chemical change by alkali. Tortillas may contain the following \rightarrow mycotoxins: fumonisin B₁ incidence: 9/11*, conc. range: 24-612 μ g / kg, Ø conc.: 227 μ g / kg, country: Canada, *dried incidence: 7/7, conc. range: 210-1070 μ g / kg, Ø conc.: 601 μ g / kg, country: Mexico

incidence: 1/2, conc.: $120 \mu g/kg$, country: USA incidence: 1/3, conc.: 60 µg/kg, country: USA incidence: 50/52, conc. range: 12-672 μ g/kg, Ø conc.: 187 μ g/kg, country: USA / Mexico \rightarrow fumonisin B₂ incidence: 6/11*, conc. range: 26-218 μ g / kg, Ø conc.: 73.5 μ g / kg, country: Canada, *dried incidence: 6/7, conc. range: 50-180 μ g / kg, Ø conc.: 88.3 μ g / kg, country: Mexico incidence: 1/2, conc.: 30 µg/kg, country: USA hydrolyzed fumonisin B₁ incidence: 5/7, conc. range: 10-50 µg/kg, \emptyset conc.: 22 μ g/kg, country: Mexico incidence: 48/52, conc. range: 13-204 μ g/kg, Ø conc.: 82 μ g/kg, country: USA/Mexico \rightarrow fumonisins (FB₁, FB₂, FB₃) incidence: 6/20*, conc. range: 10-31 $\mu g / kg$, Ø conc.: 13 $\mu g / kg$, country: UK, *as well as taco and enchilada

fumonisins (no specification) incidence: 4/5, conc. range: \leq 800 µg/kg, country: USA

Tremorgenic mycotoxins There are only a few \rightarrow mycotoxins that act on the level of the central nervous system in vertebrate animals. Members of the first class like \rightarrow citreoviridin and steltoxin are responsible for respiratory arrest and \rightarrow paralysis. The tremorgenic mycotoxins which all possess an indole moiety from tryptophan belong to the second class and induce trembling in vertebrate animals. Based on chemical similarity (nitrogen content) the tremorgens are classified into three groups: \rightarrow penitrems A, B, and C (\rightarrow Penicillium spp.) as well as aflatrem $(\rightarrow Aspergillus flavus Link)$ which was the first isolated fungal tremorgen (1964) contain only one nitrogen per molecule and belong to group A; fumitremorgins A & B (\rightarrow Aspergillus fumigatus Fres.) and verruculogens (→ Aspergillus spp., Penicillium spp.) contain three nitrogen atoms per molecule and belong to group B;



Tremorgenic mycotoxins. Penitrem A, Fumigtremorgin A, Tryptoquivaline

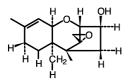
tryptoquivaline and tryptoquivalone $(\rightarrow \text{Aspergillus clavatus Desm.})$ contain four nitrogens per molecule and belong to group C (see Figure Tremorgenic mycotoxins). The members of the last group are comparatively less toxic than the remaining tremorgens. Although tremorgenic compounds are apparently uncommon in nature different fungal genera (Aspergillus, \rightarrow Claviceps, Penicillium) produce such mycotoxins. Informations e.g. about their natural occurrence are very limited. However, various naturally occurring neurological disorders, primarily of \rightarrow cattle ("staggers" syndromes) (e.g. paspalum, ryegrass, and corn staggers)) closely resemble the disorders produced under experimental conditions with fungal tremorgens.

Tremortin A (Syn.: penitrem A, \rightarrow penitrems)

Tremortin B (Syn.: penitrem B, \rightarrow penitrems)

Trichocomaceae → Eurotiales

Trichothecenes represent a family of chemically related sesquiterpenoids which all possess a tetracyclic 12,13-epoxy-trichothec-9-ene ring system (\rightarrow mycotoxins). They can be substituted at positions C-3, C-4, C-7, C-8, and C-15 (see Figure Trichothecenes 1). In 1967 the formerly called scirpenes (spiroepoxy-containing sesquiterpenoid compounds) were named trichothecenes. This name derived from the fungus *Trichothecium*. Trichothecin



Trichothecenes 1. Trichothecene nucleus

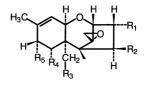
was the first trichothecene isolated from T. roseum in 1949 but correct chemical structure was not elucidated until 1964. The more than 170 known trichothecenes may be divided into simple (non-macrocyclic) and macrocyclic compounds. The latter ones contain a macrocyclic ring linking at C-4 and C-5 with diesters or triesters, e.g. verrucarins, roridins and satratoxins. There is little evidence that these compounds naturally occur in human food. The non-macrocyclic trichothecenes are divided into three groups A, B and C (see Figure Trichothecenes 2). Members of the first and largest group like \rightarrow T-2 toxin, \rightarrow HT-2 toxin, \rightarrow diacetoxyscirpenol, \rightarrow monoacetoxyscirpenol and \rightarrow neosolaniol do not contain a carbonyl group at C-8 (type A). Type B trichothecenes like \rightarrow nivalenol, \rightarrow deoxynivalenol, \rightarrow fusarenon X and diacetylnivalenol are characterized by the presence of a ketone group at C-8. An epoxide at C-7-8 is characteristic for crotocin (type C).

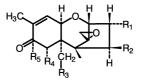
CHEMICAL DATA

For detailed information see each single trichothecene

FUNGAL SOURCES

Macrocyclic trichothecenes are produced by genera such as *Stachybotrys*, *Myrothecium*, *Cylindrocarpon*, *Phomopsis*, *Verticimonosporium*. The fusaria mainly pro-





Trichothecenes 2. Basic molecular structure of type A and B trichothecenes

duce the food relevant non-macrocyclic trichothecenes of type A (e.g. *F. acuminatum*, \rightarrow Fusarium equiseti (Corda) Sacc. sensu Gordon, \rightarrow Fusarium poae (Peck) Wollenw., \rightarrow Fusarium sambucinum Fuckel, \rightarrow Fusarium sporotrichioides Sherb.) and B (e.g. *Fusarium cerealis*, \rightarrow Fusarium culmorum (Wm. G. Smith) Sacc., \rightarrow Fusarium graminearum Schwabe). Crotocin (type C) is produced by *Trichothecium roseum*.

NATURAL OCCURRENCE

→ muesli , → muesli ingredients For further information see each single trichothecene as well as the primarily contaminated → cereals such as → barley, → maize, → wheat and → cereal products

TOXICITY

Although the *Fusarium* trichothecenes greatly vary in their toxicity they are acutely very toxic. T-2 toxin (type A) being probably the most toxic, and deoxynivalenol (type B) being among the least toxic. The A trichothecenes possess a 10 times higher toxicity than members of category B.

antibacterial, antiviral, antifungal, insecticidal (some), phytotoxic and cytostatic; primary mechanisms of toxicity are the inhibition of protein and DNA synthesis; clinical symptons: e.g. \rightarrow hypothermia, reduced respiratory rate, diarrhea, skin irritation and necrosis, emesis, hemorrhaging (\rightarrow hemorrhage), hematological changes (cellular damage in the bone marrow, spleen, and thymus leading to reductions in leucocytes and platelets $(\rightarrow \text{ aleukia})$ and reproductive problems LD₅₀: see each single trichothecene Swine and other monogastric animals (including humans) are most severely affected by these toxins. Affected animals are more susceptible to different fungal infections (e.g. Cryptococcus, Candida) and food-borne bacteria like Listeria and Salmonella which might be due to

immuno suppression (\rightarrow immunossuppressive). A high tolerance to trichothecenes was established in the case of chicken and turkey whereas ruminants were almost insensitive. If the contaminated feed source is removed there is an excellent prognosis for recovery for all species suffering from chronic trichothecene-induced toxicoses.

DETECTION

ELISA, GC-MS (best method), HPLC-MS (after derivatization), LC-MS, RIA, TLC

Possible Mycotoxicosis

→ alimentary toxic aleukia, "Cobalt-beer" cardiomyopathy, → moldy corn toxicosis, → pellagra, → red mold toxicosis.

FURTHER COMMENTS

Deoxynivalenol is the most common trichothecene in food and feed. Nivalenol, T-2 toxin, and HT-2 toxin occur to lesser extents whereas diacetoxyscirpenol is rarely isolated. It seems that trichothecenes are not very stable in cereals and feeds for longer periods. Almost no toxin could be detected in Finnish cereals after 3-6 months storage at 4 °C. However, overwintered cereals in the USSR remained toxic for several years, although no Fusaria could be isolated from the stored grains.

The naturally occurring combinations diacetoxyscirpenol / fusarenon X, diacetoxyscirpenol / deoxynivalenol and T-2 toxin / diacetoxysirpenol should act synergistically in laboratory animals. T-2 toxin synergized the negative effects of deoxynivalenol in swine. The natural combination of T-2 toxin / HT-2 toxin is synergistic in some ratios and antagonistic in other ratios.

It was speculated that these stable and relatively small molecules were used as chemical warfare agents, termed "yellow rain", in south-east Asia. However, it became obvious that this yellow rain resulted from a mass defecation of the Asian giant honey bee, *Apis dorsata*. **Production:** Temperatures below 10 °C favor the synthesis of trichothecenes whereas the greatest amounts are probably produced at low temperatures. Nevertheless, these mycotoxins have also been isolated from cereals grown in tropical (temperature 35 ± 5 °C) and subtropical areas. \rightarrow Malt is contaminated with trichothecenes only in very low concentrations (traces).

Reduction / elimination: Trichothecenes are very hard to remove from contaminated grains under moderate conditions. A transmission (up to 50%) into the endproducts like \rightarrow wheat flour, \rightarrow bread, crackers and \rightarrow baby cereals is therefore possible. Since trichothecenes are heat stable at 120 °C they probably survive the baking processes.

Alkali is effective in the destruction of trichothecenes.

Trichothecin → trichothecenes

Triticale may contain the following \rightarrow mycotoxins: \rightarrow alternario incidence: 3/19, conc. range: 80-250 μ g / kg, Ø conc.: 155 μ g / kg, country: Poland \rightarrow alternariol methyl ether incidence: 3/19, conc. range: 120-400 μ g / kg, Ø conc.: 229 μ g / kg, country: Poland → 3-acetyl deoxynivalenol incidence: 11*/50, conc. range: 1200-6000 μ g/kg, Ø conc.: 3600 μ g/kg, country: Poland, *healthy and damaged kernels, winter triticale \rightarrow citrinin incidence: 2/4, conc. range: 0.3-0.7 μ g/kg, Ø conc.: 0.5 μ g/kg, country: Switzerland \rightarrow deoxynivalenol incidence: 11/50, conc. range: 900-5900 μg/kg*, 2400-31,200 μg/kg**, Ø conc.:

10,109 µg/kg, country: Poland, *healthy kernels, **damaged kernels → moniliformin incidence: 3/3*, conc. range: 2600-15,700 µg/kg, Ø conc.: 8700 µg/kg, country: Poland, *hand-selected, visible fungal damage → ochratoxin A incidence: 9/10, conc. range: ≤ 5.6 µg/kg, Ø conc. 2.7 µg/kg, country: Germany

 \rightarrow cereals

Triticale flour may contain the following \rightarrow mycotoxins: ergometrine (\rightarrow ergot alkaloids) incidence: 2/2, conc. range: 13-31 µg/kg, country: Canada ergosine incidence: 2/2, conc. range: 9.3-16 µg/kg, country: Canada ergotamine incidence: 2/2, conc. range: 28-111 μg/kg, country: Canada ergocornine incidence: 2/2, conc. range: 12-26 µg/kg, country: Canada α -ergokryptine incidence: 2/2, conc. range: 17-21 µg/kg, country: Canada ergocristine incidence: 2/2, conc. range: 50-190 μg/kg, country: Canada

Tryptophan The indole nucleus of this important amino acid is frequently found in \rightarrow mycotoxins such as in the \rightarrow cyclopiazonic acid, \rightarrow ergot alkaloids, sporidesmins, and \rightarrow tremorgenic mycotoxins.

Tubers (ubi, gabi, tugi, singkamas, sweet potatoes) may be contaminated by \rightarrow aflatoxins due to poor storage conditions. Tubers may contain the following \rightarrow mycotoxins: aflatoxins (no specification) incidence: 6/59, conc. range: > 20- \leq 780 μg / kg, country: Philippines

Tugi \rightarrow tubers

Turkey Experimental studies show that feed tissue ratios of \rightarrow aflatoxin B₁ to AFB_1 and \rightarrow aflatoxin M_1 are high for kidney and liver but low for muscle. Turkey possess a high tolerance against \rightarrow trichothecenes. Turkey may contain the following \rightarrow mycotoxins: \rightarrow ochratoxin A incidence: 10/17, conc. range: ≤ 0.11 μ g / kg, Ø conc.: 0.02 μ g / kg, country: Denmark incidence: $3/17^*$, conc. range: ≤ 0.28 μ g / kg, Ø conc.: 0.04 μ g / kg, country: Denmark, *liver \rightarrow meat

Turkey "X" disease In 1960 a severe outbreak of the Turkey "X" disease occurred at 500 locations in Great Britain (mainly East Anglia and southern England) killing about 100,000 \rightarrow turkey poults. In addition, thousands of ducklings (\rightarrow duck) and young \rightarrow pheasants also died. Brazilian groundnut meal ("Rosetti meal") was the toxic factor which served as a protein source in the feed. The toxic factor was produced by \rightarrow Aspergillus flavus Link and \rightarrow Aspergillus parasiticus Speare which resulted in the name aflatoxin.

Using thin layer chromatography, the toxic factor could be separated into four distinct spots. These spots were named after their fluorescent color (blue, green) whereas the subscripts described their relative chromatographic mobility (\rightarrow aflatoxin B₁, \rightarrow aflatoxin B₂, \rightarrow aflatoxin G₁ and \rightarrow aflatoxin G₂). Although

the \rightarrow aflatoxins were responsible for at least the \rightarrow hepatic lesions and the high mortality they do not reproduce all signs of this disease, e.g. the strange attitudes of the head and neck. Therefore, it was suggested and proved that other \rightarrow mycotoxins like \rightarrow cyclopiazonic acid an other metabolite of *A. flavus* was also involved in Turkey "X" disease.

The Turkey "X" disease represents a turning point in mycotoxin research which greatly enhanced the scientific interest in the study of mycotoxins.

Turmeric is a dried rhizome of tropical origin. Way of mycotoxin contamination is not yet clear. Turmeric may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 2/15, Ø conc.: 12 µg/kg, country: Egypt incidence: 5/9, conc. range: 21-165 μg/kg, country: India incidence: 6/7, conc. range: tr-3.8 µg/kg, country: Canada \rightarrow aflatoxin B₂ incidence: 5/9, conc. range: 12-150 µg/kg, country: India \rightarrow aflatoxin G₁ incidence: 2/15, Ø conc.: 8 μg/kg, country: Egypt incidence: 5/9, conc. range: 20-125 μg/kg, country: India \rightarrow aflatoxin G₂ incidence: 5/9, conc. range: 14-125 μg/kg, country: India \rightarrow citrinin incidence: 2/9, conc. range: 48-52 µg/kg, \emptyset conc.: 50 µg/kg, country: India \rightarrow rubratoxin incidence: 1/9, conc.: 375 µg/kg, country: India \rightarrow spices

U

Ubi \rightarrow tubers

Urov disease → Kashin-Beck disease

V

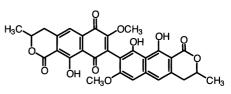
Vegetables (no specification) may contain the following \rightarrow mycotoxins: \rightarrow aflatoxin B₁ incidence: 4/51, conc. range: $< 5 \mu g / kg$, country: Germany \rightarrow aflatoxin incidence: 3/100*, conc. range: 2-20 μ g / kg (2 samples), > 20 μ g / kg (1 sa), country: Uruguay, *dried \rightarrow ochratoxin A incidence: 6/7*, conc. range: 245-7444 μg/kg, country: Tunesia, *chickpea, bean, lentil (dried) \rightarrow beans, \rightarrow cabbage, \rightarrow cowpeas, \rightarrow lentils, \rightarrow peas, \rightarrow pigeon peas, \rightarrow soybeans, \rightarrow tomatoes

Vermouth \rightarrow wine

Viomellein is structurally similar to \rightarrow xanthomegnin but is asymmetric due to the hydroxyl group at the 1' position and the lack of a ketone group at the 4' position (3,3',4,4'-tetrahydro-9',10,10'trihydroxy-7,7'-dimethoxy-3,3'-dimethyl-[8,8'-Bi-1H-naphtho[2,3-c]pyran]-1,1',6,9-tetrone). It is the second most naturally occurring fungal xanthoquinone (\rightarrow mycotoxins) (see Figure Viomellein).

Chemical Data

Empirical formula: $C_{30}H_{24}O_{11}$, molecular weight: 560



Viomellein

FUNGAL SOURCES

 \rightarrow Penicillium aurantiogriseum Dierckx, \rightarrow Penicillium crustosum Thom, *P. simplicissimum*, \rightarrow Penicillium viridicatum Westling, *Eupenicillium javanicum*, \rightarrow Aspergillus ochraceus group NATURAL OCCURRENCE

 \rightarrow barley, \rightarrow oil seed rape, \rightarrow wheat

In \rightarrow cereals, it often co-occurs with

 \rightarrow xanthomegnin and it may be associ-

ated with \rightarrow ochratoxin A and \rightarrow citrinin.

Тохісіту

toxicity similar to that of xanthomegnin, hepatotoxic, nephrotoxic (lesions)

Detection HPLC, TLC

Vomitoxin (Syn.: \rightarrow deoxynivalenol)

Vulvo-vaginitis \rightarrow F-2 toxicosis

W

Walnuts \rightarrow Aflatoxins seem to be the most important \rightarrow mycotoxins in walnuts whereas infection with aflatoxin-producing fungi is due to specific types of insects. It was estimated that the average probability of aflatoxin contamination in walnuts is one walnut in $28,250 \rightarrow$ nuts. The removal of visibly damaged nuts immediately after harvest and subsequent cool and dry storage conditions effectively prevent aflatoxin contamination. Walnuts may contain the following mycotoxins: \rightarrow aflatoxin B₁ incidence: 4/97, conc. range: $< 5 \mu g / kg$ (3 samples), conc.: 7 μ g/kg (1 sa), country: Germany incidence 3*/12, conc. range: 5-500,000 μg/kg, country: Germany, *moldy incidence: 1/14* conc.: 8 µg/kg, country: Norway, *imported \rightarrow aflatoxin B₂ incidence: 1/14* conc.: traces, country: Norway, *imported \rightarrow aflatoxin G₁ incidence: 1/14* conc.: 4 µg/kg, country: Norway, *imported \rightarrow aflatoxin G₂ incidence: 1/14* conc.: traces, country: Norway, *imported aflatoxins incidence: 10*/156, conc. range: 5-24.9 μ g/kg (9 samples), > 25 μ g/kg (1 sa), country: Canada, *AFB₁, AFB₂, AFG₁, AFG₂ incidence: 15/20, conc. range: 15-25 μg/kg, country: Egypt incidence: 4/97, conc. range: $< 5 \mu g / kg$ (3 samples), conc.: $18 \mu g / kg (1 sa)$, country: Germany incidence: 8/330, conc. range: 2-70 μ g / kg, Ø conc.: 27 μ g / kg, country: USA incidence: 2/27, conc. range: 29-41 μ g / kg, Ø conc.: 35 μ g / kg, country: USA

incidence: 2/4, conc. range: $\leq 8 \ \mu g / kg$, \emptyset conc.: 4 $\mu g / kg$, country: USA penitrem A (\rightarrow penitrems) incidence: 1/1*, conc.: nc, country: USA, *visibly moldy \rightarrow zearalenone incidence: 1/20, conc.: 125 $\mu g / kg$, country: Egypt incidence: 3/60, conc. range: 50-450 $\mu g / kg$, country: France \rightarrow nuts

Water activity $\rightarrow a_w$

Wheat is one of the most important of the cereal crops grown for human consumption. During moist weather periods the maturing seeds may be heavily invaded by \rightarrow Fusarium spp., especially \rightarrow Fusarium graminearum Schwabe the causal factor for Fusarium head blight. \rightarrow Fusarium culmorum (W. G. Smith) Sacc. and \rightarrow Fusarium avenaceum (Fr.) Sacc. are also very common on wheat. A reddish discoloration of the kernels may be associated with trichothecene contamination (\rightarrow trichothecenes). Compared to the testa the wheat embryo is an excellent substrate for aflatoxin production of \rightarrow Aspergillus flavus Link. However, \rightarrow aflatoxins do not play an important role in mycotoxin contamination of wheat. Wheat may contain the following \rightarrow mycotoxins: 3-acetoxynivalenol incidence: 3/27, conc. range: < 200 µg/kg, country: Finland → 3-acetyldeoxynivalenol incidence: 5/10, conc. range: 15-731 μ g / kg, Ø conc.: 363 μ g / kg, country: China incidence: 9/40, conc. range: 12-67 $\mu g / kg$, Ø conc.: 31 $\mu g / kg$, country: Finland

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Wheat

incidence: 50/84, conc. range: 3-18 μ g / kg, Ø conc.: 7 μ g / kg, country: Germany incidence: nc/9, conc. range: 100-30,000 μ g / kg, country: Poland incidence: 2/3*, conc. range: 100 µg/kg, \emptyset conc.: 100 µg/kg, country: Poland, *healthy and damaged kernels incidence: 13/13*, conc. range: 100-3000 (5600) μg/kg, Ø conc.: 790 μg/kg, country: Poland, *healthy and damaged kernels → 15-acetyldeoxynivalenol incidence: 3/3*, conc. range: 100-2000 μ g / kg, Ø conc.: 675 μ g / kg, country: Poland, *healthy and damaged kernels \rightarrow aflatoxin B₁ incidence: 40/545, Ø conc.: 16.3 μg/kg, country: Croatia incidence: 3/7*, conc. range: 10-15 µg/ kg, country: Germany, *moldy incidence: 1/11*, conc.: traces, country: UK, *moldy incidence: 23/31*, conc. range: 0.8-17 μ g / kg, Ø conc.: 3.37 μ g / kg, country: USA, *scabby incidence: 3/1.528, conc.: 11 μg/kg, country: USA incidence: 2/531, conc. range: 7 µg/kg, Ø conc.: 7 µg/kg, country: USA \rightarrow aflatoxin G₁ incidence: 2/531, conc. range: 2 µg/kg, \emptyset conc.: 2 µg/kg, country: USA aflatoxin (no specification) incidence: 29/123*, conc. range: 2-20 $\mu g / kg$ (28 samples), > 20 $\mu g / kg$ (1 sa), country: Uruguay, *and by-products aflatoxins (no specification) incidence: 10/30, conc. range: 15-263 µg AFB₁ / kg, 10-107 μg AFB₂ / kg, 12-95 μg AFG₁ / kg, 22-90 μ g AFG₂ / kg, country: India \rightarrow alternariol incidence: 27/33*, conc. range: ≤ 1050 μ g/kg, Ø conc.: 152 μ g/kg, country: Australia, *weather-damaged

incidence: 2/105, conc. range: 6-12 μ g / kg, Ø conc.: 9 μ g / kg, country: Germany incidence: 1/5, conc.: 590 µg/kg, country: Poland incidence: 9/49, conc. range: 20-600 $\mu g / kg$, \emptyset conc.: 131 $\mu g / kg$, country: Poland → alternariol methyl ether inicdence: 24/33*, conc. range: ≤ 46 μ g / kg, Ø conc.: 14.4 μ g / kg, country: Australia, *weather damaged incidence: 12/199, conc. range: 4-200 μ g / kg, Ø conc.: 37.3 μ g / kg, country: Germany incidence: 7/49, conc. range: 20-1600 μ g / kg, Ø conc.: 305 μ g / kg, country: Poland \rightarrow citrinin incidence: 10/15, conc. range: 70-80,000 μ g / kg, country: Canada incidence: 1/66, conc.: 2000 µg/kg, country: Poland incidence: 11/11*, conc. range: tr-4800 μg/kg, country: UK, *moldy \rightarrow deoxynivalenol incidence: 3/20, Ø conc.: 15 µg/kg, country: Argentina incidence: 56/60, conc. range: 100-9250 μ g / kg, Ø conc.: 1798 μ g / kg, country: Argentina incidence: 32/40, conc. range: 300-4500 μ g/kg, Ø conc.: 1060 μ g/kg, country: Argentina incidence: 11/12, conc. range: ≤ 6700 μ g/kg, Ø conc.: 1800 μ g/kg, country: Australia incidence: 3/4, Ø conc.: $360 \mu g / kg$, country: Austria incidence: 11/32, conc. range: 80-2110 μ g / kg, Ø conc.: 580 μ g / kg, country: Austria incidence: 4/16, conc. range: 27-1280 μ g/kg, Ø conc.: 449 μ g/kg, country: Austria incidence: 3/3*, conc. range: 465-4450 μ g/kg, Ø conc.: 3062 μ g/kg, country: Austria, *durum

incidence: 1/2, conc.: 211 μ g/kg, country: Bulgaria incidence: 55/199, conc. range: 20-1320 μg/kg, country: Canada incidence: 9/10, conc. range: 25-3475 μ g / kg, Ø conc.: 1257 μ g / kg, country: Canada incidence: 11/208, conc. range: 20-3200 μ g / kg, country: Canada incidence: 40/53*, conc. range: 50-3650 μ g / kg, Ø conc.: 434 μ g / kg, country: Canada, *suspected incidence: 412/560, conc. range: 10-5670 μ g / kg, Ø conc.: 460 μ g / kg, country: Canada incidence: 86/258, conc. range: 10-1510 μ g/kg, Ø conc.: 210 μ g/kg, country: Canada incidence: 270/1493*, conc. range: 10-10,500 μg / kg, Ø conc.: 430 μg / kg, country: Canada, *hard incidence: 5/5*, conc. range: 20-100 μ g / kg (1 sample), 101-500 μ g / kg (3 sa), > 500 µg/kg (1 sa), country: Canada, *soft incidence: 1/5, conc.: 1710 µg/kg, country: China incidence: 4/4, Ø conc.: $4284 \mu g / kg$, country: China incidence: 5/10, conc. range: 73-1051 μ g/kg, Ø conc.: 349 μ g/kg, country: China incidence: 25/27, conc. range: 1-6300 μg/kg, country: Finland incidence: 37/40, conc. range: 8-356 $\mu g / kg$, Ø conc.: 81 $\mu g / kg$, country: Finland incidence: 8/10*, conc. range: 10-68 $\mu g / kg$, Ø conc.: 35 $\mu g / kg$, country: Finland, *imported from Canada, Saudi-Arabia, USA incidence: 1/2, conc.: 86 µg/kg, country: France incidence: 1/1, conc.: 5000 µg/kg, country: France incidence: $45^{*}/51$, conc. range: ≤ 1200 μ g/kg, Ø conc.: 420 μ g/kg, country: Germany, *conventional

incidence: 38*/50, conc. range: ≤ 1000 μ g / kg, Ø conc.: 486 μ g / kg, country: Germany, *ecological incidence: 2/6, \emptyset conc: 712 µg/kg, country: Germany incidence: 14/44, conc. range: 10-5600 μ g / kg, Ø conc.: 810 μ g / kg, country: Germany incidence: 92/106*, conc. range: 70-43,800 μ g / kg, Ø conc.: 3960 μ g / kg, country: Germany, *moldy incidence: 43/45, conc. range: 40-750 μ g / kg, Ø conc.: 190 μ g / kg, country: Germany incidence: 140/154, conc. range: 40-3240 μ g / kg, Ø conc.: 170 μ g / kg, country: Germany incidence: 24/29*, conc. range: 10-2000 μg/kg, country: Germany, *food grade wheat and wheat products incidence: $2/8 \oslash$ conc.: 700 µg/kg, country: Germany incidence: 5/123, conc. range: 10-1300 µg/kg, country: Germany incidence: 7/10*, conc. range: 36-340 μ g / kg, Ø conc.: 176 μ g / kg, country: Germany, *organic produce incidence: 2/2, conc. range: 36-370 μ g/kg, Ø conc.: 203 μ g/kg, country: Germany incidence: 81/84, conc. range: 4-20,538 μ g/kg, Ø conc.: 1632 μ g/kg, country: Germany incidence: 1/1, conc.: 9 µg/kg, country: Greece incidence: 2/2, Ø conc.: 671 µg/kg, country: Hungary incidence: 1/12, conc.: 120 µg/kg, country: Italy incidence: 2/17, conc. range: 90-280 μg/kg, country: Japan inicdence: 4/6, \emptyset conc.: 23 µg/kg, country: Japan incidence: 95/101, conc. range: 10-12,400 μ g/kg, Ø conc.: 1178 μ g/kg, country: Japan incidence: 1/1, conc.: 440 µg/kg, country: Japan

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Wheat

incidence: 11/18, conc. range: ND-1800 μ g / kg, Ø conc.: 800 μ g / kg, country: Japan incidence: 8/11, conc. range: 100-9180 μ g / kg, Ø conc.: 1290 μ g / kg, country: Japan incidence: 2/3, conc. range: 160-370 μ g/kg, Ø conc.: 260 μ g/kg, country: Japan incidence: 18/18*, conc. range: 740-6920 μ g / kg, Ø conc.: 3812 μ g / kg, Japan, *scabby wheat incidence: 5/9, conc. range: \leq 170 µg/kg, \emptyset conc.: 42 µg/kg, country: Korea incidence: 1/10, conc.: 61 µg/kg, country: Nepal incidence: 78/90, conc. range: \leq 11,950 μg/kg, country: New Zealand incidence: 13/42, conc. range: \leq 310 μ g/kg, Ø conc.: 95 μ g/kg, country: Poland incidence: nc/9, conc. range: 200-30,400 μ g / kg, country: Poland incidence: 3/3*, conc. range: 2000-38,000 μ g / kg, Ø conc.: 16,216 μ g / kg, country: Poland, *healthy and damaged kernels incidence: 11/13*, conc. range: 400-39,600 μ g/kg, Ø conc.: 14,540 μ g/kg, country: Poland, *healthy and damaged kernels incidence: 1/2, conc.: 30 μ g / kg, country: Portugal incidence: 1/2, conc.: 26 μ g/kg, country: Scotland incidence: 31/43, conc. range: \leq 1180 μ g / kg, Ø conc.: 240 μ g / kg, country: Sweden incidence: 8/14, conc. range: 110-1180 μ g / kg, Ø conc.: 400 μ g / kg, country: Sweden incidence: 23/29, conc. range: 60-360 μ g/kg, Ø conc.: 190 μ g/kg, country: Sweden incidence: 12/22, conc. range: ≤ 2500 $\mu g / kg$, \emptyset conc.: 480 $\mu g / kg$, country: Taiwan

incidence: 9/12, conc. range: 45-2450 $\mu g / kg$, \emptyset conc.: 562 $\mu g / kg$, country: Taiwan incidence: 3/10, conc. range: 26-505 μ g/kg, Ø conc.: 245 μ g/kg, country: Taiwan incidence: 13/13, conc. range: 20-231 μ g/kg, Ø conc.: 115 μ g/kg, country: The Netherlands incidence: 1/35, conc.: 90 µg/kg, country: UK incidence: 20/31, conc. range: 4-312 μ g/kg, Ø conc.: 31 μ g/kg, country: UK incidence: 32/199, conc. range: 20-400 μg/kg, country: UK incidence: 23/33*, conc. range: 20-1320 µg/kg, country: UK, *imported incidence: 34/205, conc. range: 20-500 μg/kg, country: UK incidence: 1/35, conc.: 90 µg/kg, country: UK incidence: 6/55, conc. range: 80-750 μ g / kg, Ø conc.: 340 μ g / kg, country: UK incidence: 75/123, conc. range: tr-500 µg/kg (38 samples), conc. range: 500-1000 µg/kg (32 sa), conc. range: 1000-2000 μ g/kg (4 sa), conc. range: > 2000 μ g / kg (1 sa), country: USA incidence: 31/33*, conc. range: 120-5500 $\mu g / kg \emptyset$ conc.: 1782 $\mu g / kg$, country: USA, *scabby incidence: 132/247, conc. range: \leq 2650 μ g/kg, Ø conc.: 570 μ g/kg, country: USA incidence: 14/27, conc. range: 600-3800 μ g /kg, Ø conc.: 2800 μ g / kg, country: USA incidence: 23/116, \emptyset conc.: 100 μ g/kg, country: USA incidence: 12/14, conc. range: 20-100 μ g / kg (7 samples), 101-500 μ g / kg (4 sa), > 500 μ g/kg (1 sa), country: USA incidence: 156/157, conc. range: 200-43,000 µg/kg, country: USA incidence: 201/207, conc. range: 400-4000 $\mu g / kg$, country: USA incidence: 120/206, conc. range: 900-7600 μ g / kg, country: USA

incidence: 333/483, conc. range: 500-18,000 μg/kg, Ø conc.: 2000 μg/kg, country: USA incidence: 1/7, conc.: 5 μ g/kg, country: Yemen incidence: nc/5, conc. range: 3400-8000 μg/kg, country: Yugoslavia 4,7-dideoxynivalenol incidence: 3/3*, conc. range: 100-150 μ g / kg, Ø conc.: 113 μ g / kg, country: Poland, *healthy and damaged kernels → diacetoxyscirpenol incidence: 20/53*, conc. ranges: \leq 80 μg/kg, country: Canada, *suspected incidence: 1/87, conc.: 50 µg/kg, country: Germany incidence: 5/59, conc. range: 300-2000 μg/kg, country: Germany incidence: 3/100, conc. range: nc, country: France incidence: 1/nc, conc.: 50 μ g/kg, country: USSR ergocristine (\rightarrow ergot alkaloids) incidence: 1/1*, conc.: 0.2-0.3 µg/kg, country: Canada, *uncooked \rightarrow fusarenon X incidence: 3/27, conc. range: $< 40 \ \mu g / kg$, country: Finland incidence: 3/55, conc. range: 140-570 μ g / kg, Ø conc.: 350 μ g / kg, country: UK \rightarrow HT-2 toxin incidence: 24/208, conc. range: 60-590 μg/kg, country: Canada incidence: 10/53*, conc range: < 50 µg/kg, country: Canada, *suspected incidence: 2/27, conc. range: 8-40 µg/kg, \emptyset conc.: 24 µg/kg, country: Finland incidence: 2/87, conc. range: 50-60 μ g / kg, Ø conc.: 55 μ g / kg, country: Germany incidence: 1/80, conc.: 150 µg/kg, country: Germany incidence: 6/84, conc. range: 3-20 µg/kg, \emptyset conc.: 10 μ g / kg, country: Germany incidence: 1/2, conc.: 200 µg/kg, country: Hungary

→ moniliformin incidence: 6*/6, conc. range: 500-17,100 μ g / kg, Ø conc.: 8660 μ g / kg, country: Poland, *hand-selected, visible fungal growth \rightarrow neosolaniol incidence: 1/nc, conc.: 200 µg/kg, country: USSR \rightarrow nivalenol incidence: 3/4, Ø conc.: 25 µg/kg, country: Austria incidence: 1/2, conc.: $32 \mu g / kg$, country: Bulgaria incidence: 1/208, conc.: 60 μg/kg, country: Canada incidence: 4/10, conc. range: 4-40 µg/kg, Ø conc.: 23 µg/kg, country: Canada incidence: 1/5, conc.: 6644 µg/kg, country: China incidence: 3/4, Ø conc.: 162 μg/kg, country: China incidence: 8/10, conc. range: 8-373 μ g/kg, Ø conc.: 118 μ g/kg, country: China incidence: 3/27, conc. range: < 1000 μg/kg, country: Finland incidence: 2/2, Ø conc.: $42 \mu g / kg$, country: France incidence: 2/2, Ø conc.: 274 µg/kg, country: Germany incidence: 3/44, conc. range: 10-50 μ g / kg, Ø conc.: 30 μ g / kg, country: Germany incidence: 2/8, \emptyset conc.: 270 µg/kg, country: Germany incidence: 16/29, conc. range: 10-120 μg/kg, country: Germany incidence: 22/84, conc. range: 3-32 μ g / kg, Ø conc: 9 μ g / kg, country: Germany incidence: 1/1, conc.: 2 μ g/kg, country: Greece incidence: 1/2, conc.: $4 \mu g / kg$, country: Hungary incidence: 95/101, conc. range: 3-7300 μ g / kg, Ø conc.: 942 μ g / kg, country: Japan

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Wheat

incidence: 4/17, conc. range: 20-580 μg/kg, country: Japan incidence: 6/6, \emptyset conc.: 391 µg/kg, country: Japan incidence: 1/1, conc.: 160 μ g/kg, country: Japan incidence: 11/18, conc. range: ND-1000 μ g / kg, Ø conc.: 400 μ g / kg, country: Japan incidence: 8/11, conc. range: ND-3580 μ g / kg, Ø conc.: 450 μ g / kg, country: Japan incidence: 2/3, conc. range: ND-20 μ g / kg, Ø conc.: 10 μ g / kg, country: Japan incidence: 7/18*, conc range: 47-435 μ g/kg, Ø conc.: 205 μ g/kg, country: Japan, *scabby wheat incidence: 9/9, conc. range: \leq 3200 μ g / kg, Ø conc.: 534 μ g / kg, country: Korea incidence: 9/10, Ø conc.: 135 µg/kg, country: Korea incidence: 5/10, Ø conc.: 70 μg/kg, country: Nepal incidence: 81/90, conc. range: \leq 1270 μg/kg, country: New Zealand incidence: 37/48, conc. range: ≤ 350 μ g/kg, Ø conc.: 48 μ g/kg, country: Poland incidence: $1/3^*$, conc.: 10 μ g/kg, country: Poland, *healthy and damaged kernels incidence: 6/12, conc. range: 26-169 μ g / kg, Ø conc.: 74 μ g / kg, country: Taiwan incidence: 10/22, conc. range: \leq 170 μ g / kg, Ø conc.: 54 μ g / kg, country: Taiwan incidence: 4/10, conc. range: 5-52 µg/kg, \emptyset conc.: 22 µg/kg, country: Taiwan incidence: 12/13, conc. range: 7-203 μ g/kg, Ø conc.: 38 μ g/kg, country: The Netherlands incidence: 17/31, conc. range: 4-670 μ g/kg, Ø conc.: 101 μ g/kg, country: UK \rightarrow ochratoxin A incidence: 1/61, conc.: 160 µg/kg, country: Austria

incidence: 4/41, conc. range: 5-100 μg/kg, country: Austria incidence: 14/18*, conc. range: 30-27,000 μg/kg, country: Canada, *heated incidence: 4/4*, conc. range: 20-100 μ g / kg, country: Canada, *visible moldy incidence: 119/402*, conc. range: 0.05-4.9 μ g / kg (110 samples), 5-25 μ g / kg (6 sa), > 25- \leq 51 µg/kg (3 sa)*, Ø conc.: 0.7 μg/kg, country: Denmark, *conventional incidence: 29/73*, conc. range: 0.05-4.9 μ g / kg (25 samples), 5-25 μ g / kg (3 sa), $> 25- \le 36 \ \mu g / kg \ (1 \ sa), \ \emptyset \ conc.: 1.2$ μg/kg, country: Denmark, *ecological incidence: 17/45*, conc. range: 0.05-4.9 μ g / kg (16 samples), 5-25 μ g / kg (1 sa), \emptyset conc.: 0.9 μ g/kg, country: Denmark, *conventional, imported incidence: 72/194, conc. range: 0.8-37 μg/kg, country: Denmark incidence: 17/36*, conc. range: 1.2-21 μg/kg, country: Denmark, *ecological incidence: 1/3, conc.: 10 µg/kg, country: Egypt incidence: 3/97, conc. range: \leq 4.9-24.9 μg/kg, country: Germany incidence: 1/64, conc.: 0.4 µg/kg, country: Germany incidence: 8/64, conc. range: 0.1-137.3 μ g/kg, Ø conc.: 17.9 μ g/kg, country: Germany incidence: 94/719, conc. range: 0.1-12.5 μg/kg, country: Germany incidence: 3/97, conc. range: 0.4-15.4 μg/kg, country: Germany incidence: 1/30, conc.: nc, country: India incidence: 10/10*, conc.: \leq 2.6 µg/kg, \emptyset conc.: 1.4 µg/kg, country: Italy, *soft wheat incidence: 2/34, conc. range: 188-430 μ g / kg, Ø conc.: 309 μ g / kg, country: Italy incidence: 2/66, conc. range: 160-1000 μ g / kg, Ø conc.: 580 μ g / kg, country: Poland incidence: 28/239, conc. range: 5-2400 μg/kg, country: Poland

incidence: 1/209, conc.: 1.8 µg/kg, country: Saudi Arabia incidence: 5/5, conc. range: $\leq 0.8 \,\mu\text{g}/\text{kg}$, country: Spain incidence: 2/24, conc. range: $\leq 0.6 \ \mu g/$ kg, country: Spain incidence: 6/35, conc. range: \leq 4.9-8.6 μg/kg, country: Sweden incidence: 7/27, conc. range: $\leq 4.1 \ \mu g/$ kg, country: The Netherlands incidence: 6/38*, conc. range: 0.1-4.2 μ g / kg, country: The Netherlands, *imported incidence: 8/28, conc. range: 34-360 μg/kg, country: Tunesia incidence: 43/44*, conc. range: 0.1-11,064 μg/kg, country: Tunesia, *and derived food incidence: 2/8, conc. range: $\leq 2 \mu g / kg$, country: UK incidence: 2/129, conc. range: \leq 15 µg/ kg, country: UK incidence: 22/250, conc. range: \leq 4.9-31.6 μ g / kg, country: UK incidence: 10/18, conc. range: \leq 4.9-12 μ g/kg, country: UK incidence: 10/30, conc. range: \leq 1.2 µg/ kg, country: UK incidence: 8/25, conc. range: \leq 4.9-13.9 μg/kg, country: UK incidence: 2/9, conc. range: \leq 0.2 µg/kg, country: UK incidence: 15/101, conc. range: < 25-2700 μ g / kg, country: UK incidence: 9/11*, conc. range: < 50-3500 μg/kg, country: UK, *moldy incidence: 11/577*, conc. range: 5-115 µg/kg, country: USA, *hard red winter incidence: 9/848, conc. range: 20-114 μg/kg, country: USA incidence: 56/383, conc. range: 0.03-15,410 μg / kg, Ø conc.: 2.04 μg / kg, country: USA incidence: 11/130*, conc. range: 14-135 μ g / kg, country: Yugoslavia, *area with endemic nephropathy

incidence: 3/40, conc. range: 12-55 μ g / kg, Ø conc.: 34.3 μ g / kg, country: Yugoslavia \rightarrow rubratoxin incidence: 1/30, conc.: 245 µg/kg, country: India \rightarrow sterigmatocystin incidence: 1/18, conc.: ca. 300 µg/kg, country: Canada incidence: 2/30, conc. range: 110-145 μ g / kg, Ø conc.: 128 μ g / kg, country: India incidence: 7/11*, conc. range: tr-400 μ g / kg, country: UK, *moldy \rightarrow tenuazonic acid incidence: $33/33^*$, conc. range: ≤ 220 μ g / kg, Ø conc.: 50.1 μ g / kg, country: Australia, *weather-damaged \rightarrow T-2 toxin incidence: 3/208, conc. range: ≤ 18 μ g / kg, country: Canada incidence: $11/53^*$, conc. range: ≤ 200 μg/kg, country: Canada, *suspected incidence: 2/24, conc. range: 3-8 µg/kg, country: Finland incidence: 1/100, conc.: nc, country: France incidence: 1/87, conc.: 100 µg/kg, country: Germany incidence: 4/21, conc. range: 23-45 μ g / kg, Ø conc.: 25 μ g / kg, country: Germany incidence: 4/81, conc. range: 200-500 μg/kg, country: Germany incidence: 22/84, conc. range: 3-249 μ g / kg, Ø conc.: 82 μ g / kg, country: Germany incidence: 2/2, conc. range: 200-1900 μg/kg, country: Hungary incidence: 3/7*, conc. range: 2000-4000 μg/kg, country: India, *moldy incidence: 8/57, conc. range: 13-63 μg/kg, country: Poland incidence: 1/nc, conc.: 500 µg/kg, country: USSR

Wheat

 \rightarrow viomellein incidence: 8/11*, conc. range: 300-1800 μ g / kg, Ø conc.: 962 μ g / kg, country: UK, *moldy vioxanthin incidence: 8/11*, conc. range: 200-1200 μ g / kg, Ø conc.: 443 μ g / kg, country: UK, *moldy \rightarrow xanthomegnin incidence: 8/11*, conc. range: 120-1100 μ g / kg, Ø conc.: 390 μ g / kg, country: UK, *moldy \rightarrow zearalenone incidence: 20/20, \emptyset conc.: 10 μ g/kg, country: Argentina incidence: 9/10, conc. range: 2-21 µg/kg, \emptyset conc.: 9 μ g / kg, country: Canada incidence: 4/4, Ø conc.: 78 μg/kg, country: China incidence: 5/10, conc. range: 5-25 μg/kg, Ø conc.: 15 μ g/kg, country: China incidence: 2/40, conc. range: 12-32 μ g / kg, Ø conc.: 22 μ g / kg, country: Finland incidence: 8*/51, conc. range: ≤ 18 μ g / kg, Ø conc.: 6 μ g / kg, country: Germany, *conventional incidence: 18*/50, conc. range: \leq 105 μ g / kg, Ø conc.: 24 μ g / kg, country: Germany, *ecological incidence: 1/6, conc.: 5 μ g/kg, country: Germany incidence: 2/2, Ø conc.: $2 \mu g / kg$, country: Germany incidence: 15/92, conc. range: 0.5-290 μ g / kg, Ø conc.: 30 μ g / kg, country: Germany incidence: 58/106, conc. range: \leq 1560 μ g / kg, Ø conc.: 80 μ g / kg, country: Germany incidence: 12/48, conc. range: 5-20 μ g / kg, Ø conc.: 10 μ g / kg, country: Germany incidence: 3/8, Ø conc.: 4 μ g/kg, country: Germany incidence: 19/159, conc. range: 10-2000 μg/kg, country: Germany

incidence: 1/2, conc.: 10 µg/kg, country: Germany incidence: 67/84, conc. range: 1-8036 μ g/kg, Ø conc.: 178 μ g/kg, country: Germany incidence: 1/12, conc.: $4 \mu g / kg$, country: Italy incidence: 1/6, conc.: $1 \mu g/kg$, country: Japan incidence: 18/18, conc. range: 8-706 μ g / kg, Ø conc.: 189 μ g / kg, country: Japan incidence: 2/10*, conc. range: 8-40 μ g / kg, Ø conc.: 5 μ g / kg, country: Korea, *polished incidence: 5/9, \emptyset conc.: 141 µg/kg, country: Korea incidence: 2/10, Ø conc.: 4 µg/kg, country: Nepal incidence: 48/151, conc. range: \leq 460 μg/kg, country: New Zealand incidence: 1/48, conc.: 76 µg/kg, country: Poland incidence: 2/3*, conc. range: 10-2000 μ g / kg, Ø conc.: 1005 μ g / kg, country: Poland, *healthy and damaged kernels incidence: 5/13*, conc. range: 25-600 μ g / kg, Ø conc.: 425 μ g / kg, country: Poland, *healthy and damaged kernels incidence: 2/4, Ø conc.: 16 μg/kg, country: Portugal incidence: 2/2, conc. range: 3-10 µg/kg, \emptyset conc.: 6.5 µg/kg, country: Scotland incidence: 9/12, conc. range: 4-32 µg/kg, \emptyset conc.: 16 µg/kg, country: Taiwan incidence: 7/13, conc. range: 2-174 μ g / kg, Ø conc.: 45 μ g / kg, country: The Netherlands incidence: 4/31, conc. range: 1-3 µg/kg, \emptyset conc.: 1 µg/kg, country: UK incidence: 5/106*, conc. range: 100-200 $\mu g / kg$ (2 samples), > 200 $\mu g / kg$ (3 sa), country: Uruguay, *and by-products incidence: 14/27, conc. range: 400-3700 μ g / kg, Ø conc.: 950 μ g / kg, country: USA incidence: 1/116, conc.: 5000 µg / kg, country: USA

incidence: 3/33*, conc. range: 35-115 μ g/kg, Ø conc.: 80 μ g/kg, country: USA, *scabby incidence: 18/112, conc. range: 400 µg/kg (1 sample), 400-900 µg/kg (2 sa), 1000-5000 μ g/kg (13 sa), > 5000 μ g/kg (2 sa), country: USA incidence: 4/7, conc. range: 2 µg/kg, country: Yemen α -zearalenol (\rightarrow zearalenol) incidence: 4/84, conc. range: 8-71 µg/kg, \emptyset conc.: 23 µg/kg, country: Germany β-zearalenol incidence: 1/1, conc.: $12 \mu g/kg$, country: Germany \rightarrow cereals

Wheat (coarse ground) may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 1/1*, conc.: 1820 µg/kg, country: Papua, New Guinea, *imported \rightarrow zearalenone incidence: 1/1*, conc.: 1040 µg/kg, country: Papua New Guinea, *imported

Wheat (intermediate products): \rightarrow aflatoxin B₁ incidence: 35/475, Ø conc.: 11.1 µg/kg, country: Croatia

Wheat bran may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 2/3, conc. range: 170-450 μ g/kg, Ø conc.: 310 μ g/kg, country: Austria incidence: 1/2*, conc.: 45 µg/kg, country: Papua New Guinea, *imported incidence: 14/27, Ø conc.: 3400 µg/kg, country: USA \rightarrow nivalenol incidence: 1/2*, conc.: 19 µg/kg, country: Papua New Guinea, *imported \rightarrow ochratoxin A incidence: 1/1, conc.: 3 µg/kg, country: China

incidence: 6/57, conc. range: 5-20 µg/kg, country: Denmark incidence: 39/57, conc. range: 0.5-12 µg/kg, country: Denmark incidence: 10/15, conc. range: 0.1-26 μg/kg, country: Denmark incidence: 74/120*, conc. range: 0.05- 4.9 μ g / kg (72 samples), 5-12 μ g / kg (2 sa), country: Denmark, *conventional incidence: 15/22*, conc. range: 0.05-2.6 μ g/kg, Ø conc.: 0.6 μ g/kg, country: Denmark, *ecological incidence: 1/41, conc.: 0.1 µg/kg, country: Germany incidence: 3/5, conc. range: 0.2-0.8 $\mu g / kg$, \emptyset conc: 0.4 $\mu g / kg$, country: Switzerland incidence: 3/7, conc. range: $\leq 2.5 \ \mu g / kg$, country: The Netherlands \rightarrow zearalenone incidence: 14/27, Ø conc.: 2050 µg/kg, country: USA \rightarrow bran

Wheat flour During \rightarrow milling \rightarrow deoxynivalenol accumulated in the \rightarrow wheat bran whereas lower levels ($\approx 5\%$) were found in the break \rightarrow flour. Wheat flour may contain the following \rightarrow mycotoxins: \rightarrow acetyldeoxynivalenol incidence: 4/12*, conc. range: 600-2400 μg/kg, country: India, *moldy, refined \rightarrow aflatoxin B₁ incidence: 21/238, Ø conc.: 4.13 μg/kg, country: Croatia incidence: 1/83, conc.: 25.6 µg/kg, country: Malaysia \rightarrow aflatoxin B₂ incidence: 4/83, conc. range: 11.3-253 μ g/kg, Ø conc.: 75.2 μ g/kg, country: Malaysia \rightarrow aflatoxin G₁ incidence: 3/83, conc. range: 25-289 μ g/kg, Ø conc.: 135 μ g/kg, country: Malaysia

Wheat flour

 \rightarrow aflatoxin G₂ incidence: 11/83, conc. range: 16.3-436 μ g/kg, Ø conc.: 153 μ g/kg, country: Malaysia \rightarrow deoxynivalenol incidence: 61/61, conc. range: 250-9000 μ g / kg, Ø conc.: 1309 μ g / kg, country: Argentina incidence: 54/54, Ø conc.: 1210 μg/kg, country: Argentina incidence: 6/6, conc. range: 400-800 μ g / kg, Ø conc.: 467 μ g / kg, country: Argentina incidence: 11/47, conc. range: 27-830 μ g / kg, Ø conc.: 229 μ g / kg, country: Austria incidence: 43 products analysed, \emptyset conc. 400 µg/kg, country: Canada incidence: 7/7, Ø conc.: 129 μg/kg, country: China incidence: 5/5, conc. range: 11-690 μ g / kg, Ø conc.: 180 μ g / kg, country: China incidence: 42/44, conc. range: \leq 580 μ g/kg, Ø conc.: 130 μ g/kg, country: Germany incidence: 4/4*, conc. range: 41-180 μ g / kg, Ø conc.: 102 μ g / kg, country: Germany, *organic produce incidence: 3/3, conc. range: 60-90 µg/kg, \emptyset conc.: 75 µg/kg, country: Germany incidence: 9/12*, conc. range: 430-4850 μg/kg, country: India, *moldy, refined incidence: 2/5*, conc. range: 346-8380 μg/kg, country: India, *moldy, refined incidence: 26/36, conc. range: 2-239 μg/kg, country: Japan incidence: 1/1*, conc.: 1720 µg/kg, country: Papua New Guinea, *imported, fineground biscuit flour incidence: 1/1*, conc.: 2270 µg/kg, country: Papua New Guinea, *imported, raw flour incidence: 44/50, conc. range: ND-460 μg/kg, country: USA incidence: 2/27, conc. range: ND-2000, \emptyset conc.: 1500 µg/kg, country: USA

ergometrine (\rightarrow ergot alkaloids) incidence: 4/4, conc. range: 0.3-0.7 μg/kg, country: Canada ergosine incidence: 4/4, conc. range: 0.4-0.7 μg/kg, country: Canada ergotamine incidence: 4/4, conc. range: 0.3-2.3 μg/kg, country: Canada ergocornine incidence: 4/4, conc. range: 0.7-1.3 μg/kg, country: Canada α-ergokryptine incidence: 4/4, conc. range: 0-1.1 µg/kg, country: Canada ergocristine incidence: 4/4, conc. range: 0.4-4 µg/kg, country: Canada \rightarrow nivalenol incidence: 2/12*, conc. range: 30-100 μg/kg, country: India, *moldy, refined incidence: 12/36, conc. range: 4-84 μg/kg, country: Japan incidence: 1/1*, conc.: 310 µg/kg, country: Papua New Guinea, *imported, fingeground biscuit flour \rightarrow ochratoxin A incidence: 3/23*, conc. range: 0.2-0.5 μg/kg, country: Germany, *whole meal incidence: 12/13, conc. range: 0.1-1.9 μ g/kg, Ø conc.: 0.49 μ g/kg, country: Switzerland \rightarrow T-2 toxin incidence: 2/12*, conc. range: 550-800 µg/kg, country: India, *moldy, refined \rightarrow zearalenone incidence: 5/7, \emptyset conc.: 4 µg/kg, country: China incidence: 2/5, conc. range: 2-3 µg/kg, Ø conc.: 2.5 μ g/kg, country: China incidence: 3/4*, conc. range: 5.1-10 μ g / kg, Ø conc.: 6.9 μ g / kg, country: Germany, *organic produce incidence: 2/3, conc. range: 11-12 µg/kg, \emptyset conc.: 11.5 µg/kg, country: Germany incidence: 3/27, conc. range: 1-6 µg/kg, country: Japan

Wine

incidence: 1/1*, conc.: 250 μ g/kg, country: Papua New Guinea, *imported, raw flour incidence: 2/27, Ø conc.: 100 μ g/kg, country: USA \rightarrow flour, \rightarrow milling

Wheat grits may contain the following \rightarrow mycotoxins: \rightarrow citrinin incidence: 2/4*, conc. range: 0.3-0.7 µg / kg, Ø conc.: 0.5 µg / kg, country: Switzerland, *durum wheat \rightarrow deoxynivalenol incidence: 1/1, conc.: 160 µg / kg, country: Germany \rightarrow ochratoxin A incidence: 4/4*, conc. range: 0.8-2.7 µg / kg, Ø conc.: 1.65 µg / kg, country: Switzerland, *durum wheat \rightarrow barley grits, \rightarrow maize grits, \rightarrow rye grits

Wheat products may contain the following \rightarrow mycotoxins: \rightarrow deoxynivalenol incidence: 545/1257*, conc. range: 9-4060 µg/kg, Ø conc.: 260 µg/kg, country: Canada, * \rightarrow flour, \rightarrow bran, \rightarrow bread, cookies, crackers, cakes, pasta, etc. \rightarrow ochratoxin A incidence: 10/10*, conc. range: 0.2-3.5 µg/kg, Ø conc.: 1.37 µg/kg, country: Switzerland, *durum \rightarrow figazzas, \rightarrow libritos

Whey powder A storage period of 40 days did not change the \rightarrow aflatoxin M₁ of lyophylized whey powder to any significant degree. Whey powder may contain the following \rightarrow mycotoxins: aflatoxin M₁ incidence: 28/74, conc. range: 0.5-6.5 μ g/kg, country: France incidence: 88/88, conc. range: < 0.1-0.6 μ g/kg, country: UK

White cheese \rightarrow cheese (white)

 \rightarrow Ochratoxin A seems to be the Wine most important mycotoxin in wine. Red wine and red \rightarrow grape juice originating from the more southern and warmer regions of Europe and northern Africa are primarily affected. This may be due to the enhanced growth of OTA-producing \rightarrow Aspergillus species over \rightarrow Penicillium spp. and / or different practices in grape cultivation (e.g. pesticides, cultivars) and wine making (e.g. period and storage condition of the harvested grapes, maceration type, kind of fermentation). In addition, growth of OTA-producing molds in barrels and / or tanks or any other equipment as well as the failure to remove moldy \rightarrow fruits before processing might be responsible for the higher incidence and concentration of OTA in these wines. Since the climatic conditions are warm but less humid than in central Europe, it was suggested that OTA contamination of the grapes mainly occurs after harvest. Lower levels (\leq 0.005 µg OTA / l) have been found in red wines originating from the more central parts of Europe (Switzerland, Burgundy, Germany). It is assumed that OTA is probably formed prior to alcoholic fermentation since ethanol and the generally anaerobic conditions inhibit mold growth. In addition, red grape juices as well as the red wines contained similar concentrations.

No significant degradation occurs during wine making and storage. White wines contained less OTA than

rosé and these less than red wines. Besides OTA a contamination of red wines with \rightarrow ochratoxin C (ethyl ester of OTA) has been reported. This ochratoxin might be of fungal origin or an artifact. Wine may contain the following \rightarrow mycotoxins:

 \rightarrow aflatoxins (no specification)

incidence: 2/33, conc. range: $< 1 \mu g/l$, country: Germany \rightarrow ochratoxin A incidence: 14/41*, conc. range: \leq 1.2 μg/l, country: Germany, *white, partly imported incidence: 6/14*, conc. range: $\leq 2.4 \, \mu g / l$, country: Germany, *rosé, partly imported incidence: 40/89*, conc. range: \leq 7 µg/l, country: Germany, *red, partly imported incidence: 22/24*, conc. range: < 0.005-0.178 μ g/l, Ø conc.: 0.011 μ g/l, country: Switzerland, *white table wine, partly imported incidence: 77/79*, conc. range: < 0.005- $0.388 \ \mu g/l, \emptyset$ conc.: 0.039 $\mu g/l$, country: Switzerland, *red table wine, partly imported incidence: 13/15*, conc. range: < 0.005-0.123 μ g/l, Ø conc.: 0.011 μ g/l, country: Switzerland, *rosé table wine, imported incidence: $2/3^*$, conc. range: < 0.049-0.451 μg/l, Ø conc.: 0.290 μg/l, country: Switzerland, *Malaga, imported incidence: 2/2*, conc. range: < 0.044- $0.337 \ \mu g/l$, Ø conc.: 0.191 $\mu g/l$, country: Switzerland, *Marsala, imported incidence: nc/6*, conc. range: ≤ 0.17 $\mu g/l, \emptyset$ conc.: 0.011 $\mu g/l$, country: Switzerland, *Port wine, imported

incidence: 2/2*, conc. range: < 0.029-0.054 μ g/l, Ø conc.: 0.041 μ g/l, country: Switzerland, *Sherry, imported incidence: 2/2*, Ø conc.: 0.003 μ g/l, country: Switzerland, *Vermouth, imported

Wort In an experimental study an 8-day fermentation (*Saccharomyces cerevisiae*) of wort containing \rightarrow ochratoxin A, \rightarrow fumonisin B₁ and \rightarrow fumonisin B₂ at 25 °C caused mycotoxin losses in the range of 2-13%, 3-28% and 9-17%, respectively. While some OTA was taken up by the yeast (\leq 21%) almost no uptake occurred in the case of the \rightarrow fumonisins (FB₁ < 1%, FB₂ < 2%). No decrease in mycotoxin (\rightarrow mycotoxins) concentration was observed if a yeast-free wort was used.

In a further study it could be shown that OTA does not survive the malting process. If OTA was added at the start of the mashing process, simulating the use of OTA contaminated adjuncts, the finished \rightarrow beer contained OTA in the range of 2-28%.

 \rightarrow beer

X

Xanthomegnin is a lactone (3,3',4,4'-tetrahydro-10,10'-dihydroxy-7,7'-dimethoxy-3,3'-dimethyl-[8,8'-bi-1H-naphtho[2,3-c]pyran]-1,1',6,6',9,9-'hexone) mycotoxin (\rightarrow mycotoxins) which was first isolated from *Trichophyton megninii* in 1963 (see Figure Xanthomegnin).

CHEMICAL DATA

Empirical formula: $C_{30}H_{22}O_{12}$; molecular weight: 574

FUNGAL SOURCES

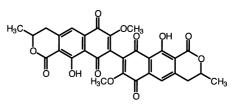
The penicillia are the main sources of xanthoquinones, e.g. \rightarrow Penicillium aurantiogriseum Dierckx, \rightarrow Penicillium crustosum Thom, *P. simplicissimum*, \rightarrow Penicillium verrucosum Dierckx, \rightarrow Penicillium viridicatum Westling, *Eupenicillium javanicum*, \rightarrow Aspergillus ochraceus group, *Trichophyton* spp.

NATURAL OCCURRENCE

 \rightarrow barley, \rightarrow oil seed rape, \rightarrow wheat Xanthomegnin may be found in ca. 50% of ochratoxin A suspected \rightarrow cereals and feed samples.

Тохісіту

hepatotoxic, nephrotoxic. strong uncoupler of oxidative phosphorylation (increased rate of respiration)



Xanthomegnin

DETECTION HPLC, TLC

Possible Mycotoxicosis

Xanthomegnin in combination with viomellein, ochratoxin A and citrinin may also be involved in kidney diseases of human and animals. Fungi producing these nephrotoxins are often co-occurring.

Further Comments

Xanthomegnin is often associated with \rightarrow viomellein. A simultaneous occurrence with \rightarrow ochratoxin A and \rightarrow citrinin is possible.

Y

Yeasts (fermentative)

Speed of fermentation (*Saccharomyces cerevisiae*) is depressed by \rightarrow T-2 toxin, \rightarrow diacetoxyscirpenol, aflatoxin (\rightarrow aflatoxins) (in decreasing order). A similar effect has been observed with \rightarrow patulin. T-2 toxin also inhibits yeast growth. However, a substantial reduction in patulin concentration (< 1% of the original levels) occurred during fermentation of \rightarrow apple juice.

Yellow rice disease is an intoxication which mainly occurred in the 19th and early 20th centuries as well as shortly after World War II in Japan. Many human deaths have been reported due to the consumption of moldy (yellow) \rightarrow rice imported form south-east Asia which had been declared unfit for human consumption. The syndrome involved \rightarrow acute cardiac beriberi. Moldy rice toxins should mainly be responsible for emesis, ascending \rightarrow paralysis, \rightarrow convulsions and respiratory arrest. Death may occur. More than 15 kinds of molds have been incriminated in the yellow rice syndrome but \rightarrow Penicillium islandicum Sopp $(\rightarrow \text{ islanditoxin}, \rightarrow \text{ luteoskyrin}), \rightarrow \text{Penicil-}$ lium citrinum Thom (\rightarrow citrinin), \rightarrow Penicillium citreonigrum Dierckx (synonym P. *citreo-viride*) (\rightarrow citreoviridin), and *P*. rugulosum (\rightarrow rugulosin) are the most important. Their \rightarrow mycotoxins primarily act on the liver but other organs such as the kidneys may also be affected.

Yogurt Although the contamination rate of yogurt with \rightarrow aflatoxin M₁ due to the \rightarrow carry over of \rightarrow aflatoxin B₁ from the feed into the \rightarrow milk (AFM₁) seems to be low, this aflatoxin is the most important mycotoxin in this kind of foodstuff. Different reports concerning the behavior and influence of AFM₁ and aflatoxin B₁ in / on yogurt do exist. The results are as follows: (i) no influence of yogurt manufacture and refrigerated storage on AFM₁ content, (ii) variable increases of AFM₁ content in yogurt, (iii) a high reduction of AFM₁ in yogurt, (iiii) complete transformation of AFB₁ in its hydroxy derivative AFB_{2a}. AFB₁ caused a delay in curdling.

In addition, AFM_1 caused thickening of the cell walls of *Lactobacillus bulgaricus* and *Streptococcus thermophilus*. A change in cell shape from coccoid to oval (*S. thremophilus*) and shortening of cell chain length (*L. bulgaris*) was also observed.

Yogurt may contain the following \rightarrow mycotoxins:

aflatoxin M₁

incidence: 44/54, conc. range: 0.05-0.47 μ g/kg, Ø conc.: 0.2 μ g/kg, country: Germany

incidence: 91/114, conc. range: < 0.001-0.496 μ g/kg, Ø conc.: 0.018 μ g/kg, country: Italy

incidence: $1/1^*$, conc.: $0.19 \ \mu g/kg$, country: Syria, *Koshk (sundried mixture of parboiled \rightarrow wheat and yogurt) milk

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Z

Zearalenol (Syn.: α -zearalenol) is a hydroxylated derivative of \rightarrow zearalenone due to zearalenone reductases present in animal tissues. Formation by F. semitectum has been reported. It is used as a growth promoter in livestock due to its anabolic potential. Apparently no residues accumulate in animal tissues and it does not exert potent uterotropic effects. α -Zearalenol possess a ten-times higher estrogenic activity than zearalenone whereas the β -isomer is considerably less active (similar or slightly less than that of zearalenone). Zearalenol may be of concern to food hygienists if it is transmitted into \rightarrow milk and other edible tissues.

Zearalenone (Syn.: F-2 toxin) is a 6-(10hydroxy-6-oxo-*trans*-1-undedenyl)- β resorcyclic acid lactone which is produced by \rightarrow Fusarium spp., primarily \rightarrow Fusarium graminearum Schwabe and \rightarrow Fusarium culmorum (W. G. Smith) Sacc. (see Figure Zearalenone). Originally (1962) this mycotoxin which was recovered from cultures of *Giberella zea* (sexual stage of *Fusarium roseum*) was called F-2 toxin. Determination of molecular structure followed in 1966.

CHEMICAL DATA

Empirical formula: $C_{18}H_{22}O_5$, molecular weight: 318

FUNGAL SOURCES

Fusarium spp.: e.g. \rightarrow Fusarium avenaceum (Fr.) Sac. (?), *F. culmorum*, \rightarrow Fusarium equiseti (Corda) Sacc. sensu Gordon, *F. graminearum*, \rightarrow Fusarium moniliforme Sheldon, \rightarrow Fusarium oxysporum Schlecht. emend. Snyd. & Hans., \rightarrow Fusarium sambucinum Fuckel, *F. semitectum*, \rightarrow Fusarium sporotrichioides Sherb.

NATURAL OCCURRENCE

- \rightarrow bananas, \rightarrow barley, \rightarrow barley malt,
- \rightarrow beans, \rightarrow beer, \rightarrow beer, joala, \rightarrow beer,

Zearalenone

opaque maize, \rightarrow beer, pito, \rightarrow bread, \rightarrow breakfast cereals, cereals, \rightarrow chilli powder, \rightarrow chilli sauce, \rightarrow coriander, \rightarrow corn flakes, \rightarrow curry, \rightarrow curry paste, \rightarrow fennel, \rightarrow fermented products, \rightarrow foods, \rightarrow garlic pickle, \rightarrow grains, \rightarrow job's-tears, \rightarrow maize, \rightarrow maize flour, \rightarrow maize malt, \rightarrow maize meal, \rightarrow maize, brewers, \rightarrow millet, \rightarrow millet meal, \rightarrow muffin mix, \rightarrow oats, \rightarrow oil, \rightarrow oil seeds, \rightarrow pepper, \rightarrow pop corn, \rightarrow rice, \rightarrow rye, \rightarrow rye bran, \rightarrow rye flour, \rightarrow snack food, \rightarrow sorghum, \rightarrow spices, \rightarrow tapioca, \rightarrow walnuts, \rightarrow wheat Zearalenone is commonly found in food, mainly in \rightarrow cereals and \rightarrow cereal products of the temperate regions. Zearalenone is of worldwide importance since it occurs in maize in international trade.

High levels of zearalenone in cereals usually accumulate during storage of mature, *Fusarium* infected grains that have not sufficiently dried because of wet weather at harvest or in grains that were stored wet (e.g. maize: moisture content > 22%). Beside this, zearalenone production has been reported on grains in the field, during harvest, commercial grain processing, and / or subsequently during storage of any food- or feedstuff containing the grain.

According to the mean zearalenone levels naturally found in feed transmission of this mycotoxin into tissues and \rightarrow milk of ruminants generally does not pose a significant human health risk. A normal daily intake (cow) of 50-165 mg zearalenone from protein rations did not result in any detectable residues. Although experimental \rightarrow carry over of zearalenone residues into \rightarrow poultry products was shown, rates of carry over due to naturally contaminated feed may be neglected. Residues of zearalenone in \rightarrow meat, milk and eggs seem to be negligible.

M. Weidenbörner, *Encyclopedia of Food Mycotoxins* © Springer-Verlag Berlin Heidelberg 2001

Zearalenone

TOXICITY

not acutely toxic (20,000 mg/kg oral application did not cause deaths in mice and rats), hyperestrogenic in swine; \rightarrow cattle are less, \rightarrow poultry are minimal affected; weakly \rightarrow teratogenic (pigs), \rightarrow mutagenic (?), possibly carcinogenic (class 2B carcinogen, IARC)

 LD_{50} (po): > 4000 - > 10,000 mg/kg bw rat/ LD_{50} sodium chloride (po): 3750 mg/kg bw rat)

hyperestrogenic syndromes: e.g. uterine enlargement, swelling of the vulva (vulvovaginitis), mammary glands and nipples, prolapse of the vagina or rectum, prolonged or interrupted estrus, pseudopregnancy, infertility especially prepubertal gilts but other species like rats, mice or monkeys are also affected Transmission of zearalenone via sow's milk to piglets cause estrogenism in the young pig.

Since the very high LD_{50} of zearalenone it might better be called a non-steroidal fungal hormon (estrogen), rather than a direct mycotoxin. Besides estrogenic zearalenone also possesses anabolic activity.

DETECTION

ELISA, GC-MS, HPLC, LC-MS, TLC

Possible Mycotoxicosis

Although an estrogenic syndrome in humans could not be correlated with the consumption of foods containing zearalenone, this mycotoxin has been implicated in several incidents of precocious pubertal changes in children (\rightarrow premature thelarche).

FURTHER COMMENTS

This mycotoxin seems to be a suitable indicator for the presence of other

 \rightarrow Fusarium mycotoxins in cereals such as

 \rightarrow trichothecenes (e.g. deoxynivalenol, \rightarrow nivalenol).

Temperatures between 12 and 14 °C are required for significant zearalenone formation but production also occurs at temperatures below 10 °C and even below freezing.

Zearalenone often co-occurs with deoxynivalenol in grain worldwide. At low concentrations the effect of pure zearalenone is antagonized by the presence of pure deoxynivalenol whereas zearalenone slightly enhanced the effects of deoxynivalenol over a range of concentrations. One strain of *F. semitectum* not only produced zearalenone but also \rightarrow zearalenol and 8'-hydroxyzearalenone.

Acremonium species of New Zealand produce zearalenols. These fungi may be important in maize grown in subtropical countries.

Reduction / elimination: Cleaning

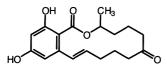
removed only 3-10% of zearalenone. Wet \rightarrow milling of maize led to an accumulation in the gluten (49-56%) > solubles > fiber > germ whereas zearalenone was not present in the starch fraction. The steeping procedure did not destroy zearalenone. During dry-milling, high levels were found in the maize germ, degermer fines, bran meal, hull, and high fat fractions. Low zearalenone levels (10-22%) occurred in the prime products (grits, low-fat meal, and flour).

Sieving of coarsely ground barley, wheat and maize caused substantial reductions in zearalenone (and \rightarrow deoxynivalenol) concentrations.

Zearalenone possess a relatively high heat stability - most survived a temperature of 180 °C for 30 min - and it is insensitive to hydrolytic cleavage.

Making \rightarrow bread caused losses in the range of 34-40% of the zearalenone originally present in \rightarrow wheat flour; instant \rightarrow noodles 48-62%, and \rightarrow biscuits 16-27%.

Zearalenone can survive the process of **brewing** corn, corn malt and other substrates whereas only little destruction of the mycotoxin occurred. The recovered solids contained about twice the levels of zearalenone originally present in maize.



Zearalenone

The stability of zearalenone during **fermentation** is further documented by its natural occurrence in maize \rightarrow beer. No zearalenone was found in ethanol resulting from the distillation of fermented

maize naturally contaminated with zearalenone. Saccharomyces cerevisiae converted zearalenone largely to β -zearalenol and, to a minor degree, to α -zearalenol.

Zwieback may contain the following

- \rightarrow mycotoxins:
- \rightarrow ochratoxin A
- incidence: 6/9, conc. range: 0.1-0.49
- $\mu g \, / \, kg$ (5 samples), 0.50-1.49 $\mu g \, / \, kg$
- (1 sa), country: Germany
- \rightarrow bread

Mycotoxin legislation. Maximum tolerated levels of mycotoxins in foodstuffs, dairy products and animal feedstuffs (FAO 1997, modified)

Country / food & feedstuff	Commodity	(Sum of) Mycotoxin(s)	Level µg / kg	Responsible authority	Remarks
Antiuga & Barbu	da: no regulations	1	1		
-	ion 1991, see also Mercosu	 r			·····
food	baby food	AFB ₁	0	1	
	groundnut, maize and by-products	$\begin{array}{c} AFB_1 \\ AFB_1, B_2, G_1, \end{array}$	5 20		
dairy	liquid milk, powdered milk	G ₂ AFM ₁	0.05		
	milk products	AFM ₁	0.5		
feed	soya meal	AFB ₁	30		
Australia, adopte	d by all states and territori	es			
food	all foods	AFB ₁ , B ₂ , G ₁ , G ₂ Phomopsin	5 5	Natl Food Auth	
	peanut butter, nuts and the nut proportion of products	AFB ₁ , B ₂ , G ₁ , G ₂	15	3)	
Austria (see Euro	pean Union):				
Food	all foods	$\begin{array}{c} AFB_1 \\ AFB_2, G_1, G_2 \end{array}$	1 5	Min Pub Health	
	milling and shelled prod- ucts and derived prod- ucts	AFB ₁ AFB ₂ , G ₁ , G ₂	2 5	33	
	children's foods (in pre- pared foods)	AFB ₁ , B ₂ , G ₁ , G ₂ , M ₁	0.02	"	
	wheat, rye	OTA DON ZEA	5 500 60		Guideline level
	durum wheat	OTA DON ZEA	5 750 60		
	fruit juice	Patulin	50	Min Pub Health	
dairy	milk(products)	AFM ₁	0.05	"	
	whey powder, whey paste	AFM ₁	0.4	"	Calcd on dry matter
	whey, liquid whey prod- ucts	AFM ₁	0.025	"	
	cheese	AFM ₁	0.25	»	
	butter	AFM ₁	0.02	"	
	pasteurized fresh milk for infants / children; chil- dren's food	AFM ₁	0.01	"	Calcd on recon stituted produc
	powdered milk(prod- ucts), condensed milk, milk concentrates	AFM ₁	0.4	"	Calcd on dry matter
feed	see European Union			······	

Country / food & feedstuff	Commodity	(Sum of) Mycotoxin(s)	Level µg / kg	Responsible authority	Remarks
Bahamas: situati	on 1991; no national regula	tions; FDA regu	lations ar	e used	
food	all foods, all grains	AFB ₁ , B ₂ , G ₁ , G ₂	20		
Bahrain: no regu	L lations	2			
Barbados: situati					GANGET -
Foods	all foods	AFB ₁ , B ₂ , G ₁ , G ₂	20		
Dairy	milk	AFM ₁	0.05		
Feed	all feedstuffs	AFB ₁ , B ₂ , G ₁ , G ₂	50		
Belgium (see Eur	opean Union):			•	
Food	peanuts	AFB ₁	5	Min Pub Health	
Dairy	milk	AFM ₁	0.05	"	
Feed	see European Union				
Belize:	J	•	•	•	• • • • • • • • • • • • • • • • • • • •
Food	maize, groundnut	AFB ₁ , B ₂ , G ₁ , G ₂	20		Situation 1991
Bolivia: situation	1991; no regulations		- L	L	
Bosnia and Herz	egowina: situation 1981			1	
Food	wheat, maize, rice, cereals	AFB ₁ , G ₁	1	Fed Comm La- bour Health Soc Welf	
	beans	AFB ₁ , G ₁	5	"	-
feed	feedstuffs	?	?		
Brazil: situation	1987; proposals; see also Me	ercosur	- I	L	
food	all foodstuffs	AFB ₁ AFB ₁ , B ₂ , G ₁ , G ₂	15 30		
	imported foodstuffs	AFB ₁ AFB ₁ , B ₂ , G ₁ , G ₂	5 10		
	industrially prepared foodstuffs for children from 0-2 years and for school meals	AFB ₁ , B ₂ , G ₁ , G ₂	3		
	rice, barley, beans, maize	ΟΤΑ	50		
	maize	ZEA	200		
	maize, groundnut	AFB ₁ , G ₁	30		Situation 1991
dairy	milk(products)	AFM ₁	0.5		Situation 1987 proposal
	imported milk(products)	AFM ₁	0.1		Situation 1987 proposal
feed	peanut meal (export)	AFB ₁ , B ₂ , G ₁ , G ₂	50		Situation 1977

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Country / food & feedstuff	Commodity	(Sum of) Mycotoxin(s)	Level µg / kg	Responsible authority	Remarks
Bulgaria: situatio	n 1992		10 0	· · · · · · · · · · · · · · · · · · ·	L
food	peanut(product)s, ker- nel(product)s, cocoa beans, cocoa butter, co- coa powder	AFB ₁ , B ₂ , G ₁ , G ₂	5		
	grain(products), cere- al(products)	AFB ₁ , B ₂ , G ₁ , G ₂	2.5		
dairy	liquid milk products powdered milk powdered milk for diete- tics and infant feeding cheese and similar pro-	AFM ₁ AFM ₁ AFM ₁ AFM ₁	0.5 0.1 0 0.5		
Canada:	ducts			l	
food	nut(product)s	AFB ₁ , B ₂ , G ₁ , G ₂	15	Health Can	Calcd on the nut meat portion
	uncleaned soft wheat	DON	2000	"	
feed	animal feeding stuffs	all aflatoxins	20	Agric Food Can- ada	
	diets for cattle/poultry	DON HT-2 toxin	5000 100		Recommenda- tion
	diets for swine/young calves/lactating dairy ani- mals	DON HT-2 toxin	1000 25		Recommenda- tion
	feedstuffs for reprodu- cing animals	all mycotoxins	0		Recommenda- tion
Chile: situation 19	991	•		·····	·
feed	feedstuffs feedstuffs	AFB ₁ AFB ₁ , B ₂ , G ₁ , G ₂ AFB ₁ AFB ₁ , B ₂ , G ₁ , G ₂	20 50 5 20		
China (People's R	epublic of China):	L	L	h	
food	rice, edible oils	AFB ₁	10	Min Health	
	maize, peanut(prod- uct)s, maize, peanut oil	AFB ₁	20	33	
	wheat, barley, oats, beans, sorghum, other grains, fermented food- stuffs	AFB ₁	5	"	
dairy	cow milk, milk products (calcd. on the basis of milk)	AFB ₁	0.5	"	
feed	compound feed for chick- ens	AFB ₁	10	St Tech Sup Bur	

Country / food & feedstuff	Commodity	(Sum of) Mycotoxin(s)	Level µg / kg	Responsible authority	Remarks
	compound feed for laying hens, compound feed and mixed feed for fattening pigs	AFB ₁	20	St Tech Sup Bur	
	maize, peanut cake, pea- nut residues	AFB ₁	50	St Tech Sup Bur	
Colombia: situati	on 1991	•	- L		
food	foods cereals (sorghum/millet)	$\begin{array}{c} \text{AFB}_1, \text{B}_2, \text{G}_1, \\ \text{G}_2 \\ \end{array}$	20		
		AFB ₁ , B ₂ , G ₁ , G ₂	30		
	oil seeds	AFB ₁ , B ₂ , G ₁ , G ₂	10		
feed	cattle feed	AFB ₁ , B ₂ , G ₁ , G ₂	50		
	sesame seeds	AFB ₁ , B ₂ , G ₁ , G ₂	20		
	poultry feedstuffs	$\begin{array}{c} \text{AFB}_1, \text{B}_2, \text{G}_1, \\ \text{G}_2 \end{array}$	20		
Costa Rica: situat	ion 1991	I	- J	L	L
Food	maize	AFB ₁ , B ₂ , G ₁ , G ₂	35		
Feed	maize	AFB ₁ , B ₂ , G ₁ , G ₂	50		
Côte d'Ivoire: situ	ation 1987; proposals, type	es of aflatoxins n	ot precise	ely stated	4
feed	straight feedstuffs	AFB ₁ , B ₂ , G ₁ , G ₂	100	Min Pub Health Min Animal Prod Min Commerce	
	complete feedstuffs	AFB ₁ , B ₂ , G ₁ , G ₂	10	33	
	complete feedstuffs for pigs / poultry (except young animals / ducks)	AFB ₁ , B ₂ , G ₁ , G ₂	38	»	
	complete feedstuffs for cattle / sheep, goats	AFB ₁ , B ₂ , G ₁ , G ₂	75		
	complete feedstuffs for dairy cattle	AFB ₁ , B ₂ , G ₁ , G ₂	50		
Cuba: situation 19		L	L	L	L
food	foods	AFB ₁ , B ₂ , G ₁ , G ₂	5		
	cereals, groundnuts	AFB ₁ , B ₂ , G ₁ , G ₂	5		
feed	feedstuffs, raw materials for feedstuffs	AFB ₁ , B ₂ , G ₁ , G ₂	5		
	feedstuffs, raw materials for feedstuffs	AFB ₁ , B ₂ , G ₁ , G ₂	5		

Country / food &	Commodity	(Sum of)	Level	Responsible	Remarks
feedstuff	ļ	Mycotoxin(s)	µg / kg	authority	
Cyprus: situation				·····	
food	cereals, pulses, dried fruit, sesame and foods produced exclusively from these, caraway seed, poppy seed, seeds used in bakery products and con- fectionery	AFB ₁ , B ₂ , G ₁ , G ₂	5		
dairy	milk, dairy products	all mycotoxins	0.5		
Czech Republic:		L	L	L	
food	all foods	AFB ₁ AFB ₂ , G ₁ , G ₂	5 10	Min Health	
	infant's /children's foods	$\begin{array}{c} \text{AFB}_1 \\ \text{AFB}_2, \text{G}_1, \text{G}_2 \end{array}$	1 2	"	
	all foods	Patulin OTA	50 20	"	
	children's foods	Patulin OTA	30 5))	
	infant's foods	Patulin OTA	20 1	23	
dairy	all foods	AFM ₁	5		
	milk	AFM ₁	0.5	>>	
	infant's / children's foods	AFM ₁	1	»	
	infant's foods on milk ba- sis	AFM ₁ AFB ₁ AFB ₂ , G ₁ , G ₂	0.1 0.1 0.2		Calcd on recon- stituted product
Denmark (see Eu	ropean Union):		I	L	
food	peanut(product)s	AFB ₁ AFB ₁ , B ₂ , G ₁ , G ₂	2 4	"	
	brazil nuts	$ \begin{array}{c} AFB_1 \\ AFB_1, B_2, G_1, \\ G_2 \end{array} $	2 4	"	
	dried figs	AFB ₁ AFB ₁ , B ₂ , G ₁ , G ₂	2 4	»	
	pig kidney	ΟΤΑ	25	Dan Vet Serv	whole carcass condemned; vis- ibly damaged kidneys are ana- lyzed chemically
	pig kidney	ΟΤΑ	10		viscera con- demned; visibly damaged kid- neys are ana- lyzed chemically
	cereal(product)s	OTA	5		
feed	see European Union				1

Country / food & feedstuff	Commodity	(Sum of) Mycotoxin(s)	Level µg / kg	Responsible authority	Remarks
Dominican Repu	blic: situation 1991		.		i
food	maize(product)s, pea- nut, soya, tomato(prod- ucts)	AFB ₁ , G ₁	0		
	imported maize	AFB ₁ , B ₂ , G ₁ , G ₂	20		
Ecuador: situatio	n 1991; no regulations				
Egypt:					
food	peanut(product)s, oil seed(product)s, cere-	AFB ₁ , B ₂ , G ₁ , G ₂	10		
	al(product)s	AFB ₁	5		
	maize	$\begin{array}{c} AFB_1 \\ AFB_1, B_2, G_1, \\ G_2 \end{array}$	10 20		
	starch and its derivatives	$ \begin{array}{c} \text{AFB}_1\\ \text{AFB}_1, \text{B}_2, \text{G}_1,\\ \text{G}_2 \end{array} $	0 0		
dairy	milk, dairy products	AFG ₁ , G ₂ , M ₁ , M ₂ AFM ₁	0		
feed	animal and poultry fod- ders	$ \begin{array}{c} \text{AFB}_1 \\ \text{AFB}_1, \text{B}_2, \text{G}_1, \\ \text{G}_2 \end{array} $	10 20		

European Union: All European Union tolerances refer to a commodity content of 12%; United Kingdom has extra regulation for feedstuff ingredients.

 1^{st} January 1999: 2 µg / kg AFB₁ and 4 µg / kg sum of AFB₁, B₂, G₁ and G₂ for cereals, peanuts, nuts, dried fruits and their products intended for direct human consumption or use as an ingredient in foodstuff. 8 µg / kg AFB₁ and 15 µg / kg sum of AFB₁, B₂, G₁ and G₂ for peanuts and 5 µg / kg AFB₁ and 10 µg / kg sum of AFB₁, B₂, G₁ and G₂ for peanuts and 5 µg / kg AFB₁ and 10 µg / kg sum of AFB₁, B₂, G₁ and G₂ for peanuts and 5 µg / kg AFB₁ and 10 µg / kg sum of AFB₁, B₂, G₁ and G₂ for nuts and dried fruits to be subjected to sorting, or other physical treatment, before human consumption or use as an ingredient in foodstuffs. 0.05 µg / kg AFM₁ in milk(products).

feed	straight feedstuffs	AFB ₁	50	various	
	straight feedstuffs: pea- nut(products), co- pra(products), cotton seed(products), palm- nut(products), babas- su(products), maize(products)	AFB1	20	33	
	complete feedstuffs for pigs and poultry (except young animals)	AFB ₁	20	>>	
	complete feedstuffs for cattle / sheep / goats (ex- cept dairy cattle / calves / lambs)	AFB ₁	50	»	
	complete feedstuffs for dairy cattle	AFB ₁	5	"	
	complete feedstuffs for calves and lambs	AFB ₁	10	"	

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Country / food & feedstuff	Commodity	(Sum of) Mycotoxin(s)	Level µg / kg	Responsible authority	Remarks
	other complete feedstuffs	AFB ₁	10	"	
	complementary feed- stuffs for pigs and poultry (except young animals)	AFB ₁	30	"	
	complementary feed- stuffs for catle / sheep / goats (except dairy ani- mals / calves / lambs)	AFB ₁	50	"	
	other complementary feedstuffs	AFB ₁	5	"	
	raw materials: ground- nut(product)s, co- pra(products), palmnut(products), cot- ton seed(products), ba- bassu(products), maize(products)	AFB1	200	»	
Finland (see Euro	pean Union):		.L		
food	all foods	AFB ₁ , B ₂ , G ₁ , G ₂	5	Min Trade Ind Natl Brd Trade Cons Int	
	all foods	Patulin	50	Min Trade Ind Natl Brd Trade Cons Int	
feed	see European Union			Min Agr For	
France (see Europ	pean Union):				I
food	all foods	AFB ₁	10		
	peanuts, pistachio nuts, almonds, oil seeds, chil- dren foods	AFB ₁	1	Min Consump	Not intended for the production of vegetable oils
	wheat meal	AFB ₁	3	"	
	wheat bran	AFB ₁	10	"	
	vegetable oils, cereals, wheat meal (complete)	AFB ₁	5	»	
	apple juice (products)	Patulin	50	33	
	cereals, vegetable oils	ZEA	200	"	
	cereals	OTA	5	"	
dairy	milk, milk powder(calcd on reconstituted pro- duct)	AFM ₁	0.05	"	
	milk, milk powder(calcd on reconstituted pro- duct) for infants under 3 years	AFM ₁	0.03	"	
feed	see European Union				

Country / food & feedstuff	Commodity	(Sum of) Mycotoxin(s)	Level µg / kg	Responsible authority	Remarks
Germany (see Eu	ropean Union):		10.0	. ,	
food	all foods	AFB ₁ AFB ₁ , B ₂ , G ₁ , G ₂	2 4	Bundes Ges	
	enzyme(preparation)s intended for the produc- tion of foodstuffs	AFB ₁ , B ₂ , G ₁ , G ₂	0.05	»	
	foods for infants and young children	AFB ₁ , B ₂ , G ₁ , G ₂	0.05	"	
dairy	milk	AFM ₁	0.05	>>	
	foods for infants and young children	AFM ₁	0.01	"	
feed	see European Union			Min Agr For	
Greece (see Europ	pean Union):	-			
food	peanuts, hazelnuts, wal- nuts, cashewnuts, pista- chio nuts, almonds, pumpkin seeds, sunflow- er seeds, pine, seeds, apri- cot seeds	AFB ₁ AFB ₁ , B ₂ , G ₁ , G ₂	5 10	Min Agr	
	maize, dried figs, dried apricots, dried prunes, dates, raisins	AFB_1AFB_1, B_2, G_1,G_2	5 10	"	
	raw coffee beans apple juice, apple pro- ducts	OTA Patulin	20 50	»	
feed	see European Union				
Guatemala: situat	ion 1991				
Food	maize, kidney beans, rice, sorghum	AFB ₁ , B ₂ , G ₁ , G ₂	20		
	groundnuts, groundnut butter	AFB ₁ , B ₂ , G ₁ , G ₂	20		Guide value un- til regulation is approved
Feed	concentrate	AFB ₁ , B ₂ , G ₁ , G ₂	20		Guide value un- til regulation is approved
Honduras: situati	on 1991				
Food	all foods	AFB ₁ , B ₂ , G ₁ , G ₂	1		
	maize (grounded or whole grain)	AFB ₁	1		
	baby food	$\begin{array}{c} \text{AFB}_1, \text{B}_2, \text{G}_1, \\ \text{G}_2 \\ \end{array}$	0.01		
		AFM ₁	0.02		
Dairy	milk(products)	AFM ₁	0.05		
	cheeses	AFM ₁	0.25		

Country / food &	Commodity	(Sum of)	Level	Responsible	Remarks
feedstuff		Mycotoxin(s)	μg / kg	authority	
Hong Kong:					
Food	foods	$ \begin{array}{c} AFB_1, B_2, G_1, \\ G_2, M_1, M_2, P_1, \\ a flatoxicol \end{array} $	15	Dep Health	
	peanut(products)	AFB ₁ , B ₂ , G ₁ , G ₂ , M ₁ , M ₂ , P ₁ , aflatoxicol	20	»	
Hungary:		• •••••••	•		
Food	all foods	AFB ₁	5	Min Health	Situation 1987
	groundnut kernels	AFB ₁	30		Situation 1987
	preserved foods	all mycotoxins	0	Min Health	Situation 1992
1	groundnuts	AFB ₁ , B ₂ , G ₁ , G ₂	5		Situation 1992
India: situation 19	987		1	L	1
food	all foods	AFB ₁	30	Min Health Fam Welf Dept Health	
feed	peanut meal (export)	AFB ₁	120	Min Fd Cvl Supp Dept Civil Supp	
Indonesia:		•			•
food	peanuts, maize, herbs, seeds			Min Health	Proposal in preparation
feed	copra in cow / pig / duck / sheep feed	AFB ₁ , B ₂ , G ₁ , G ₂	1000	Dir Anim Husb	Proposal ultimo 1994; includes max %-ages of raw material in various feed- stuffs for all cow / pig / duck / sheep feedstuffs
	groundnut / sesame seed / rape seed meal	AFB ₁ , B ₂ , G ₁ , G ₂	200	»	Proposal ultimo 1994
	cassava in chicken feed	AFB ₁ , B ₂ , G ₁ , G ₂	120	"	Proposal ultimo 1994
	capok seed / coconut meal in chicken feed, co- conut meal in cow / pig / duck / sheep feed	AFB ₁ , B ₂ , G ₁ , G ₂	100	»	Proposal ultimo 1994
	sunflower seed meal in chicken feed	AFB ₁ , B ₂ , G ₁ , G ₂	90	"	Proposal ultimo 1994
	soya bean / capok seed / fish / meat / bone meal / rice / maize bran, leucae- na (?), maize / wheat pol- lar (?), and sorghum in cow / pig / duck / sheep feed, maize / meat / bone / cotton seed meal in chicken feed	AFB ₁ , B ₂ , G ₁ , G ₂	50	»	Proposal ultimo 1994

Country / food & feedstuff	Commodity	(Sum of) Mycotoxin(s)	Level µg / kg	Responsible authority	Remarks
	soya bean / leucaena (?) / fish / meat / bone meal, rice / maize bran, wheat pollar (?), sorghum, co- pra in chicken feed	AFB ₁ , B ₂ , G ₁ , G ₂	20	33	Proposal ultimo 1994
Iran: no regulatio	ons				
Iraq: no regulatio	ons				
Ireland (see Euro	pean Union):				
food	all foods	AFB ₁ AFB ₁ , B ₂ , G ₁ , G ₂	5 30		Situation 1987
feed	see European Union				
Israel:		•		•	• 10
food	nut(product)s, pea- nut(product)s, maize flour (products), fig(products)	AFB ₁ AFB ₁ , B ₂ , G ₁ , G ₂	5 15		Proposal
	apple juice	Patulin	50		
	cereal(product)s, pulse(product)s	ΟΤΑ	50		Proposal
dairy	milk, milk powder (calcd on the basis of milk)	AFM1	0,05		Proposal
feed	according to European Union				Situation 1987
	grain for feed	AFB ₁ OTA T-2 toxin DAS	20 300 100 1000		Situation 1991
Italy (see Europea	an Union):			L	
food	all foods	AFB ₁ AFB ₁ , B ₂ , G ₁ , G ₂	5 10	ISS	
	dried figs	AFB ₁ AFB ₁ , B ₂ , G ₁ , G ₂	5 10	Min Health	
	spices	$\begin{array}{c} AFB_1 \\ AFB_1, B_2, G_1, \\ G_2 \end{array}$	20 40	ISS	
feed	see European Union				
Jamaica: situatior	i 1991			••••	•
Food	food, grains	AFB ₁ , B ₂ , G ₁ , G ₂	20		
Japan:					
Food	all foods	AFB ₁	10	Min Health Welf	

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Country / food & feedstuff	Commodity	(Sum of) Mycotoxin(s)	Level µg / kg	Responsible authority	Remarks
Feed	peanut meal (import)	AFB1	1000	Min Agr Forest- Fish	Not more than 2% in feed for dairy cattle; not more than 4% in feed for chicken (over 4 weeks of age), swine (over 30 kg) and cattle (over 3 months of age, except dairy cat- tle); not for use in feed for other livestock
Jordan: situation	1981				
Food	almonds, cereals, maize, peanuts, pistachio nuts, pine nuts, rice	AFB ₁ AFB ₁ , B ₂ , G ₁ , G ₂	15 30	Min Health	
Feed	all feedstuffs	$\begin{array}{c} AFB_1 \\ AFB_1, B_2, G_1, \\ G_2 \end{array}$	15 30	33	
Kenya: situation	1981	I	- I		1
Food	peanut(product)s, vege- table oils	AFB ₁ , B ₂ , G ₁ , G ₂	20	Min Health	
Kuwait: no regula	ations	L			L
Luxembourg (see	European Union):				
Food	peanut(product)s	AFB ₁	5	Min Pub Health	Situation 1981
Feed	see European Union				
Macedonia: situa	tion 1981	L			1
Food	wheat, maize, rice, cereals	AFB ₁ , G ₁	1	Fed Comm La- bour Health Soc Welf	
	beans	AFB_1, G_1	5	»	
Feed	feedstuffs				
Malawi: situation					
food	peanuts (export)	AFB ₁	5		
Malaysia: situatio					
food	all foods	AFB ₁ , B ₂ , G ₁ , G ₂	35		
Mauritius: situati	on 1987	•			
Food	all foods	AFB ₁ AFB ₁ , B ₂ , G ₁ , G ₂ , M ₁ , M ₂	5 10		
	groundnuts	AFB ₁ AFB ₁ , B ₂ , G ₁ , G ₂	5 15		

Country / food & feedstuff	Commodity	(Sum of) Mycotoxin(s)	Level µg / kg	Responsible authority	Remarks
	ntina, Uruguay, Brazil, and ears, will overrule national		oosals for	common regula	ations, probably ef-
Food	maize kernels (whole, pieces, ground, peeled), maize flour / meal, pea- nuts (in shell, raw, roast- ed) peanut cream, peanut butter	AFB ₁ , B ₂ , G ₁ , G ₂	20		
Dairy	liquid milk	AFM ₁	0.5		
	milk powder	AFM ₁	5		
Mexico:					
food	flours	all aflatoxins	20		
feed	cereals for bovine and porcine fattening feed- stuffs	AFB ₁ , B ₂ , G ₁ , G ₂	200		Situation 1991; less than 10% of cereals in feed- stuffs
	tle / poultry	AFB ₁ , B ₂ , G ₁ , G ₂	0		Situation 1991
Morocco: current	ly no regulations; Codex A	limentarius is fo	llowed		
Netherland, The ((see European Union)				
food	all foods and food ingre- dients except groundnuts used for the preparation of peanut oil	AFB ₁	5	Min VWS	
	cereal(product)s, pulse(product)s, leg- ume(product)s	all mycotoxins	0	Min VWS C Board	
dairy	milk(products), milk powder (calcd on recon- stituted product)	AFM ₁	0.05	Min VWS	
	cheese	AFM ₁	0.2	»	
	butter	AFM ₁	0.02	"	
	infant foods on milk basis	AFM ₁	0.05	"	As a proportion of the milk basis in infant food
feed	see European Union				
New Zealand: situ	ation 1987				
Food	all foods	AFB ₁ , B ₂ , G ₁ , G ₂	5		
	peanut butter, shelled nuts, nut portion of pro- ducts containing nuts	AFB ₁ , B ₂ , G ₁ , G ₂	15		
-	on 1991: no regulations				
Nigeria: situation					
Food	all foods	AFB ₁	20	FDA	
	infant foods	AFB ₁	0	>>	

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Country / food feedstuff	1 & Commodity	(Sum of) Mycotoxin(s)	Level µg / kg	Responsible authority	Remarks
Dairy	fluid milk	AFM ₁	1	"	
Feed	feedstuffs	AFB ₁	50	"	
Norway: situat	tion 1987				
Food	all foodstuffs	$\begin{array}{c} \text{AFB}_1, \text{B}_2, \text{G}_1, \\ \text{G}_2 \end{array}$	5		
	brazil nuts, buckwheat	AFB ₁ , B ₂ , G ₁ , G ₂	5		
	apple juice (concentrat- ed)	Patulin	50		Calcd on recon- stituted product
Feed	mixed feedstuffs depen- ding on type of animal	AFB ₁	10-50	Min Agr	Groundnut meal and cottonseed meal are not al- lowed entry
Oman: situatio	on 1987				
Feed	complete feedstuffs	AFB ₁	10	Min Comm Ind	Maximum con- tent referred to a moisture con- tent of 12%
	complete feedstuffs for poultry	AFB ₁	20	»	Maximum con- tent refered to a moisture con- tent of 12%
Panama: situa	tion 1991: no regulations		1	I	L,
Peru: situation	1991; no national regulations	, Codex Alimen	tarius pro	posals used	
Food	all foodstuffs	AFB ₁ , B ₂ , G ₁ , G ₂	10		
Feed	all feedstuffs	AFB ₁ , B ₂ , G ₁ , G ₂	10		
	complementary pro- ducts for milk, animal products, feedstuffs	AFB ₁	10		
	cereals for porcine grow- ing feedstuffs	AFB ₁ , B ₂ , G ₁ , G ₂	100		Situation 1991
Philippines:		•		•	
Food	nut(products)	AFB ₁ , B ₂ , G ₁ , G ₂	20		
Feed	poultry feedstuffs	AFB ₁ , B ₂ , G ₁ , G ₂	20	Bur Anim Husb	
	livestock feedstuffs	AFB ₁ , B ₂ , G ₁ , G ₂	50	»	
Poland:					
Food	all foods	AFB ₁	0	Min Publ Health	
feed	feedstuffs, feedstuff in- gredients, complete feed- stuffs for cattle / sheep / goats	AFB ₁	50		
	complete feedstuffs for pigs / poultry / dairy cows	AFB ₁	20		

Country / food & feedstuff	Commodity	(Sum of) Mycotoxin(s)	Level µg / kg	Responsible authority	Remarks
Portugal: situatio	n 1987 (see European Unic	on):		•	
Food	all foodstuffs	AFB1	20	Min Pub Health Min Agr Min Commerce	Situation 1987
	peanuts	AFB ₁	25	"	Situation 1987
	infant foods	AFB ₁	5	"	Situation 1987
Feed	see European Union				
Qatar: no regulati	ions	•			•
Romania: situatio	on 1987				
Food	all foods	AFB ₁	0	Min Pub Health Min Agr	
	all foods	Patulin	30	"	
		OTA	5		
D :		ZEA	30))	
Dairy	milk, dairy products	AFM ₁	0	» »	
Feed	all feedstuffs	AFB ₁ , B ₂ , G ₁ , G ₂	50		
	all feedstuffs	Patulin OTA DON Stachyobotrio- toxin Chaetomin	30 5 5 0 0	»	
Russia:			-		
Food	animal fats	AFB ₁ AFM ₁	0 0.5	Min Health	
	bottled / canned / potted fruits and berries	Patulin	50	"	
	bottled / canned / potted vegetables	AFB ₁ Patulin	5 50	"	
	casein	AFB ₁ AFM ₁	0 5	»	
	cereals (wheat of hard and strong types), flour, wheat bran	AFB ₁ ZEA T-2 toxin DON	5 1000 100 1000	"	
	fruits, berries and vegeta- bles (bottled / canned / potted juices and puree), cacao, cacao powder, chocolate, coffee, eggs, dehydrated egg, meat and poultry (fresh / chilled / frozen / tinned / potted / bottled), sausage and cu- linary products from meat and poultry, sub- products of farming ani- mals and poultry, sweets	AFB1	5	"	

Country I food 9	Commoditu	(Sum of)	Level	Deenoneihle	Remarks
Country / food & feedstuff	Commodity	(Sum of) Mycotoxin(s)	μg / kg	Responsible authority	Kennarks
	leguminous, protein iso- lators and concentrators, vegetable oil	AFB ₁ ZEA	5 1000	"	
	nut(kernel)s	AFB ₁ ZEA	? 1000	"	
Dairy	milk, sour dairy pro- ducts, concentrated milk, cheese and cottage cheese products, cow butter	AFB ₁ AFM ₁	0 0.5	33	
Salvador, El: situa	ation 1991	L		L	1
Food	foods	AFB ₁ , B ₂ , G ₁ , G ₂	20		
Feed	all feedstuffs	AFB ₁	10		
	supplementary feeds for porcine / poultry / dairy cattle; single composite feedstuffs; bovine / caprine / ovine feedstuffs	AFB ₁	20		
Saudi Arabia: no	regulations	•			
Senegal: situation	1987				
Feed	peanut products (straight feedstuffs)	AFB ₁	50	Min Commerce Min Pub Health	
	peanut products (feed- stuff ingredients)	AFB ₁	300	"	
Serbia: situation 1	981	L	- J		
Food	wheat, maize, rice, cereals	AFB ₁ , G ₁	1	Fed Comm La- bour Health Soc Welf	
	beans	AFB_1, G_1	5	"	
feed	feedstuffs	?	?		
Singapore: situati	on 1987		-		
Food	all foods	$\begin{array}{c} AFB_1 \\ AFB_1, B_2, G_1, \\ G_2 \end{array}$	0 0	Min Env	
South Africa:					
Food	all foods	AFB ₁ AFB ₁ , B ₂ , G ₁ , G ₂	5 10	Dept Health	
Spain (see Europe	ean Union):	•			• • • • • • • • • • • • • • • • • • •
Food	all foods	AFB ₁ AFB ₁ , B ₂ , G ₁ , G ₂	5 10	Min Pub Health Cons	
Feed	see European Union				
Sri Lanka:					
Food	foods	all aflatoxins	30		

Country / food & feedstuff	Commodity	(Sum of) Mycotoxin(s)	Level µg / kg	Responsible authority	Remarks
	foods intended for chil- dren up to 3 years of age	all aflatoxins	1		
Dairy	milk(products)	all aflatoxins	1		
Suriname: situati	on 1991	- 1		·····	······································
Food	maize	AFB ₁ , B ₂ , G ₁ , G ₂	30		
	groundnut(products), legumes	AFB ₁	5		
Feed	feedstuffs	AFB ₁ , B ₂ , G ₁ , G ₂	30		
Sweden : replacem (see European Ur	ient of Swedish feestuff regu iion)	lations with EU	regulatio	ns to be reconside	red near 31-12-9
Food	all foods	AFB ₁ , B ₂ , G ₁ , G ₂	5	Natl Food Adm	
	berries, fruits, juices	Patulin	50	"	Situation 1987
Dairy	liquid milk products	AFM ₁	0.05	"	
Feed	feedstuff ingredients	AFB ₁	50	"	
	feedstuff ingredients for dairy cattle	AFB ₁	10	"	
	cereal grains and forages as feedstuff ingredients for dairy cattle	AFB ₁	1	33	
	mixed feedstuffs (exclu- ding forages) for dairy cattle	AFB ₁	3	»	
	complete feedstuffs	AFB ₁	10	"	
	complete feedstuffs for cattle / sheep / goats ex- cept dairy cattle / lambs / kids	AFB ₁	50	»	
	complete feedstuffs for pigs and poultry except young animals	AFB ₁	20	33	
	complete feedstuffs (in- cluding forages) for dairy cattle	AFB ₁	1.5	»	
	complete feedstuffs for poultry	ΟΤΑ	200	»	
	complete feedstuffs for pigs	ΟΤΑ	100	33	
Switzerland:		•			• • • • • • • • • • • • • • • • • • • •
Food	all foods (except maize / cereals / herbs)	$\begin{array}{c} \text{AFB}_1 \\ \text{AFB}_2, \text{G}_1, \text{G}_2 \end{array}$	1 5	Lab Cantons	
	maize cereals (granular or ground)	AFB ₁ AFB ₂ , G ₁ , G ₂	2 5	"	
	herbs	AFB ₁ AFB ₂ , G ₁ , G ₂	5 5	"	

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Country / food & feedstuff	Commodity	(Sum of) Mycotoxin(s)	Level µg / kg	Responsible authority	Remarks
	babie's / infant's food	AFB ₁ , B ₂ , G ₁ , G ₂	0,01	"	Calcd on recon stituted produc
	cereal(product)s	OTA	2	Bund Amt Ges	Provisional
	maize(products)	Fumonisin B ₁ +B ₂	1000))	Provisional
	fruit juice	Patulin	50	Lab Cantons	
Dairy	milk(products)	AFM ₁	0.05	"	
	whey(products)	AFM ₁	0.025	"	
	cheese	AFM ₁	0.25	>>	
	butter, baby / infant food	AFM ₁	0.02	"	
feed	prohibit feeding cattle with peanut bruise	\$?	For Viehw	
Taiwan, Province	of China: situation 1991	• • • • • • • • • • • • • • • • • • • •		•	<u> </u>
food	cereals	AFB ₁ , B ₂ , G ₁ , G ₂	50	Dept Health Council Agr	
feed	feed, oilseed meals for feed under 4 % of mixed feed	AFB ₁ , B ₂ , G ₁ , G ₂	1000		
Thailand: situatio	n 1987		····	.	· · · · · · · · · · · · · · · · · · ·
food	all foods	AFB ₁ , B ₂ , G ₁ , G ₂	20	Min Pub Health	
Trinidad & Tobag	o: situation; no national re	gulations, Code	x Alimen	tarius proposals u	sed
Food	foods	AFB ₁ , B ₂ , G ₁ , G ₂	10		
Feed	feedstuffs	AFB ₁ , B ₂ , G ₁ , G ₂	10		
	complementary products	AFB ₁	10		
	ice cream	all mycotoxins	0		Situation 1992
UAE (United Aral	Emirates): no regulations			·	• • • • • • • • • • • • • • • • • • • •
UK (United Kingd	lom) (see European Union):			
food	nut(product)s, dried fig (product)s	AFB ₁ , B ₂ , G ₁ , G ₂	4	Min Agr Fish Fd	
feed	see European Union				
	groundnut, copra, palm- kernel, cottonseed, ba- bassu, maize and derived products (raw materials)	AFB ₁	20	»	Levels refer to a moisture con- tent of 12%
Uruguay: see also					
Food	foods and spices	AFB ₁ , B ₂ , G ₁ , G ₂	20	Min Pub Health	
	texturized soy protein products: flour, starch, concentrate, isolate	AFB ₁ , B ₂ , G ₁ , G ₂	30	"	
	peanuts, dried fruit(product)s	AFB ₁ , B ₂ , G ₁ , G ₂	30	"	

Country / food & feedstuff	Commodity	(Sum of) Mycotoxin(s)	Level µg / kg	Responsible authority	Remarks
	cocoa beans	AFB ₁ , B ₂ , G ₁ , G ₂	10	"	
	infant foods, produced industrially	AFB ₁ , B ₂ , G ₁ , G ₂	3	"	
	rice, barley, beans, coffee, maize	ΟΤΑ	50	»	
	maize, barley	ZEA	200	"	
	fruit juice	Patulin	50	"	
dairy	milk(products)	AFM ₁	0.5	>>	
USA: United State	es of America				
food	all foods	AFB ₁ , B ₂ , G ₁ , G ₂	20	FDA	
	finished wheat products	DON	1000	»	
dairy	whole milk, low fat milk, skim milk	AFM ₁	0.5	"	
feed	feedstuff(ingredient)s	AFB ₁ , B ₂ , G ₁ , G ₂	20))	
	cottonseed meal intended for beef cattle / swine / poultry feedstuffs (re- gardless of age or bree- ding status)	AFB ₁ , B ₂ , G ₁ , G ₂	300	»	
	maize and peanut prod- cuts intended for bree- ding beef cattle / swine or mature poultry	AFB ₁ , B ₂ , G ₁ , G ₂	100	»	
	maize and peanut pro- ducts intended for finish- ing swine of 100 pounds or greater	AFB ₁ , B ₂ , G ₁ , G ₂	200	»	
	maize and peanut pro- ducts intended for finish- ing beefcattle	AFB ₁ , B ₂ , G ₁ , G ₂	300	"	
	grains and grain by-pro- ducts destined for rumi- nating beef and feedlot cattle older than 4 months and for chickens (not exceeding 50% of the cattle or chicken total diet)	DON	10,000	»	
	grains and grain by-pro- ducts (not exceeding 40% of the diet)	DON	5000	»	
	grains and grain by-pro- ducts destined for swine (not exceeding 20% of their diet)	DON	5000	"	

(continued)

Country / food & feedstuff	Commodity	(Sum of) Mycotoxin(s)	Level µg / kg	Responsible authority	Remarks
Venezuela: situat	ion 1991			L	
Food	rice flour	AFB ₁ , B ₂ , G ₁ , G ₂	5		
Feed	feedstuffs	AFB ₁ , B ₂ , G ₁ , G ₂	20		
Zimbabwe:	L			· · · · · · · · · · · · · · · · · · ·	
Food	foods	AFB ₁ AFG ₁	5 4	Min Agr	
	groundnuts, maize, sor- ghum	AFB ₁ AFG ₁	5 4	"	
dairy	feedstuffs	AFB ₁ , B ₂ , G ₁ , G ₂		"	Levels vary with type of animal
	poultry feed	AFB_1, G_1	10 10	»	

 $AFB_1 = aflatoxin B_1$, DAS = diacetoxyscirpenol, DON = deoxynivalenol, OTA = ochratoxin A, ZEA = Zearalenone

Literature

Anonymous (1978) Interactions of Mycotoxins in Animal Production. Proceedings of a Symposium July 13, 1978 Michigan State University. National Academy of Science, Washington DC

Anonymous (1989) Mycotoxins: Economic and Health Risks. Council for Agricultural Science and Technology, Ames, Iowa

Applebaum RS, Brackett RE, Wiseman DW, Marth EH (1982) Aflatoxin: Toxicity to dairy cattle and occurrence in milk and milk products – a review. J Food Prot 45:752–777

- Arora DK, Mukerji KG, Marth EH (1991) (Eds) Handbook of Applied Mycology, Vol. 3, Foods and Feeds. Marcel Dekker, New York etc
- Battaglia R, Hatzold T, Kroes R (Eds) (1996) Occurrence and Significance of Ochratoxin A in Food. 10– 12 January 1996 Aix-en-Provence, France, ILSI, Europe. Food Add & Contam 13, Supplement:1–57

Betina V (Ed) (1984) Mycotoxins - Production, Isolation, Separation and Purification. Elsevier, Amsterdam etc

Betina V (Ed) (1989) Mycotoxins - Chemical, Biological and Environmental Aspects. Elsevier, Amsterdam etc

Beuchat LR (Ed) (1987) Food and Beverage Mycology. 2nd Ed, AVI, New York

Bhatnagar D, Lillehoj EB, Arora DK (eds) (1992) Handbook of Applied Mycology. Marcel Dekker, mc, New York

- Bullerman LB (1979) Significance and mycotoxins to food safety and human health. J Food Protect 42:65-86
- Champ BR, Highley E, Hocking AD, Pitt JI (1991) Fungi and Mycotoxins in Stored Products. Proceedings of an International Conference held at Bangkok, Thailand, 23–26 April 1991. ACIAR Proceedings No 36

Chelkowski J (Ed) (1989) Fusarium - Mycotoxins, Taxonomy and Pathogenicity. Elsevier, Amsterdam

Chelkowski J (Ed) (1991) Cereal Grain. Mycotoxins, Fungi and Quality in Drying and Storage. Elsevier, Amsterdam

Cole RJ, Cox RH (Eds) (1981) Handbook of Toxic Fungal Metabolites. Academic Press, New York etc

Creppy EE, Castegnaro M, Dirheimer G (Eds) (1993) Human Ochratoxicosis and its Pathologies. John Libbey Eurotext, Montrouge, France

- Douglas King A Jr, Schade JE (1984) Alternaria toxins and their importance in food. J Food Protect 47:886-901
- Doyle ME, Steinhart CE, Cochrane BA (1994) Food Safety 1994. Marcel Dekker, New York etc
- Eaton DL, Groopman JD (Eds) (1994) The Toxicology of Aflatoxins: Human Health, Veterinary, and Agricultural Significance. Academic Press Inc, San Diego
- Egmond HP van (Ed) (1989) Mycotoxins in Dairy Products. Elsevier Applied Science, London, New York
- Egmond HP van, Speijers GJA (1994) Survey of data of the incidence and levels of ochratoxin A in food and animal feed worldwide. Natural Toxins 3:125–144
- Ellis WO, Smith JP, Simpson BK, Oldham JH (1991) Aflatoxins in food: occurrence, biosynthesis, effects of organisms, detection, and methods of control. Crit Rev Food Sci Nutr 30:403-439

FAO (1997) Food and Nutrition Paper 64. Worldwide Regulations for Mycotoxins. A compendium. Rome

- Flannigan B (1991) Mycotoxins. In: D´Mello JP, Duffus CM, Duffus JH (Eds) Toxic Substances in Crop Plants, pp 226–257. Royal Society of Chemistry, Cambridge
- Forgacs J, Carll WT (1962) Mycotoxicoses. Adv Vet Sci 7:273-382

Literature

- Frank HK (1974) Aflatoxine. Bildungsbedingungen, Eigenschaften und Bedeutung für die Lebensmittelwirtschaft. BEHR'S Verlag, Hamburg
- Frank HK (1992) Citrinin. Z Ernährungswiss 31:164-177
- Frisvad J (1988) Fungal species and their specific production of mycotoxins. In: Samson RA, Reenen-Hoeckstra E.S (Eds) Introduction of Food-borne Fungi, pp 239-249. Centraalbureau voor Schimmelcultures, Baarn, Netherlands
- Frisvad J (1989) The connection between the penicillia and aspergilli and mycotoxins with special emphasis on misidentified isolates. Arch Environ Cont Toxicol 18:452–467
- Galvano F, Galofaro V, Galvano G (1996) Occurrence and stability of aflatoxin M₁ in milk and milk products: a worldwide review. J Food Protect 59:1079–1090
- Gedek B (1989) Mykotoxine. In: Gemeinhardt, H (Ed) Endomykosen. Gustav Fischer Verlag, Jena
- Gelda CS, Luyt LJ (1977) Survey of total aflatoxin content in peanuts, peanut butter, and other foodstuffs. Ann Nutr Alim 31:477-483
- Goldblatt LA (Ed) (1969) Aflatoxin. Scientific Background, Control, and Implications. Academic Press, New York London
- Hawksworth DL, Kirk PM, Sutton BC, Pegler DN (Eds) (1995) Ainsworth & Bisby's Dictionary of the Fungi. 8th Ed, CAB International, Wallingford
- Hesseltine CW (1974) Natural occurrence of mycotoxins in cereals. Mycopath Mycol Appl 53:141-153
- Höhler D (1998) Ochratoxin A in food and feed: occurrence, legislation and made at action. Z Ernährungswiss 37:2–12
- IARC (1993) IARC monographs on the evaluation of carcinogenic risk of chemicals to humans. Ochratoxin A. Nr. 56:489-521
- IARC (1993) IARC monographs on the evaluation of carcinogenic risk of chemicals to humans. Toxins derived from *Fusarium moniliforme*: fumonisins B₁ and B₂ and Fusarin C. Nr. 56:445–466

Jackson LS, de Vries JW, Bullerman LB (Eds) (1996) Fumonisins in Food. Plenum Press, New York

- Jarvis B (1976) Mycotoxins in food. In: Skinner, FA, Carr, JG (Eds) Symposium: Microbiology in Agriculture, Fisheries & Food, pp 251–267. Academic Press, London
- Jelinek CF, Pohland AE, Wood G (1989) Worldwide occurrence of mycotoxins in foods and feeds an update. J Assoc Off Anal Chem 72:223-230
- Kiermeier F (1973) Mykotoxine in Milch und Milchprodukten. Z Lebensm Unters Forsch 151:237-240
- Krogh P (1987) Mycotoxins in Food. Academic Press, London etc
- Kubacki SJ (1986) The analysis and occurrence of patulin in apple juice. In: Steyn PS, Vlegaar R (Eds) Mycotoxins and Phycotoxins. A collection of invited papers presented at the sixth International IU-PAC Symposium of Mycotoxins and Phycotoxins, Pretoria. Bioactive Molecules, pp 293–304. Elsevier, Amsterdam etc
- Kurata H, Ueno Y (1984) Toxigenic Fungi Their Toxins and Health Hazard. Developments in Food Science 7. Elsevier, Amsterdam etc
- Kühl H (1910) Über eine eigenartige Veränderung der Paranuss. Pharm Zentralhalle 106
- Lacey J (Ed) (1985) Trichothecenes and other Mycotoxins. John Wiley and Sons, New York
- Marasas WFO, Nelson PE (Eds) (1987) Mycotoxicology: Introduction to the Mycology, Plant Pathology, Chemistry, Toxicology, and Pathology of Naturally Occurring Mycotoxicoses in Animals and Man. The Pennsylvania State University Press, University Park, PA
- Marasas WFO, Nelson PE, Tousson TA (1984) Toxigenic *Fusarium* Species, Identity and Mycotoxicology. The Pennsylvania State University Press, University Park, PA
- Marasas WFO (1995) Fumonisins: their implications for human and animal health. Natural Toxins 3:193-198
- McKee LH (1995) Microbial contamination of spices and herbs: a review. Lebensm-Wiss Technol 28:1-11
- Miller JD (1995) Fungi and mycotoxin in grain: implications for stored product research. J Stored Prod Res 31:1-16
- Miller JD, Trenholm HL (1994) Mycotoxins in Grain. Compounds Other Than Aflatoxin. Eagan Press, St. Paul, Minnesota

Ministry of Agriculture, Fisheries and Food (1980) Survey of Mycotoxins in the United Kingdom. The Fourth Report of the Steering Group on Food Surveillance. The Working Party on Mycotoxins. Food Surveillance Paper No. 4. London, HMSO

Ministry of Agriculture, Fisheries and Food (1987). Survey of Mycotoxins in the United Kingdom. The Eighteenth Report of the Steering Group on Food Surveillance. The Working Party on Naturally Occurring Toxicants in Food: Sub-Group on Mycotoxins. Food Surveillance Paper No. 18. London, HMSO

Moreau C, Moss M (Eds) (1979) Moulds, Toxins and Food. John Wiley and Sons, New York

Mücke W, Lemmen Ch (1999) Schimmelpilze. Vorkommen, Gesundheitsgefahren, Schutzmaßnahmen. Ecomed, Landsberg/Lech

Natori S, Hashimoto K, Ueno Y (Eds) (1989) Mycotoxins and Phycotoxins. Elsevier, Amsterdam etc

Pestka JJ (1986) Fungi and Mycotoxins in Meats. In: Pearson AM, Dutson TR (Eds) Advances in Meat Research, pp 277–309. Mac Millan Publishers, Michigan

Pitt JI (1979) The Genus *Penicillium* and its Teleomorphic States *Eupenicillium* and *Talaromyces*. Academic Press, London

Pohland AE, Nesheim S, Friedman L (1992) Ochratoxin A: a review (Technical report). Pure & Appl Chem 64:1029–1046

- Pohland AE (1993) Mycotoxins in review. Food Add Cont 10:17-28
- Purchase IFH (1971) Mycotoxins in Human Health. Macmillan, London
- Purchase IFH (1974) (Ed) Mycotoxins. Elsevier, Amsterdam
- Purchase KA (1998) Mycotoxins. In: Watson D (Ed) Natural Toxins in Food, pp 147–181. Academic Press, London

Reiss J (Ed) (1981) Mykotoxine in Lebensmitteln. Gustav Fischer Verlag, Stuttgart, New York

Reiss J (1998) Schimmelpilze. Lebensweise, Nutzen, Schaden, Bekämpfung. 2. Aufl. Springer, Berlin Heidelberg New York

Rodricks JV (Ed) (1976) Mycotoxins and Other Fungal Related Food Problems. Advances in Chemistry Series 149. American Chemical Society, Washington, DC

Rodricks JV, Hesseltine CW, Mehlmann MA (Eds) (1977) Mycotoxins in Human and Animal Health. Pathotox Publishers Inc, Park Forest South, IL

Roth L, Frank H, Kromann K (1990) Giftpilze - Pilzgifte. Ecomed, Landsberg/Lech

Samson RA, Hoekstra ES, Frisvad JC, Filtenborg O (1998) Introduction to Food-borne Fungi. Centraalbureau voor Schimmelcultures, Baarn, Netherlands

Sauer DB (Ed) (1992) Storage of Cereal Grains and Their Products: 4th Ed. American Association of Cereal Chemists, St. Paul, Minnesota

Scott, PM (1990) Trichothecenes in grains. Cereal Foods World 35:661-666

Scott, PM (1997) Multi-year monitoring of Canadian grains and grain-based foods for trichothecenes and zearalenone. Food Add Cont 14:333-339

Sharma RP, Salunkhe DK (1991) Mycotoxins and Phytoalexins. CRC Press, Boca Raton, Florida

Shepard GS, Thiel PG, Stockenström S, Sydenham EW (1996). Worldwide survey of fumonisin contamination of corn and corn-based products. JAOAC 79:671-687

Smith JE, Moss MO (1985) Mycotoxins. Formation, Analysis and Significance. John Wiley and Sons, New York

Stoloff L. (1976): Incidence, distribution, and disposition of products containing aflatoxins. Proc Am Phytopathol Soc 3:156-172

Tanaka T, Hasegawa A, Yamamoto S, Lee U-S, Sugiura Y, Ueno Y (1988) Worldwide contamination of cereals by the *Fusarium* mycotoxins nivalenol, deoxynivalenol, and zearalenone. 1. Survey of 19 counries. J Agric Food Chem 36:979–983

Ueno Y (Ed) (1983) Trichothecenes – Chemical, Biolgical and Toxicological Aspects. Kodansha/Elsevier, Tokyo

Ueno Y (1985) The toxicology of mycotoxins. Crit Rev Toxicol 14:99-132

Watson DH (1985) Toxic Fungal Metabolites in Food. CRC Crit Rev Food Sci Nutr 22:177-198

Weidenbörner M (1998) Lebensmittel-Mykologie. BEHR'S Verlag, Hamburg

Weidenbörner M (1999) Lexikon der Lebensmittelmykologie. Springer-Verlag, Berlin etc

WHO (1979) Environmental Health Criteria 11. Mycotoxins. World Health Organization, Geneva

Literature

Wilson DM, Abramson D (1992) Mycotoxins. In: Sauer DB (Ed) Storage of Cereal Grains. 4th Ed. pp 341-391. Amercian Association of Cereal Chemists, St. Paul, Minnesota

Wyllie TD, Morehouse LG (1978) (Eds) Mycotoxic Fungi, Mycotoxins, Mycotoxicoses. An Encyclopedic Handbook. Vol. 3 Marcel Dekker Inc, New York

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